Supporting Information

Sulfonic Acid-Containing Flavonoids from the Roots of *Phyllanthus acidus*

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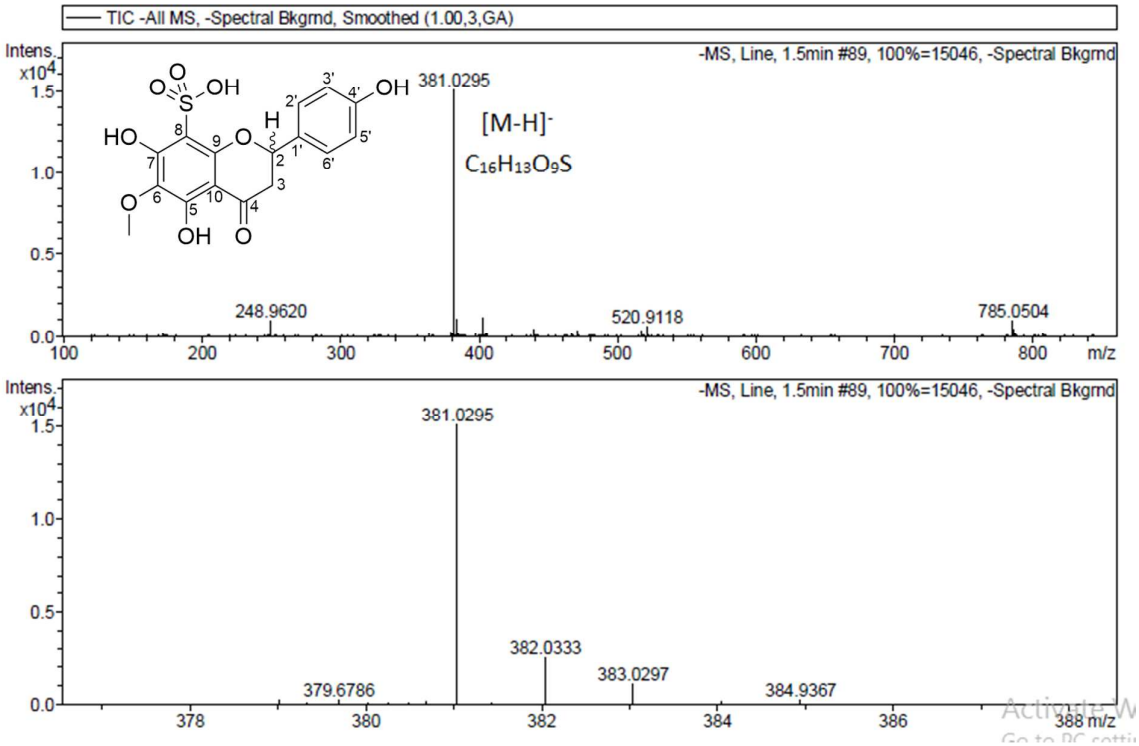
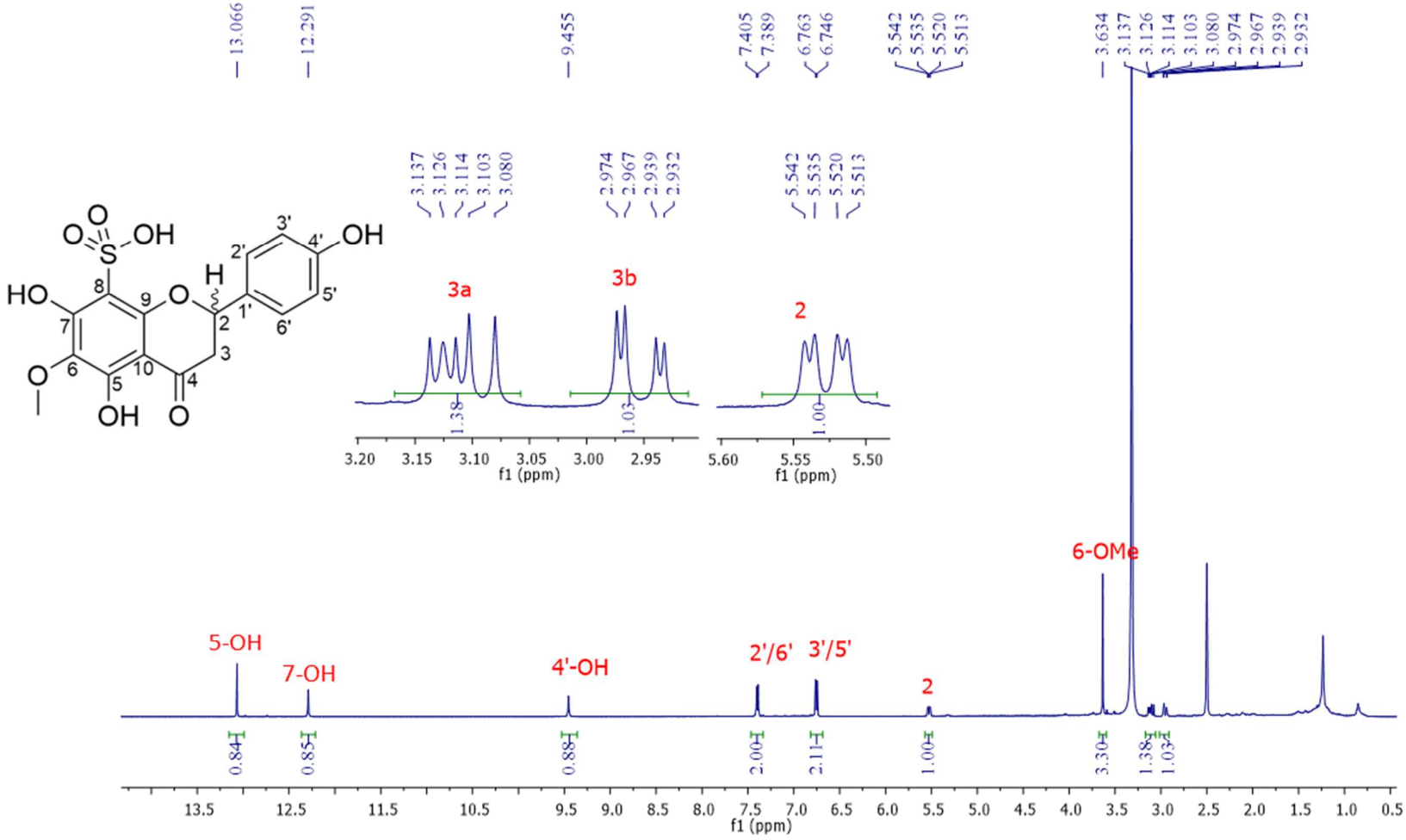
