

Supporting Information

Modeling Bioaccumulation of Metallic Engineered Nanomaterials in a Freshwater Ecosystem

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Data selection tiers

Parameter selection for each species and each ENM was conducted in a tiered process where the preference ranking was as follows:

1. parameter measured for specific ENM and specific species
2. parameter measured for specific ENM for similar species
3. parameter measured for any ENM for specific species
4. parameter measured for any ENM for similar species
5. parameter measured for metal ion of ENM for specific species
6. parameter measured for metal ion of ENM for similar species
7. parameter measured for similar metal ion for specific species
8. parameter measured for similar metal ion for similar species
9. parameter measured for any metal ion for specific species
10. parameter measured for any metal ion for similar species

The search process started by looking for the specific parameter for each individual organism for each ENM. If no ENM specific parameters could be identified, the metal ion equivalent was explored. If parameters for the specific metal ion could not be found, then other commonly studied metal ions (e.g. mercury, cadmium, etc.) were investigated. This was done separately species by species for each parameter and ENM. When a parameter for the specific species could not be identified, the parameter for a similar species (e.g. *D. pulex* instead of *D. magna*) was substituted. The uncertainty increases moving down the ranking table.

Bioaccumulation Equations by Species (parameters are defined in Tables S2-S4)

1. *S. capricornutum*

$$\begin{aligned}\frac{dC_{b,1}}{dt} &= k_{u1,1}C_w - k_{e,1}C_{b,1} - k_{dis}C_{b,1} - D_1C_{b,1} \\ \frac{dC_{dis,1}}{dt} &= k_{u3,1}\alpha_{bio1}C_{w,dis} + k_{dis}C_{b,1} - k_{edis,1}C_{dis,1} - D_1C_{b,1}\end{aligned}$$

2. *F. crotonensis*

$$\begin{aligned}\frac{dC_{b,2}}{dt} &= k_{u1,2}C_w - k_{e,2}C_{b,2} - k_{dis}C_{b,2} - D_2C_{b,2} \\ \frac{dC_{dis,2}}{dt} &= k_{u3,2}\alpha_{bio2}C_{w,dis} + k_{dis}C_{b,2} - k_{edis,2}C_{dis,2} - D_2C_{b,2}\end{aligned}$$

3. *D. magna*

$$\begin{aligned}\frac{dC_{b,3}}{dt} &= k_{u1,3}C_w + k_{u2,3}C_{ss} + \alpha_3\alpha_{ENM,3}k_{d,3}C_{b,1} - k_{e,3}C_{b,3} - k_{dis}C_{b,3} - D_3C_{b,3} \\ \frac{dC_{dis,3}}{dt} &= k_{u3,3}\alpha_{bio3}C_{w,dis} + k_{dis}C_{b,3} - k_{edis,3}C_{dis,3} - D_3C_{b,3}\end{aligned}$$

4. *H. azteca*

$$\begin{aligned}\frac{dC_{b,4}}{dt} &= k_{u1,4}C_w + k_{u2,4}C_{sed} + \alpha_4\alpha_{ENM,4}k_{d,4}C_{b,1} - k_{e,4}C_{b,4} - k_{dis}C_{b,4} - D_4C_{b,4} \\ \frac{dC_{dis,4}}{dt} &= k_{u3,4}\alpha_{bio4}C_{w,dis} + k_{dis}C_{b,4} - k_{edis,4}C_{dis,4} - D_4C_{b,4}\end{aligned}$$

5. *V. constricta*

$$\begin{aligned}\frac{dC_{b,5}}{dt} &= k_{u1,5}C_w + k_{u2,5}C_{sed} + \alpha_5\alpha_{ENM,5}k_{d,5}C_{b,3} - k_{e,5}C_{b,5} - k_{dis}C_{b,5} - D_5C_{b,5} \\ \frac{dC_{dis,5}}{dt} &= k_{u3,5}\alpha_{bio5}C_{w,dis} + k_{dis}C_{b,5} - k_{edis,n}C_{dis,5} - D_5C_{b,5}\end{aligned}$$

6. *P. promelas*

$$\begin{aligned}\frac{dC_{b,6}}{dt} &= k_{u1,6}C_w + \alpha_6\alpha_{ENM,6}k_{d,6}C_{b,3} - k_{e,6}C_{b,6} - k_{dis}C_{b,6} - D_6C_{b,6} \\ \frac{dC_{dis,6}}{dt} &= k_{u3,6}\alpha_{bio6}C_{w,dis} + k_{dis}C_{b,6} - k_{edis,6}C_{dis,6} - D_6C_{b,6}\end{aligned}$$

7. *O. mykiss*

$$\begin{aligned}\frac{dC_{b,7}}{dt} &= k_{u1,7}C_w + \alpha_7\alpha_{ENM,7}k_{d,7}C_{b,6} - k_{e,7}C_{b,7} - k_{dis}C_{b,7} - D_7C_{b,7} \\ \frac{dC_{dis,7}}{dt} &= k_{u3,7}\alpha_{bio7}C_{w,dis} + k_{dis}C_{b,7} - k_{edis,7}C_{dis,7} - D_7C_{b,7}\end{aligned}$$

Table S1. Environmental parameter values used in nanoBio model for San Francisco Bay

Definition	Parameter	Value	Units
Volume of water compartment	V_W	1.5956E+12	L
Volume of suspended sediment compartment	V_{ss}	1.5956E+07	kg
Volume of sediment solids compartment	V_{sed}	3.8295E+10	kg
Volume Sediment Water compartment	V_{sedw}	6.3825E+10	L

Table S2. ENM dissolution rates considered in nanoFate and nanoBio

ENM	Dissolution Rate (1/day)	Metal ion Bioavailability (α_{bio})	References
CuO	0.00195	0.116	1-13
TiO ₂	0	0	
ZnO	0.08675	0.7542	1,6,10,11,13-30

Table S3. Species specific parameter values considered in nanoBio

Definition	Parameter	Units	<i>S. capricornutum</i>	<i>F. crotonensis</i>	<i>D. magna</i>	<i>H. azteca</i>	<i>V. constricta</i>	<i>P. promelas</i>	<i>O. mykiss</i>
Wet body mass of individual organism	M_i	mg	$3.58 \cdot 10^{-8}$	$6.8 \cdot 10^{-7}$	3	8	$2.657 \cdot 10^3$	$3 \cdot 10^3$	$4 \cdot 10^6$
Lifespan	L_n	days	2	3	60	365	3650	912.5	2920
Biomass density	B_n	mg/m ³	1	1	12.39	52.17	40.83	1.67	$2.65 \cdot 10^{-2}$
Species pH	pH	--	7.32	7.32	7.32	6	7	4.5	5
Assimilation Efficiency	α_n	--	NA	NA	0.2	0.5	0.5	0.35	0.8