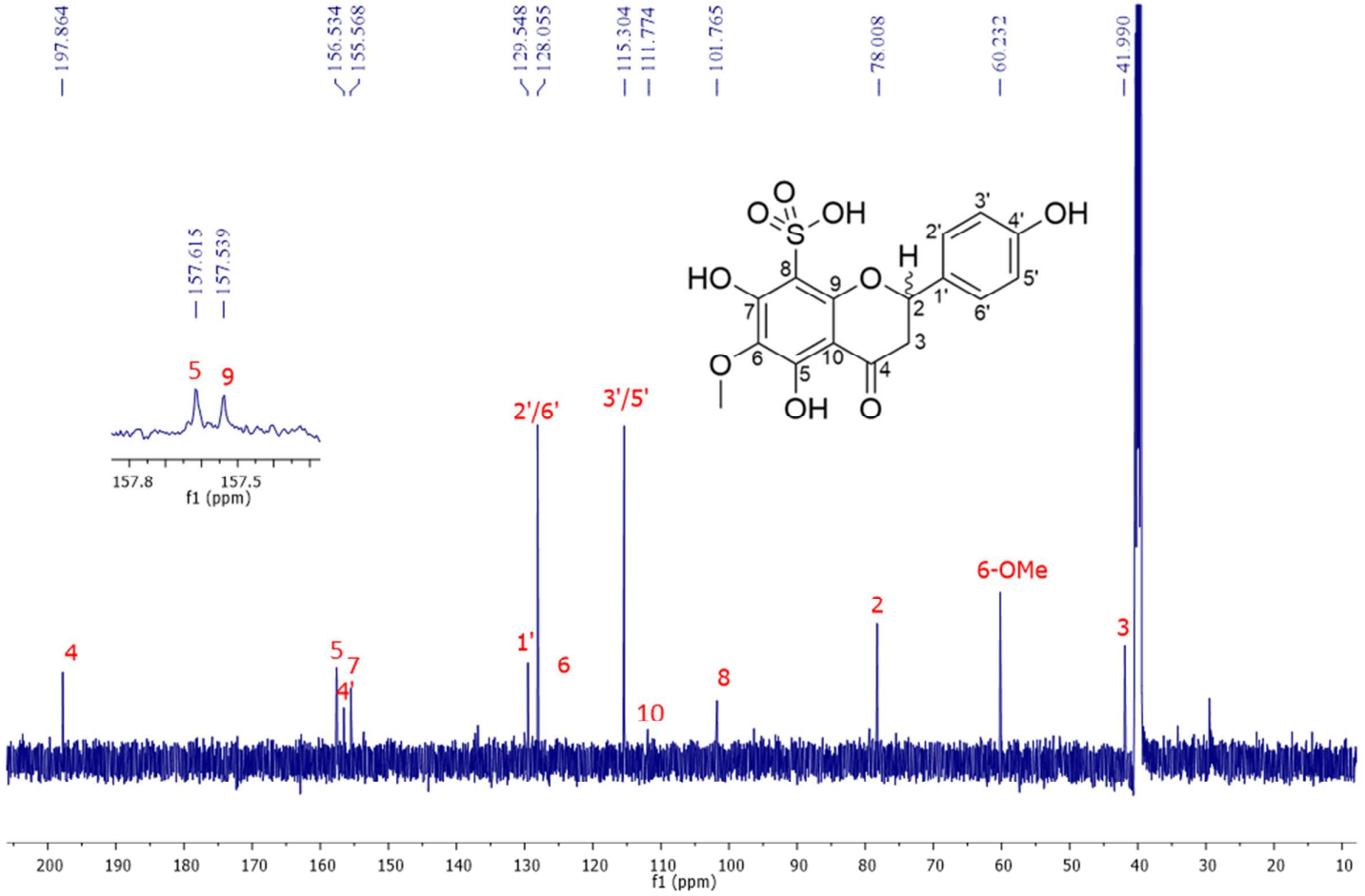


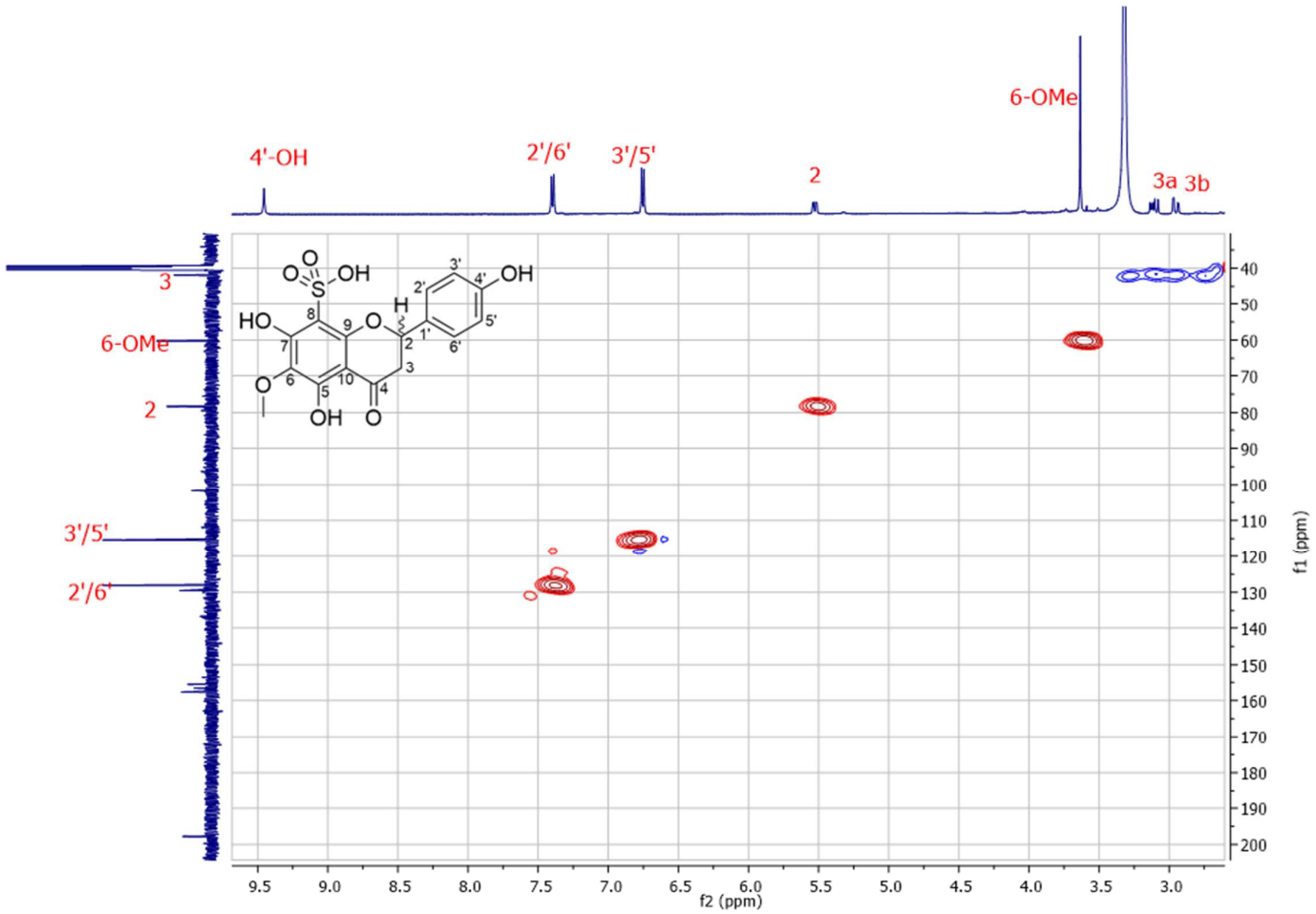