Table S4. ENM-specific parameter values for the various species used in nanoBio

CuO									
Definition	Parameter	Units	<i>S. capricornutum</i>	<i>F. crotonensis</i>	<i>D. magna</i>	<i>H. azteca</i>	<i>V. constricta</i>	<i>P. promelas</i>	<i>O. mykiss</i>
Uptake from water	$k_{u1,n}$	L/mg-day	$2.78 \cdot 10^{-2}$	$2.78 \cdot 10^{-1}$	$1.6 \cdot 10^{-1}$	$5.79 \cdot 10^{-4}$	$7.68 \cdot 10^{-3}$	$9.6 \cdot 10^{-4}$	$4.32 \cdot 10^{-5}$
Uptake of solids	$k_{u2,n}$	L/mg-day	NA	NA	$1.6 \cdot 10^{-14}$	$4.8 \cdot 10^{-5}$	$7.68 \cdot 10^{-5}$	0	0
Uptake of dissolved ion from water	$k_{u3,n}$	L/mg-day	$1.41 \cdot 10^{-7}$	$1.41 \cdot 10^{-7}$	$1.6 \cdot 10^{-2}$	$8.66 \cdot 10^{-1}$	$7.29 \cdot 10^{-3}$	$5.13 \cdot 10^{-3}$	$6.3 \cdot 10^{-2}$
Ingestion rate	$k_{d,n}$	g/mg-day	NA	NA	$1.6 \cdot 10^{-8}$	$4.8 \cdot 10^{-5}$	$7.68 \cdot 10^{-6}$	$7.02 \cdot 10^{-6}$	$7.69 \cdot 10^{-2}$
ENM/Metallic Assimilation Efficiency	α_{ENM}	--	NA	NA	0.332	0.72	0.41	0.06	0.06
Fecal elimination rate	$k_{e,n}$	g/mg-day	$1.73 \cdot 10^{-4}$	$3 \cdot 10^{-3}$	0.029	$6.38 \cdot 10^{-5}$	$7.68 \cdot 10^{-6}$	$7.02 \cdot 10^{-7}$	$2 \cdot 10^{-6}$
Fecal elimination rate of dissolved ion	$k_{edis,n}$	g/mg-day	0	0	0.29	$2.52 \cdot 10^{-3}$	1.92	$1.9 \cdot 10^{-2}$	$1.7 \cdot 10^{-2}$
TiO ₂									
Definition	Parameter	Units	<i>S. capricornutum</i>	<i>F. crotonensis</i>	<i>D. magna</i>	<i>H. azteca</i>	<i>V. constricta</i>	<i>P. promelas</i>	<i>O. mykiss</i>
Uptake from water	$k_{u1,n}$	L/mg-day	$3.25 \cdot 10^{-3}$	$3.25 \cdot 10^{-3}$	$1.6 \cdot 10^{-1}$	$5.79 \cdot 10^{-4}$	$7.68 \cdot 10^{-3}$	$9.6 \cdot 10^{-4}$	$4.32 \cdot 10^{-5}$
Uptake of solids	$k_{u2,n}$	L/mg-day	0	0	$1.6 \cdot 10^{-4}$	$4.8 \cdot 10^{-5}$	$7.68 \cdot 10^{-5}$	0	0
Uptake of dissolved ion from water	$k_{u3,n}$	L/mg-day	$1.41 \cdot 10^{-7}$	$1.41 \cdot 10^{-7}$	$1.6 \cdot 10^{-2}$	$4.44 \cdot 10^{-1}$	$7.29 \cdot 10^{-3}$	$5.13 \cdot 10^{-3}$	$2 \cdot 10^{-6}$
Ingestion rate	$k_{d,n}$	g/mg-day	NA	NA	$1.6 \cdot 10^{-8}$	$4.8 \cdot 10^{-5}$	$7.68 \cdot 10^{-6}$	$7.02 \cdot 10^{-6}$	$7.69 \cdot 10^{-2}$
ENM/Metallic Assimilation Efficiency	α_{ENM}	--	NA	NA	0.326	0.137	0.03	0.3	0.3
Fecal elimination rate	$k_{e,n}$	g/mg-day	$1.73 \cdot 10^{-4}$	$3 \cdot 10^{-4}$	$1.6 \cdot 10^{-3}$	$6.38 \cdot 10^{-5}$	$7.68 \cdot 10^{-6}$	$7.02 \cdot 10^{-7}$	$2 \cdot 10^{-6}$
Fecal elimination rate of dissolved ion	$k_{edis,n}$	g/mg-day	0	0	0.29	$1.04 \cdot 10^{-2}$	1.92	$1.9 \cdot 10^{-2}$	$2 \cdot 10^{-3}$
ZnO									
Definition	Parameter	Units	<i>S.</i>	<i>F.</i>	<i>D. magna</i>	<i>H. azteca</i>	<i>V.</i>	<i>P.</i>	<i>O. mykiss</i>

			<i>capricornutum</i>	<i>crotonensis</i>			<i>constricta</i>	<i>promelas</i>	
Uptake from water	$k_{u1,n}$	L/mg-day	$3.19 \cdot 10^{-2}$	$3.19 \cdot 10^{-2}$	$1.6 \cdot 10^{-2}$	$5.79 \cdot 10^{-4}$	$7.68 \cdot 10^{-3}$	$9.6 \cdot 10^{-4}$	$4.32 \cdot 10^{-5}$
Uptake of solids	$k_{u2,n}$	L/mg-day	0	0	$1.5 \cdot 10^{-4}$	$4.8 \cdot 10^{-5}$	$7.68 \cdot 10^{-2}$	0	0
Uptake of dissolved ion from water	$k_{u3,n}$	L/mg-day	$1.41 \cdot 10^{-7}$	$1.41 \cdot 10^{-7}$	$2.1 \cdot 10^{-4}$	0.44	$7.29 \cdot 10^{-3}$	$5.13 \cdot 10^{-3}$	$2 \cdot 10^{-6}$
Ingestion rate	$k_{d,n}$	g/mg-day	NA	NA	$1.6 \cdot 10^{-8}$	$4.8 \cdot 10^{-5}$	$7.68 \cdot 10^{-6}$	$7.02 \cdot 10^{-6}$	$7.69 \cdot 10^{-2}$
ENM/Metallic Assimilation Efficiency	α_{ENM}	--	NA	NA	0.33	0.49	0.332	0.3	0.3
Fecal elimination rate	$k_{e,n}$	g/mg-day	$1.73 \cdot 10^{-4}$	$3 \cdot 10^{-4}$	$1.6 \cdot 10^{-2}$	$6.38 \cdot 10^{-5}$	$7.68 \cdot 10^{-6}$	$7.02 \cdot 10^{-7}$	$2 \cdot 10^{-6}$
Fecal elimination rate of dissolved ion	$k_{edis,n}$	g/mg-day	0	0	0.29	$1.04 \cdot 10^{-2}$	1.92	$1.9 \cdot 10^{-2}$	$1.1 \cdot 10^{-2}$

References CuO: ³¹⁻⁶⁷; TiO₂: ^{31-35,37-70}; ZnO: ^{31-35,37-69,71-75}

#adsorption rate of nanoparticle to particulate matter

Table S5. Summary of exposure conditions and concentrations in biological tissue for laboratory experiments

ENM	Species	Medium	Exposure (mg/L or mg/kg for dietary and/or sediment exposure)	Time Period (days)	Concentration in Biological Tissue (mg/g)	Ref.
CuO	<i>M. balthica</i>	Sed.	147.7 mg/kg DW	35	0.9 – 2.4 DW	76
CuO	<i>P. antipodarum</i>	Sed.	240 mg/kg DW	14	0.05 – 0.065 DW	77
CuO	<i>L. variegatus</i>	Sed.	40 mg/kg DW	10	0.058 DW	78
CuO	<i>C. carpio</i>	FW	100 mg/L	30	0.07 – 0.13 DW	79
CuO	<i>L. stagnalis</i>	FW	0.013 mg/L	3-5	0.26 DW	9
CuO	<i>C. Vulgaris</i>	FW	50 mg/L	1	3.5 mg Cu/g C	79
CuO	<i>D. magna</i>	FW	0.07 mg/L	9	1.84 DW	79
TiO ₂	<i>D. magna</i>	FW	0.1 mg/L	1	4.52 DW	80
TiO ₂	<i>D. magna</i>	FW	1 mg/L	1	61.1 DW	80
TiO ₂	<i>D. magna</i>	FW	2 mg/L	4	92.3 DW	81
TiO ₂	<i>D. rerio</i>	FW	4,520 mg/kg DW [^]	14	0.107 DW	82
TiO ₂	<i>D. rerio</i>	FW	61,086 mg/kg DW [^]	14	0.522 DW	82
TiO ₂	<i>C. carpio</i>	FW	10 mg/L	10	1.60 DW	83
TiO ₂	<i>C. carpio</i>	FW	10 mg/L	25	3.39 DW	83
TiO ₂	<i>C. carpio</i>	FW	10 mg/L	10	4.95 DW	84
TiO ₂	<i>O. Mykiss</i>	FW	0.1 mg/L	14	0.12 – 0.16 DW	85
TiO ₂	<i>O. Mykiss</i>	FW	5.0 mg/L	14	0.39 – 0.88 DW	86
ZnO	<i>M. galloprovincialis</i>	Mar.	0.1 mg/L	84	0.16 – 0.18 DW	87
ZnO	<i>M. galloprovincialis</i>	Mar.	0.5 mg/L	84	0.18 – 0.31 DW	87
ZnO	<i>M. galloprovincialis</i>	Mar.	0.1	112	0.40 DW	88
ZnO	<i>O. Mykiss</i>	FW	0.5 mg/L	14	0.34 – 0.51 DW	86

[^] Dietary exposure

[§] Wet weight

[¶] DW = Dry weight

ND – no data/not reported

Bioaccumulation not reported, estimated by authors based on reported exposure and accumulation