S2. ^{1}H NMR (DMSO-d_6, 500 MHz) spectrum of **1**



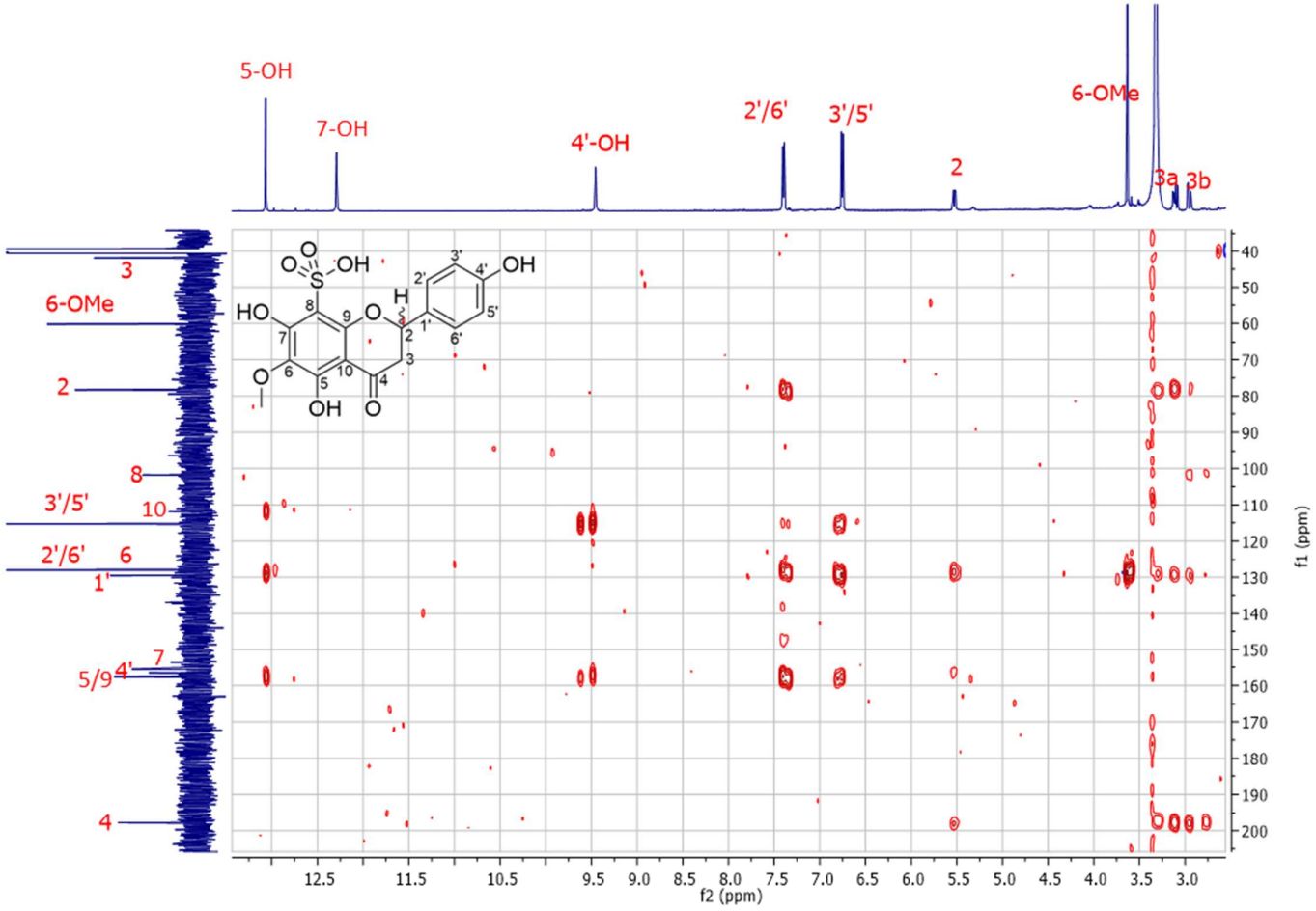
S3. ^{13}C NMR (DMSO-d_6, 125 