& BMF = Biomagnification factor

*Dietary exposure plus ambient exposure

FW = freshwater

Sed. = sediment

Mar. = marine water

Ref. = reference

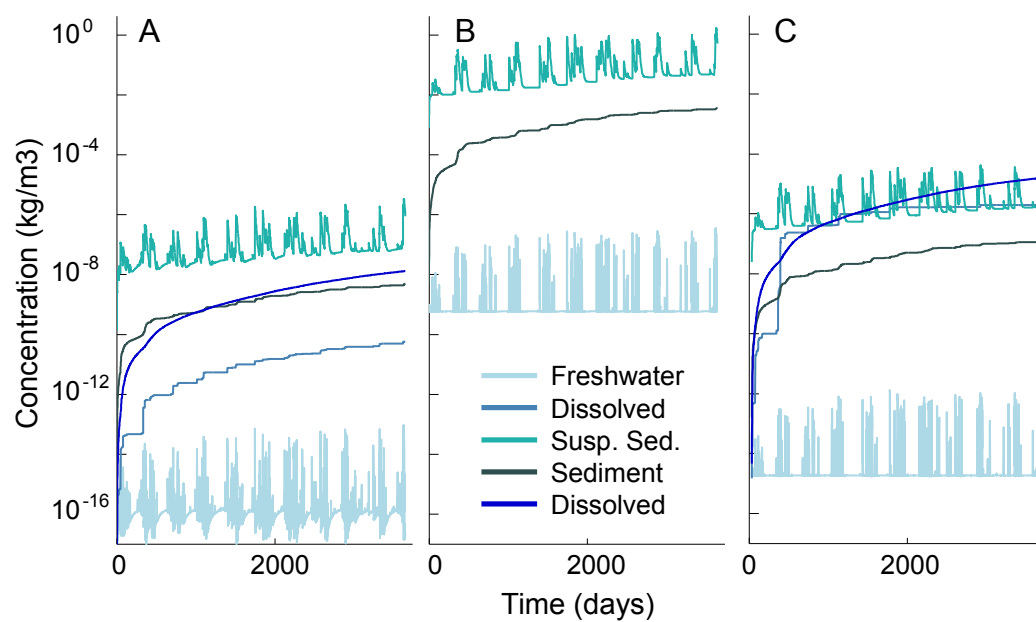


Figure S1. Environmental exposure concentrations from nanoFate model for (A) nCuO, (B) nTiO₂, and (C) nZnO.

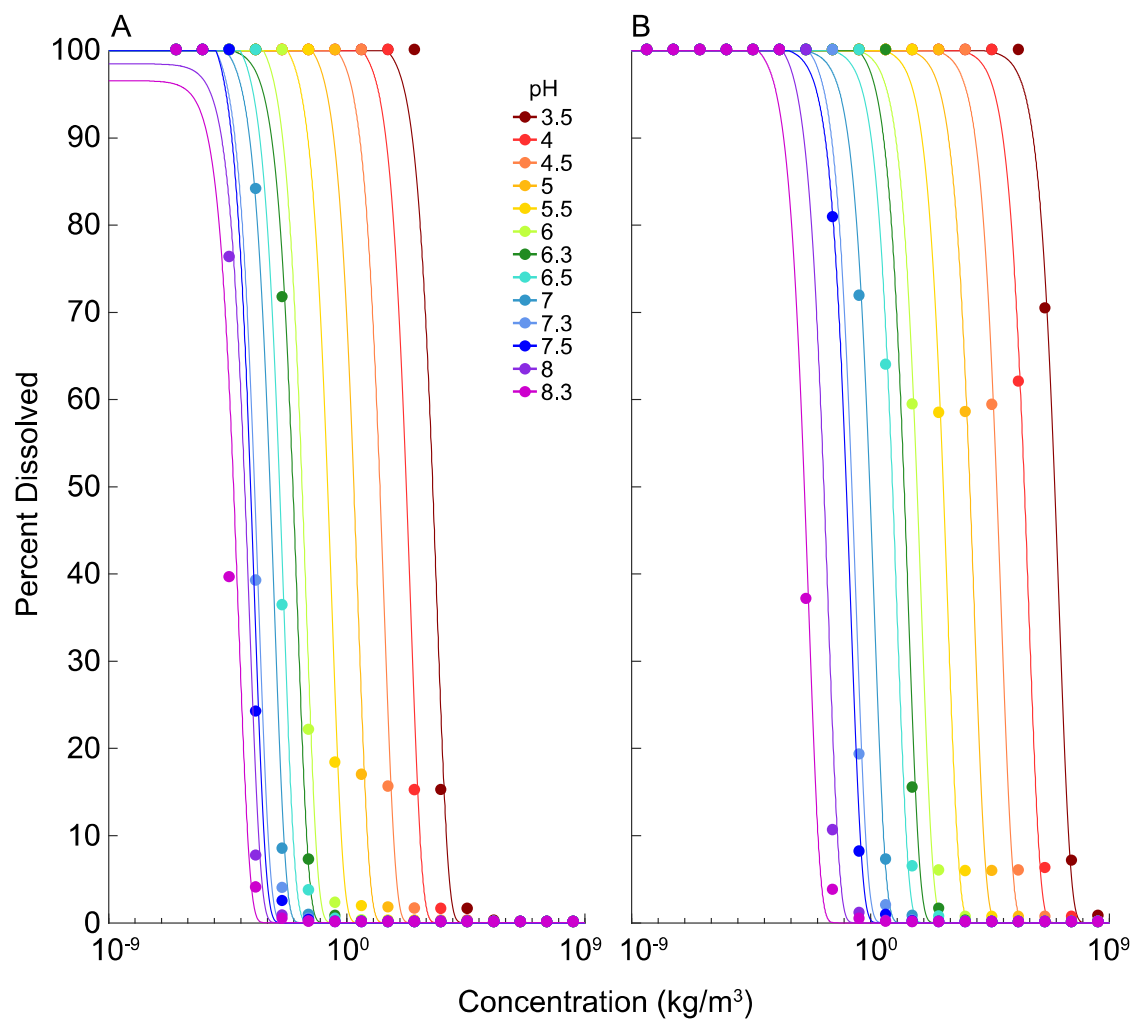


Figure S2. Dissolution curves from *MINTEq*. (A) CuO to Cu^{2+} ; and (B) ZnO to Zn^{2+} .

Table S6. Pseudo-steady state mean concentration and time to mean for nCuO

Species	nCuO		Cu ²⁺	
	Time to Pseudo-Steady-State (d)	Pseudo-State-State Concentration (ug/mg)	Time to Pseudo-Steady-State (d)	Pseudo-State-State Concentration (ug/mg)
S. capricornutum	46	1.02E-13	1100	9.03E-16
F.crotonensis	46	1.02E-13	1508	1.19E-15
D.magna	352	2.95E-13	3327	2.76E-15
H. Azteca	3327	1.16E-12	3327	2.76E-15
V.constricta	3327	1.16E-12	3327	2.76E-15
P.promelas	1748	1.01E-12	3327	2.76E-15
O. mykiss	2282	2.10E-10	3327	2.76E-15

Table S7. Pseudo-steady state mean concentration and time to mean for nTiO₂

Species	nTiO ₂		Ti ⁺	
	Time to Pseudo-Steady-State (d)	Pseudo-State-State Concentration (ug/mg)	Time to Pseudo-Steady-State (d)	Pseudo-State-State Concentration (ug/mg)
S. capricornutum	335	7.52E-08	NA	0
F.crotonensis	335	7.52E-08	NA	0
D.magna	352	1.32E-07	NA	0
H. Azteca	2893	5.17E-07	NA	0
V.constricta	2660	3.06E-07	NA	0
P.promelas	1881	1.30E-07	NA	0
O. mykiss	2660	3.06E-07	NA	0

Table S8. Pseudo-steady state mean concentration and time to mean for nZnO

Species	nZnO		Zn ²⁺	
	Time to Pseudo-Steady-State (d)	Pseudo-State-State Concentration (ug/mg)	Time to Pseudo-Steady-State (d)	Pseudo-State-State Concentration (ug/mg)
S. capricornutum	335	2.69E-12	2892	2.15E-10
F.crotonensis	335	2.69E-12	2893	2.19E-10
D.magna	352	4.81E-12	2891	2.07E-10
H. Azteca	3328	1.21E-11	2891	2.07E-10
V.constricta	2892	1.57E-11	2891	2.07E-10
P.promelas	2240	8.54E-12	2891	2.07E-10
O. mykiss	2243	1.96E-12	2891	2.07E-10

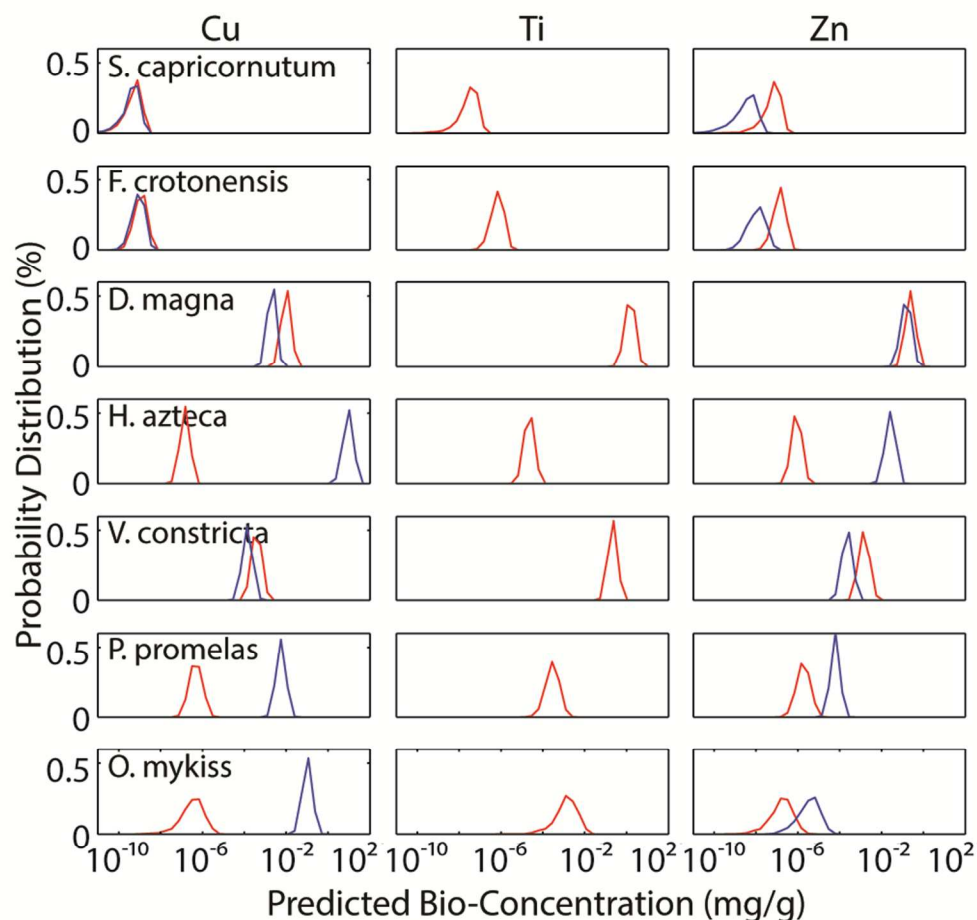


Figure S3. Probability distribution of predicted organism ENM (red) and dissolved ion (blue) concentrations. Each row is for a single species. The x-axis of each graph shows the predicted body-burden (mg/g) and the y-axis shows the frequency predicted over 10,000 Monte Carlo simulations.

Table S9. Geometric standard deviations for each chemical and biological species

Species	nCuO	Cu	nTiO ₂	nZnO	Zn
<i>S. capricornutum</i>	3.007	3.090	3.094	3.000	4.155
<i>F. crotonensis</i>	1.946	1.958	1.953	1.902	2.582
<i>D. magna</i>	1.609	1.525	1.689	1.630	1.718
<i>H. azteca</i>	1.606	1.683	1.686	1.672	1.670
<i>V. constricta</i>	1.667	1.654	1.563	1.713	1.692
<i>P. promelas</i>	1.964	1.609	2.063	2.026	1.531
<i>O. mykiss</i>	3.632	1.643	3.5053	3.505	3.328

Table S10. Sensitivity ranking of all model parameters for each ENM

CuO								
Definition	Parameter	S. capricornutum	F. crotonensis	D. magna	H. azteca	V. constricta	P. promelas	O. mykiss
Assimilation efficiency	α_n	0.0000	0.0000	0.0983	0.0964	0.1021	0.2144	0.3235
Biomass density	B_n	0.0951	0.0926	0.1006	0.0950	0.0998	0.0901	0.1008
Dissolution rate	k_{dis}	0.1007	0.0995	0.0953	0.1707	0.2392	0.1723	0.1502
Dissolved Ion concentration in water	C_{wdis}	0.0983	0.1015	0.0963	0.1001	0.0955	0.1087	0.1001
Elimination rate	$k_{e,n}$	0.1051	0.0947	0.1624	0.1005	0.0938	0.1062	0.0962
Elimination rate of dissolved ion	$k_{edis,n}$	0.0000	0.0000	0.0918	0.0961	0.0998	0.0962	0.1016
ENM concentration in sediment	C_{sed}	0.0992	0.0967	0.1047	0.2934	0.2627	0.1076	0.1058
ENM concentration in suspended sediment	C_{ss}	0.0928	0.1134	0.2903	0.1061	0.1019	0.2105	0.1497
ENM concentration in water	C_w	0.1815	0.2081	0.1010	0.1017	0.1012	0.1125	0.1029
Ingestion rate	$k_{d,n}$	0.0000	0.0000	0.0934	0.1009	0.0963	0.2092	0.1501
Lifespan	L_n	0.4223	0.3510	0.1361	0.1493	0.0986	0.1150	0.0930
Uptake by water	$k_{u1,n}$	0.1946	0.2242	0.0954	0.0889	0.0882	0.0993	0.1053
Uptake of dissolved ion	$k_{u3,n}$	0.0945	0.0780	0.1011	0.0981	0.0994	0.0893	0.0995
Uptake of solids	$k_{u2,n}$	0.0000	0.0000	0.2908	0.2790	0.2718	0.0000	0.0000
Wet body mass of individual organism	M_i	0.0938	0.0957	0.0906	0.1115	0.1001	0.0984	0.0957
Cu ²⁺								
Definition	Parameter	S. capricornutum	F. crotonensis	D. magna	H. azteca	V. constricta	P. promelas	O. mykiss
Assimilation efficiency	α_n	0.0000	0.0000	0.0966	0.1001	0.1019	0.1036	0.1046
Biomass density	B_n	0.1010	0.1007	0.1069	0.0943	0.0958	0.1002	0.0979
Dissolution rate	k_{dis}	0.0915	0.0975	0.1526	0.0986	0.1009	0.0995	0.0936
Dissolved Ion concentration in water	C_{wdis}	0.1918	0.2190	0.1993	0.2700	0.2539	0.2703	0.2754
Elimination rate	$k_{e,n}$	0.1030	0.1015	0.1171	0.0996	0.0972	0.0975	0.1009
Elimination rate of dissolved ion	$k_{edis,n}$	0.0000	0.0000	0.2114	0.1000	0.2626	0.1838	0.2137
ENM concentration in sediment	C_{sed}	0.1155	0.0900	0.0889	0.0972	0.1071	0.0952	0.0965
ENM concentration in suspended sediment	C_{ss}	0.0964	0.1022	0.1625	0.1058	0.0890	0.0998	0.1065
ENM concentration in water	C_w	0.0981	0.0907	0.1003	0.0989	0.0967	0.0958	0.0933
Ingestion rate	$k_{d,n}$	0.0000	0.0000	0.1010	0.1016	0.0922	0.0936	0.0990