MHz) spectrum of **1**



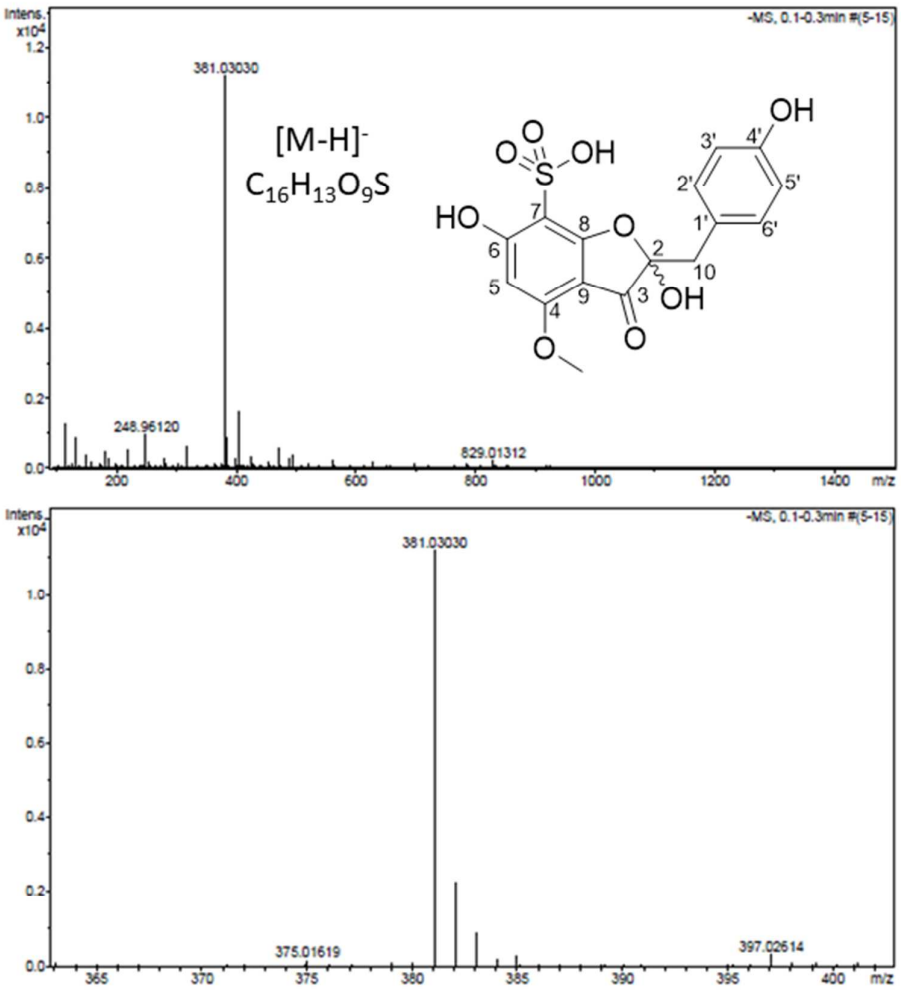
S4. HSQC (DMSO-d_6, 500 MHz, 125 MHz) spectrum of **1**