Lifespan	L_n	0.4421	0.3619	0.1908	0.2411	0.0959	0.1350	0.1085
Uptake by water	$k_{u1,n}$	0.1033	0.1053	0.1038	0.0963	0.0974	0.0905	0.0907
Uptake of dissolved ion	$k_{u3,n}$	0.1848	0.1972	0.1910	0.2754	0.2572	0.2800	0.2785
Uptake of solids	$k_{u2,n}$	0.0000	0.0000	0.1564	0.0927	0.1000	0.0000	0.0000
Wet body mass of individual organism	M_i	0.1009	0.1018	0.1004	0.1024	0.1021	0.1053	0.0935
TiO ₂								
Definition	Parameter	S. capricornutum	F. crotonensis	D. magna	H. azteca	V. constricta	P. promelas	O. mykiss
Assimilation efficiency	α_n	0.0000	0.0000	0.0880	0.1066	0.1037	0.1932	0.3366
Biomass density	B_n	0.1002	0.0893	0.0906	0.0984	0.1007	0.0987	0.1069
Dissolution rate	k_{dis}	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000
Dissolved Ion concentration in water	C_{wdis}	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000	0.0000
Elimination rate	$k_{e,n}$	0.0961	0.0952	0.0989	0.0985	0.0929	0.0919	0.1004
Elimination rate of dissolved ion	$k_{edis,n}$	0.0000	0.0000	0.0970	0.0995	0.1001	0.0985	0.1003
ENM concentration in sediment	C_{sed}	0.0914	0.1033	0.1058	0.2486	0.2917	0.1048	0.1089
ENM concentration in suspended sediment	C_{ss}	0.1006	0.1010	0.2606	0.1081	0.0963	0.1985	0.1545
ENM concentration in water	C_w	0.1772	0.1993	0.1004	0.0927	0.1030	0.0996	0.0991
Ingestion rate	$k_{d,n}$	0.0000	0.0000	0.0865	0.0939	0.0881	0.2002	0.1531
Lifespan	L_n	0.4409	0.3567	0.2413	0.2592	0.1537	0.1765	0.1146
Uptake by water	$k_{u1,n}$	0.1970	0.2153	0.0917	0.0969	0.0908	0.0989	0.1035
Uptake of dissolved ion	$k_{u3,n}$	0.0911	0.0875	0.0992	0.0888	0.0951	0.1071	0.1048
Uptake of solids	$k_{u2,n}$	0.0000	0.0000	0.2708	0.2417	0.2986	0.0000	0.0000
Wet body mass of individual organism	M_i	0.0966	0.1091	0.1012	0.1008	0.1007	0.1088	0.0940
ZnO								
Definition	Parameter	S. capricornutum	F. crotonensis	D. magna	H. azteca	V. constricta	P. promelas	O. mykiss
Assimilation efficiency	α_n	0.0000	0.0000	0.0995	0.0985	0.1048	0.2007	0.3223
Biomass density	B_n	0.0915	0.1000	0.0950	0.0992	0.1010	0.1094	0.0987
Dissolution rate	k_{dis}	0.0929	0.1017	0.1791	0.2549	0.2620	0.1901	0.1418
Dissolved Ion concentration in water	C_{wdis}	0.1025	0.0892	0.0964	0.1014	0.0992	0.1050	0.0964
Elimination rate	$k_{e,n}$	0.1103	0.1000	0.1020	0.0993	0.1049	0.1053	0.0948
Elimination rate of dissolved ion	$k_{edis,n}$	0.0000	0.0000	0.1001	0.0931	0.0929	0.0942	0.0933

ENM concentration in sediment	C_{sed}	0.1035	0.0943	0.1070	0.2704	0.2695	0.1110	0.0982
ENM concentration in suspended sediment	C_{ss}	0.0922	0.0935	0.2958	0.1045	0.0965	0.2076	0.1402
ENM concentration in water	C_w	0.1818	0.2107	0.0941	0.0982	0.1145	0.1079	0.0984
Ingestion rate	$k_{d,n}$	0.0000	0.0000	0.0899	0.0895	0.0905	0.2028	0.1550
Lifespan	L_n	0.4295	0.3431	0.1343	0.1070	0.1073	0.0976	0.1006
Uptake by water	$k_{u1,n}$	0.2065	0.2100	0.0913	0.1062	0.1005	0.0966	0.0911
Uptake of dissolved ion	$k_{u3,n}$	0.0802	0.0968	0.1043	0.1046	0.1049	0.1012	0.0953
Uptake of solids	$k_{u2,n}$	0.0000	0.0000	0.2845	0.2542	0.2588	0.0000	0.0000
Wet body mass of individual organism	M_i	0.1056	0.0943	0.0927	0.0994	0.1114	0.0930	0.0973
Zn ²⁺								
Definition	Parameter	S. capricornutum	F. crotonensis	D. magna	H. azteca	V. constricta	P. promelas	O. mykiss
Assimilation efficiency	α_n	0.0000	0.0000	0.0903	0.0980	0.0979	0.1280	0.3308
Biomass density	B_n	0.0993	0.0966	0.0846	0.1071	0.1033	0.1086	0.0939
Dissolution rate	k_{dis}	0.1427	0.1507	0.1347	0.1053	0.1062	0.0991	0.0979
Dissolved Ion concentration in water	C_{wdis}	0.1036	0.0928	0.1036	0.2591	0.0990	0.2469	0.1011
Elimination rate	$k_{e,n}$	0.1097	0.1025	0.0967	0.0920	0.0941	0.0932	0.0967
Elimination rate of dissolved ion	$k_{edis,n}$	0.0000	0.0000	0.1616	0.1249	0.2523	0.1995	0.1373
ENM concentration in sediment	C_{sed}	0.1020	0.0945	0.1032	0.1047	0.2735	0.1088	0.0972
ENM concentration in suspended sediment	C_{ss}	0.0928	0.0941	0.2650	0.1110	0.0992	0.1451	0.1511
ENM concentration in water	C_w	0.1449	0.1658	0.0953	0.0942	0.1078	0.0958	0.1013
Ingestion rate	$k_{d,n}$	0.0000	0.0000	0.0873	0.1062	0.0880	0.1429	0.1591
Lifespan	L_n	0.4786	0.4130	0.1980	0.2177	0.0898	0.1553	0.1111
Uptake by water	$k_{u1,n}$	0.1597	0.1565	0.0940	0.0978	0.1065	0.0905	0.0963
Uptake of dissolved ion	$k_{u3,n}$	0.0815	0.0947	0.1044	0.2875	0.0886	0.2200	0.0949
Uptake of solids	$k_{u2,n}$	0.0000	0.0000	0.2472	0.0981	0.2578	0.0000	0.0000
Wet body mass of individual organism	M_i	0.1054	0.0932	0.1003	0.1003	0.1086	0.0979	0.0960

Visual MINTEQ simulation

The Visual MINTEQ model was used to predict the equilibrium concentration of free metal ion (Cu^{2+} or Zn^{2+}) in the various compartments. For the freshwater compartment, the details are presented in Table S10 – S12, since this is most relevant for the analysis presented for a freshwater aquatic food chain. To setup the Visual MINTEQ model, the composition of freshwater is presented in Table S10. The chemical species and solid phases considered for the Cu and Zn systems are presented in Table S10, as well as the equilibrium distribution at an initial concentration of $4.4\text{E-}06$ M for zincite (ZnO) and $4.5\text{E-}06$ M for tenorite (CuO). The solubility equilibrium constants (K_{so}) was adjusted to reflect particle size via the surface area. Table S12 presents the fraction of dissolved metal ion (Cu^{2+} or Zn^{2+}) in this freshwater system at different initial concentrations of CuO or ZnO . The fraction of dissolved metal ion decreases markedly as the initial concentrations increase. These responses were then built into the nanoFate model to predict the available concentration of nanoCuO and free Cu^{2+} , or nanoZnO and free Zn^{2+} .

Table S11. Freshwater composition considered in MINTEQ

pH	7.63	
IS	$1.84\text{E-}02$	equiv/kg
HCO_3^-	$3.30\text{E-}03$	mol/kg
TOC	$1.64\text{E-}04$	mol/kg
Al	$4.74\text{E-}05$	mol/kg
As (V)	$0.00\text{E+}00$	mol/kg
B	$3.42\text{E-}05$	mol/kg
Ba	$0.00\text{E+}00$	mol/kg
Ca^{2+}	$2.76\text{E-}03$	mol/kg
Cl-	$3.53\text{E-}03$	mol/kg
Fe	$0.00\text{E+}00$	mol/kg
I	$0.00\text{E+}00$	mol/kg
K+	$7.95\text{E-}05$	mol/kg
Li	$4.32\text{E-}06$	mol/kg
Mn^{2+}	$0.00\text{E+}00$	mol/kg
Mg^{2+}	$1.40\text{E-}03$	mol/kg
Na^+	$2.18\text{E-}03$	mol/kg
NO_3^-	$7.01\text{E-}05$	mol/kg
NO_2^-	$0.00\text{E+}00$	mol/kg
PO_4^{3-}	$5.90\text{E-}06$	mol/kg

Table S12. Cu and Zn speciation considered in MINTEQ

% of total concentration	Species name	% of total concentration	Species name
0.944	Cu^{+2}	38.608	Zn^{+2}
0.235	Cu DOM1	0.383	Zn DOM1
4.298	CuOH^+	5.558	ZnOH^+
0.014	Cu(OH)_3^-	13.115	$\text{Zn(OH)}_2 \text{ (aq)}$
1.48	$\text{Cu(OH)}_2 \text{ (aq)}$	0.01	Zn(OH)_3^-
0.019	$\text{Cu}_2\text{(OH)}_2^{+2}$	0.228	ZnCl^+
0.159	$\text{CuSO}_4 \text{ (aq)}$	6.192	$\text{ZnSO}_4 \text{ (aq)}$
0.013	$\text{CuHPO}_4 \text{ (aq)}$	0.116	$\text{Zn(SO}_4)_2^{-2}$
82.906	$\text{CuCO}_3 \text{ (aq)}$	0.084	$\text{ZnHPO}_4 \text{ (aq)}$
0.102	CuHCO_3^+	33.11	$\text{ZnCO}_3 \text{ (aq)}$
9.804	$\text{Cu(CO}_3)_2^{-2}$	2.084	ZnHCO_3^+
0.022	$\text{CuH}_2\text{BO}_3^+$	0.504	$\text{Zn(CO}_3)_2^{-2}$

Table S13. Fraction of dissolved Cu^{2+} or Zn^{2+} in freshwater as calculated using MINTEQ and the freshwater composition from Table S10

Cu^{2+}			Zn^{2+}		
Concentration		%	Concentration		%
kg/m^3	Molality	Dissolved	kg/m^3	Molality	Dissolved
3.57E-05	4.49E-07	100.000%	3.57E-04	4.39E-06	100.000%
3.57E-04	4.49E-06	27.287%	3.57E-03	4.39E-05	37.468%
3.57E-03	4.49E-05	2.792%	3.57E-02	4.39E-04	3.747%
3.57E-02	4.49E-04	0.273%	3.57E-01	4.39E-03	0.375%
1.00E-01	1.26E-03	0.097%	4.88E-01	6.00E-03	0.274%
2.00E-01	2.51E-03	0.049%	5.70E-01	7.00E-03	0.235%
			6.10E-01	7.50E-03	0.219%

References

- (1) Mortimer, M.; Kasemets, K.; Kahru, A. Toxicity of ZnO and CuO nanoparticles to ciliated protozoa *Tetrahymena thermophila*. *Toxicology* **2010**, *269* (2–3), 182–189.
- (2) Adeleye, A. A. S. A. S.; Conway, J. R. J. R. J.; Perez, T.; Rutten, P.; Keller, A. A. A. Influence of extracellular polymeric substances on the long-term fate, dissolution, and speciation of copper-based nanoparticles. *Environ. Sci. Technol.* **2014**, *48* (21), 12561–12568.
- (3) Buffet, P.-E.; Richard, M.; Caupos, F.; Vergnoux, A.; Perrein-Ettajani, H.; Luna-Acosta, A.; Akcha, F.; Amiard, J.-C.; Amiard-Triquet, C.; Guibbolini, M.; et al. A Mesocosm Study of Fate and Effects of CuO Nanoparticles on Endobenthic Species (*Scrobicularia plana*, *Hediste diversicolor*). *Environ. Sci. Technol.* **2013**, 130110104824003.
- (4) Bielmyer-Fraser, G. K.; Jarvis, T. A.; Lenihan, H. S.; Miller, R. J. Cellular Partitioning of Nanoparticulate versus Dissolved Metals in Marine Phytoplankton. *Environ. Sci. Technol.* **2014**, *48* (22), 13443–13450.
- (5) Conway, J. R.; Adeleye, A. S.; Gardea-Torresdey, J. L.; Keller, A. A. Aggregation, dissolution, and transformation of copper nanoparticles in natural waters. *Environ. Sci. Technol.* **2015**, *49* (5), 2749–2756.
- (6) Ivask, A.; Bondarenko, O.; Jephthina, N.; Kahru, A. Profiling of the reactive oxygen species-related ecotoxicity of CuO, ZnO, TiO₂, silver and fullerene nanoparticles using a set of recombinant luminescent *Escherichia coli* strains: differentiating the impact of particles and solubilised metals. *Anal. Bioanal. Chem.* **2010**, *398*, 701–716.
- (7) Wang, Z.; Li, J.; Zhao, J.; Xing, B. Toxicity and Internalization of CuO Nanoparticles to Prokaryotic Alga *Microcystis aeruginosa* as Affected by Dissolved Organic Matter. *Environ. Sci. Technol.* **2011**, *45* (14), 6032–6040.
- (8) Zhao, J.; Wang, Z.; Liu, X.; Xie, X.; Zhang, K.; Xing, B. Distribution of CuO nanoparticles in juvenile carp (*Cyprinus carpio*) and their potential toxicity. *J. Hazard. Mater.* **2011**, *197*, 304–310.
- (9) Croteau, M.-N.; Misra, S. K.; Luoma, S. N.; Valsami-Jones, E. Bioaccumulation and Toxicity of CuO Nanoparticles by a Freshwater Invertebrate after Waterborne and Dietborne Exposures. *Environ. Sci. Technol.* **2014**, *48* (18), 10929–10937.
- (10) Blinova, I.; Ivask, A.; Heinlaan, M.; Mortimer, M.; Kahru, A. Ecotoxicity of nanoparticles of CuO and ZnO in natural water. *Environ. Pollut.* **2010**, *158* (1), 41–47.
- (11) Aruoja, V.; Dubourguier, H.-C.; Kasemets, K.; Kahru, A. Toxicity of nanoparticles of CuO, ZnO and TiO₂ to microalgae *Pseudokirchneriella subcapitata*. *Sci. Total Environ.* **2009**, *407* (4), 1461–1468.
- (12) Griffitt, R. J.; Luo, J.; Gao, J.; Bonzongo, J.-C.; Barber, D. S. Effects of particle composition and species on toxicity of metallic nanomaterials in aquatic organisms. *Environ. Toxicol. Chem.* **2008**, *27* (9), 1972–1978.
- (13) Heinlaan, M.; Ivask, A.; Blinova, I.; Dubourguier, H.-C.; Kahru, A. Toxicity of nanosized and bulk ZnO, CuO and TiO₂ to bacteria *Vibrio fischeri* and

- crustaceans *Daphnia magna* and *Thamnocephalus platyurus*. *Chemosphere* **2008**, 71 (7), 1308–1316.
- (14) Franklin, N. M.; Rogers, N. J.; Apte, S. C.; Batley, G. E.; Gadd, G. E.; Casey, P. S. Comparative Toxicity of Nanoparticulate ZnO, Bulk ZnO, and ZnCl₂ to a Freshwater Microalga (*Pseudokirchneriella subcapitata*): The Importance of Particle Solubility. *Environ. Sci. Technol.* **2007**, 41 (24), 8484–8490.
 - (15) Poynton, H. C.; Lazorchak, J. M.; Impellitteri, C. A.; Smith, M. E.; Rogers, K.; Patra, M.; Hammer, K. A.; Allen, H. J.; Vulpe, C. D. Differential Gene Expression in *Daphnia magna* Suggests Distinct Modes of Action and Bioavailability for ZnO Nanoparticles and Zn Ions. *Environ. Sci. Technol.* **2011**, 45 (2), 762–768.
 - (16) Li, M.; Pokhrel, S.; Jin, X.; Mädler, L.; Damoiseaux, R.; Hoek, E. M. V. Stability, bioavailability, and bacterial toxicity of ZnO and iron-doped ZnO nanoparticles in aquatic media. *Environ. Sci. Technol.* **2011**, 45 (2), 755–761.
 - (17) Kool, P. L.; Ortiz, M. D.; van Gestel, C. A. M. Chronic toxicity of ZnO nanoparticles, non-nano ZnO and ZnCl₂ to *Folsomia candida* (Collembola) in relation to bioavailability in soil. *Environ. Pollut.* **2011**, 159 (10), 2713–2719.
 - (18) Waalewijn-Kool, P. L.; Diez Ortiz, M.; van Straalen, N. M.; van Gestel, C. A. M. Sorption, dissolution and pH determine the long-term equilibration and toxicity of coated and uncoated ZnO nanoparticles in soil. *Environ. Pollut.* **2013**, 178, 59–64.
 - (19) Wang, H.; Wick, R. L.; Xing, B. Toxicity of nanoparticulate and bulk ZnO, Al₂O₃ and TiO₂ to the nematode *Caenorhabditis elegans*. *Environ. Pollut.* **2009**, 157 (4), 1171–1177.
 - (20) Heggelund, L. R.; Diez-Ortiz, M.; Lofts, S.; Lahive, E.; Jurkschat, K.; Wojnarowicz, J.; Cedergreen, N.; Spurgeon, D.; Svendsen, C. Soil pH effects on the comparative toxicity of dissolved zinc, non-nano and nano ZnO to the earthworm *Eisenia fetida*. *Nanotoxicology* **2013**, 8 (5), 559–572.
 - (21) Wong, S. W. Y.; Leung, P. T. Y.; Djurišić, A. B.; Leung, K. M. Y. Toxicities of nano zinc oxide to five marine organisms: influences of aggregate size and ion solubility. *Anal. Bioanal. Chem.* **2010**, 396 (2), 609–618.
 - (22) Ma, H.; Kabengi, N. J.; Bertsch, P. M.; Unrine, J. M.; Glenn, T. C.; Williams, P. L. Comparative phototoxicity of nanoparticulate and bulk ZnO to a free-living nematode *Caenorhabditis elegans*: The importance of illumination mode and primary particle size. *Environ. Pollut.* **2011**, 159 (6), 1473–1480.
 - (23) Peng, X.; Palma, S.; Fisher, N. S.; Wong, S. S. Effect of morphology of ZnO nanostructures on their toxicity to marine algae. *Aquat. Toxicol.* **2011**, 102 (3–4), 186–196.
 - (24) Miller, R. J.; Lenihan, H. S.; Muller, E. B.; Tseng, N.; Hanna, S. K.; Keller, A. A. Impacts of metal oxide nanoparticles on marine phytoplankton. *Environ. Sci. Technol.* **2010**, 44 (19), 7329–7334.
 - (25) Fairbairn, E. A.; Keller, A. A.; Mädler, L.; Zhou, D.; Pokhrel, S.; Cherr, G. N. Metal oxide nanomaterials in seawater: Linking physicochemical characteristics with biological response in sea urchin development. *J. Hazard. Mater.* **2011**, 192 (3).
 - (26) Bian, S.-W.; Mudunkotuwa, I. A.; Rupasinghe, T.; Grassian, V. H. Aggregation and dissolution of 4 nm ZnO nanoparticles in aqueous environments:

- influence of pH, ionic strength, size, and adsorption of humic acid. *Langmuir* **2011**, 27 (10), 6059–6068.
- (27) Rathnayake, S.; Unrine, J. M.; Judy, J.; Miller, A.-F.; Rao, W.; Bertsch, P. M. Multitechnique Investigation of the pH Dependence of Phosphate Induced Transformations of ZnO Nanoparticles. *Environ. Sci. Technol.* **2014**, 48 (9), 4757–4764.
 - (28) Yu, L.; Fang, T.; Xiong, D.; Zhu, W.; Sima, X. Comparative toxicity of nano-ZnO and bulk ZnO suspensions to zebrafish and the effects of sedimentation, 'OH production and particle dissolution in distilled water. *J. Environ. Monit.* **2011**, 13 (7), 1975.
 - (29) Lv, J.; Zhang, S.; Luo, L.; Han, W.; Zhang, J.; Yang, K.; Christie, P. Dissolution and microstructural transformation of ZnO nanoparticles under the influence of phosphate. *Environ. Sci. Technol.* **2012**, 46 (13), 7215–7221.
 - (30) Reed, R. B.; Ladner, D. A.; Higgins, C. P.; Westerhoff, P.; Ranville, J. F. Solubility of nano-zinc oxide in environmentally and biologically important matrices. *Environ. Toxicol. Chem.* **2012**, 31 (1), 93–99.
 - (31) Claesson, A. Variation in cell composition and utilization of N and P for growth of *Selenastrum capricornutum*. *Physiol. Plant.* **1980**, 48 (1), 59–64.
 - (32) Reynolds, C. S. *The ecology of freshwater phytoplankton*; Cambridge University Press, 1984.
 - (33) Loayza-Muro, R.; Elias-Letts, R. Responses to the mussel *Anodonta trapesialis* (unionidae) to environmental stressors: Effect of pH, temperature, and metals on filtration rate. *Environ. Pollut.* **2007**, 149 (2), 209–215.
 - (34) Panov, V. E.; McQueen, D. J. Effects of temperature on individual growth rate and body size of a freshwater amphipod. *Can. J. Zool.* **1998**, 76 (6), 1107–1116.
 - (35) Hargrave, B. T. Distribution, Growth, and Seasonal Abundance of *Hyaella azteca* (Amphipoda) in Relation to Sediment Microflora. *J. Fish. Res. Board Canada* **1970**, 27 (4), 685–699.
 - (36) Carbonell, G.; Tarazona, J. V. Toxicokinetics of copper in rainbow trout. *Aquat. Toxicol.* **1994**, 29, 213–221.
 - (37) Hargrave, B. T. The utilization of benthic microflora by *Hyaella azteca* (amphipoda). *J. Anim. Ecol.* **1970**, 39 (2), 427–437.
 - (38) Hargrave, B. T. Prediction of egestion by the despoti-feeding amphipod *hyaella azteca*. *Oikos* **1972**, 23 (1), 116–124.
 - (39) Bertram, P. E.; Brooks, A. S. Kinetics of accumulation of selenium from food and water by fathead minnows. *Water Res.* **1986**, 20 (7), 877–884.
 - (40) Borgmann, U.; Norwood, W. P. Kinetics of excess (above background) copper and zinc in *Hyaella azteca* and their relationship to chronic toxicity. *Can. J. Fish. Aquat. Sci.* **1994**, 52 (4), 864–874.
 - (41) Herwig, B. R.; Zimmer, K. D. Population ecology and prey consumption by fathead minnows in prairie wetlands: importance of detritus and larval fish. *Ecol. Freshw. Fish* **2007**, 16 (3), 282–294.
 - (42) van Dam, A. A.; Penning De Vries, F. W. T. Parameterization and calibration of a model to simulate effects of feeding level and feed composition on growth of *Oreochromis niloticus* and *Oncorhynchus mykiss*. *Aquac. Res.* **1995**, 26, 415–

425.

- (43) Burns, C. W. Relation Between Filtering Rate, Temperature, and Body Size in Four Species of Daphnia. *Limnol. Oceanogr.* **1969**, *14*, 693–700.
- (44) Taipale, S.; Brett, M.; Martin-Creuzburg, D.; Yeung, S.; Hiltunen, M.; Strandberg, U.; Kankaala, P. Differing daphnia magna assimilation efficiencies for terrestrial, bacterial, and algal carbon and fatty acids. *Ecology* **2014**, *95* (2), 563–576.
- (45) Patterson, M. A. Energy Reserves in native freshwater mussels (bivalvia: unionidae) with and without attached zebra mussels: effects of food deprivation, Virginia Tech: Virginia, 1998.
- (46) Lapointe, D.; Couture, P. Influence of the Route of Exposure on the Accumulation and Subcellular distribution of nickel and thallium in juvenile fathead minnows. *Arch. Environ. Contam. Toxicol.* **2009**, *57*, 571–580.
- (47) Thomann, R. V.; Shkreli, F.; Harrison, S. A pharmacokinetic model of cadmium in rainbow trout. *Environ. Toxicol. Chem.* **1997**, *16* (11), 2268–2274.
- (48) Nalewajko, C. Photosynthesis and excretion in various planktonic algae. *Limnol. Oceanogr.* **1966**, *11* (1), 1–10.
- (49) Le Faucheur, S.; Campbell, P. G. ; Fortin, C.; Slaveykova, V. L. Interactions between mercury and phytoplankton: SPeciation, bioavailability, and internal handling. *Environ. Toxicol. Chem.* **2014**, *33* (6), 1211–1224.
- (50) Guan, R.; Wang, W.-X. Dietary assimilation and elimination of Cd, Se, and Zn by Daphnia magna at different metal concentrations. *Environ. Toxicol.* **2004**, *23* (11), 2689–2698.
- (51) Dietz, T. H. Uptake of sodium and chloride by freshwater mussels. *Can. J. Zool.* **1979**, *57*, 156–160.
- (52) Winhold, L. Unionidae. *Animal Diversity Web*; University of Michigan.
- (53) Wen, Y. H. Life History and production of Hyalella azteca (Crustacea: Amphipoda) in a hypereutrophic prairie pond in southern Alberta. *Can. J. Zool.* **1992**, *70* (7), 1417–1424.
- (54) Othman, M. S.; Pascoe, D. Growth, Development and Reproduction of Hyalella Azteca (Saussure, 1858) in Laboratory Culture. *Crustaceana* **2001**, *74* (2), 171–181.
- (55) Elenbaas, M. Daphnia Magna. *Animal Diversity Web*.
- (56) Gotham, I. J.; Rhee, G.-Y. Effects of nitrate and phosphate limitations on cyclost growth of two freshwater diatoms. *J. Gen. Microbiol.* **1982**, *128*, 199–205.
- (57) Blaise, C.; Ferard, J.-F. *Small-scale freshwater toxicity investigations: Hazard assessment schemes*; Springer: The Netherlands, 2005; Vol. 2.
- (58) Fakioglu, O. Phytoplankton biomass impact on the lake water quality. In *Biomass Now - Cultivation and Utilization*; Darko, M., Ed.; Intech Open, 2013.
- (59) Stich, H. B.; Maier, G. Distribution and abundance of Daphnia pulicaria, a large Daphnia of the “pulex group”, in Lake Constance (Lower Lake). *Limnol. - Ecol. Manag. Inl. Waters* **2007**, *37* (4), 303–310.
- (60) Ricciardi, A.; Bourget, E. Weight-to-weight conversion factors for marine benthic macroinvertebrates. *Mar. Ecol. Prog. Sereies* **1998**, *163*, 245–251.
- (61) Harvey, B. C.; White, J. L.; Nakamoto, R. J. Habitat-specific biomass, survival,

- and growth of rainbow trout (*Oncorhynchus mykiss*) during summer in a small coastal stream. *Can. J. Fish. Aquat. Sci.* **2005**, *62*, 650–658.
- (62) Henry, R.; Simao, C. A. Evaluation of density and biomass of a bivalve population (*diploa delodontus expansus*) in a small tropical reservoir. *Rev. Hydrobiol. trop* **1984**, *17* (4), 309–318.
 - (63) Sommer, A. *Pimephales promelas* (Black-head minnow). *Animal Diversity Web*; University of Michigan.
 - (64) Ridolfi, K. *Oncorhynchus mykiss*. *Animal Diversity Web*; University of Michigan.
 - (65) Nichols, J. W.; McKim, J. M.; Lien, G. J.; Hoffman, A. D.; Bertelsen, S. L. Physiologically based toxicokinetic modeling of three waterborne chloroethanes in rainbow trout (*Oncorhynchus mykiss*). *Toxicol. Appl. Pharmacol.* **1991**, *110* (3), 374–389.
 - (66) Duffy, W. G. Population dynamics, production, and prey consumption of fathead minnows (*Pimephales promelas*) in prairie wetlands: a bioenergetics approach. *Can. J. Fish. Aquat. Sci.* **1998**, *54*, 15–27.
 - (67) Errecalde, O.; Campbell, P. G. C. Cadmium and zinc bioavailability to *selenastrum capricornutum*: accidental metal uptake and toxicity in the presence of citrate. *J. Phycol.* **2000**, *36*, 473–483.
 - (68) Aslop, D. H.; Wod, Ch. M. Influence of waterborne cations on zinc uptake and toxicity in rainbow trout, *Oncorhynchus mykiss*. *Can. J. Fish. Aquat. Sci.* **1999**, *56* (11), 2112–2119.
 - (69) Borgmann, U. A mechanistic model of copper accumulation in *hyalella azteca*. *Sci. Total Environ.* **1998**, *219* (2–3), 137–145.
 - (70) Trudel, M.; Rasmussen, J. Modeling the elimination of mercury by fish. *Environ. Sci. Technol.* **1997**, *31* (6), 1716–1722.
 - (71) Nichols, J. W.; McKim, J. M.; Andersen, M. E.; Gargas, M. L.; Clewell, H. J.; Erickson, R. J. A physiologically based toxicokinetics model for the uptake and disposition of waterborne organic chemicals in fish. *Toxicol. Appl. Pharmacol.* **1990**, *106* (3), 433–447.
 - (72) Yu, R.-Q.; Wang, W.-X. Kinetic uptake of bioavailable cadmium, selenium, and zinc by *daphnia magna*. *Environ. Toxicol.* **2002**, *21* (11), 2348–2355.
 - (73) Thomann, R. V. Bioaccumulation model of organic chemical distribution in aquatic food chains. *Environ. Sci. Technol.* **1989**, *23* (6), 699–707.
 - (74) Blaise, C.; Gagné, F.; Férard, J. f.; Eullaffroy, P. Ecotoxicity of selected nanomaterials to aquatic organisms. *Environ. Toxicol.* **2008**, *23* (5), 591–598.
 - (75) Gotham, I. J.; Rhee, G. Y. Comparative kinetic studies of phosphate-limited growth and phosphate uptake in phytoplankton in continuous culture. *J. Phycol.* **1981**, *17*, 257–265.
 - (76) Dai, L.; Syberg, K.; Banta, G. T.; Selck, H.; Forbes, V. E. Effects, Uptake, and Depuration Kinetics of Silver Oxide and Copper Oxide Nanoparticles in a Marine Deposit Feeder, *Macoma balthica*. *ACS Sustain. Chem. Eng.* **2013**, *1* (7), 760–767.
 - (77) Ramskov, T.; Selck, H.; Banta, G.; Misra, S. K.; Berhanu, D.; Valsami-Jones, E.; Forbes, V. E. Bioaccumulation and effects of different-shaped copper oxide nanoparticles in the deposit-feeding snail *Potamopyrgus antipodarum*.

Environ. Toxicol. Chem. **2014**, 33 (9), 1976–1987.

- (78) Thit, A.; Ramskov, T.; Croteau, M.-N.; Selck, H. Biodynamics of copper oxide nanoparticles and copper ions in an oligochaete - Part II: Subcellular distribution following sediment exposure. *Aquat. Toxicol.* **2016**, 180, 25–35.
- (79) Zhao, J.; Wang, Z.; Liu, X.; Xie, X.; Zhang, K.; Xing, B. Distribution of CuO nanoparticles in juvenile carp (*Cyprinus carpio*) and their potential toxicity. *J. Hazard. Mater.* **2011**, 197, 304–310.
- (80) Zhu, X.; Chang, Y.; Chen, Y. Toxicity and bioaccumulation of TiO₂ nanoparticle aggregates in *Daphnia magna*. *Chemosphere* **2010**, 78 (3), 209–215.
- (81) Rosenfeldt, R. R.; Seitz, F.; Schulz, R.; Bundschuh, M. Heavy Metal Uptake and Toxicity in the Presence of Titanium Dioxide Nanoparticles: A Factorial Approach Using *Daphnia magna*. *Environ. Sci. Technol.* **2014**, 48 (12), 6965–6972.
- (82) Zhu, X.; Wang, J.; Zhang, X.; Chang, Y.; Chen, Y. Trophic transfer of TiO₂ nanoparticles from daphnia to zebrafish in a simplified freshwater food chain. *Chemosphere* **2010**, 79 (9), 928–933.
- (83) Zhang, X.; Sun, H.; Zhang, Z.; Niu, Q.; Chen, Y.; Crittenden, J. C. Enhanced bioaccumulation of cadmium in carp in the presence of titanium dioxide nanoparticles. *Chemosphere* **2007**, 67 (1), 160–166.
- (84) Sun, H.; Zhang, X.; Niu, Q.; Chen, Y.; Crittenden, J. C. Enhanced Accumulation of Arsenate in Carp in the Presence of Titanium Dioxide Nanoparticles. *Water. Air. Soil Pollut.* **2006**, 178 (1–4), 245–254.
- (85) Federici, G.; Shaw, B. J.; Handy, R. D. Toxicity of titanium dioxide nanoparticles to rainbow trout (*Oncorhynchus mykiss*): Gill injury, oxidative stress, and other physiological effects. *Aquat. Toxicol.* **2007**, 84 (4), 415–430.
- (86) Johnston, B. D.; Scown, T. M.; Moger, J.; Cumberland, S. A.; Baalousha, M.; Linge, K.; van Aerle, R.; Jarvis, K.; Lead, J. R.; Tyler, C. R. Bioavailability of Nanoscale Metal Oxides TiO₂, CeO₂, and ZnO to Fish. *Environ. Sci. Technol.* **2010**, 44 (3), 1144–1151.
- (87) Hanna, S. K.; Miller, R. J.; Muller, E. B.; Nisbet, R. M.; Lenihan, H. S. Impact of Engineered Zinc Oxide Nanoparticles on the Individual Performance of *Mytilus galloprovincialis*. *PLoS One* **2013**, 8 (4), e61800.
- (88) Muller, E. B.; Hanna, S. K.; Lenihan, H. S.; Miller, R. J.; Nisbet, R. M. Impact of engineered zinc oxide nanoparticles on the energy budgets of *Mytilus galloprovincialis*. *J. Sea Res.* **2014**, 94, 29–36.