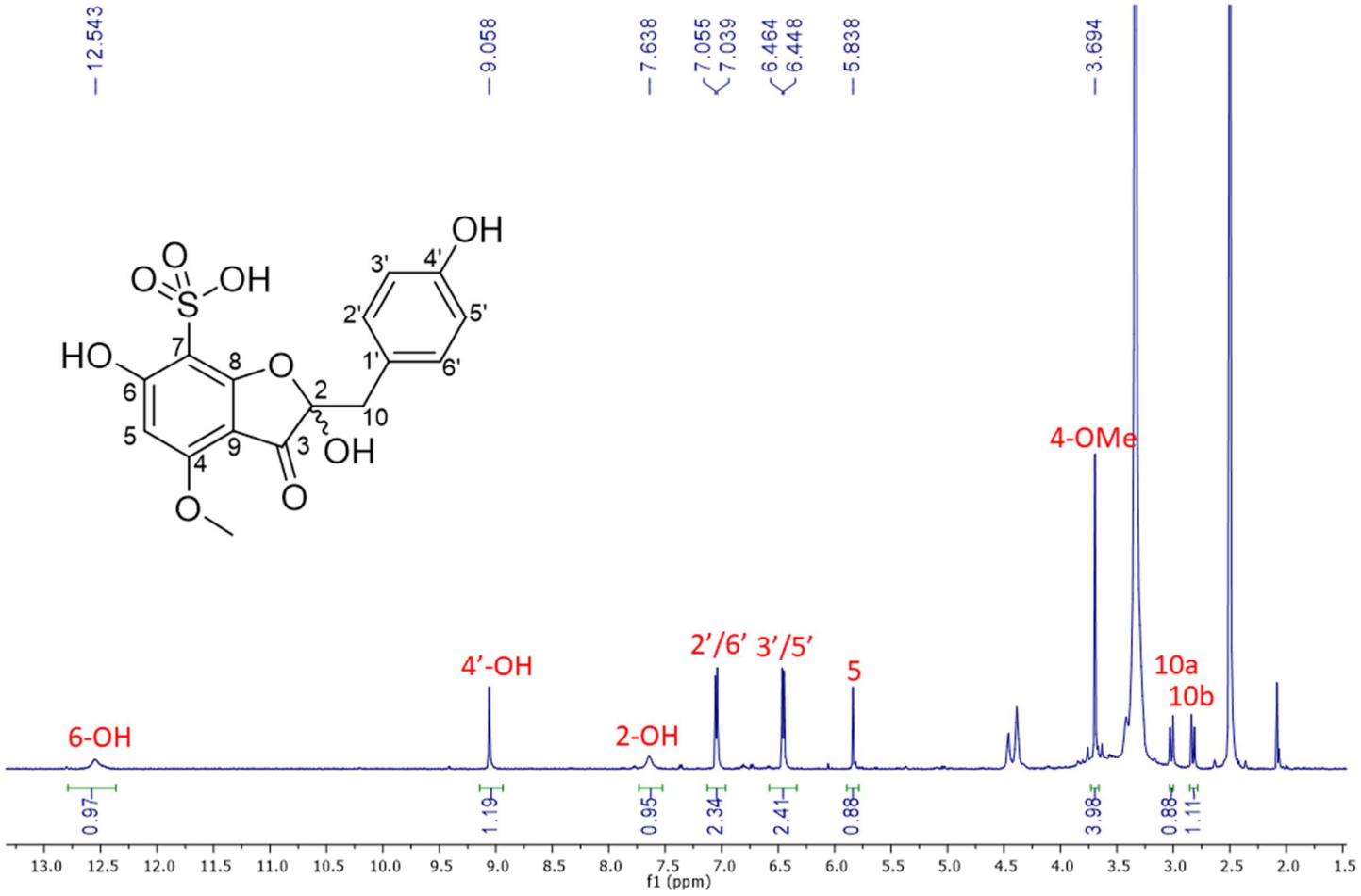
S5. HMBC (DMSO-d_6, 500 MHz, 125 MHz) spectrum of **1**



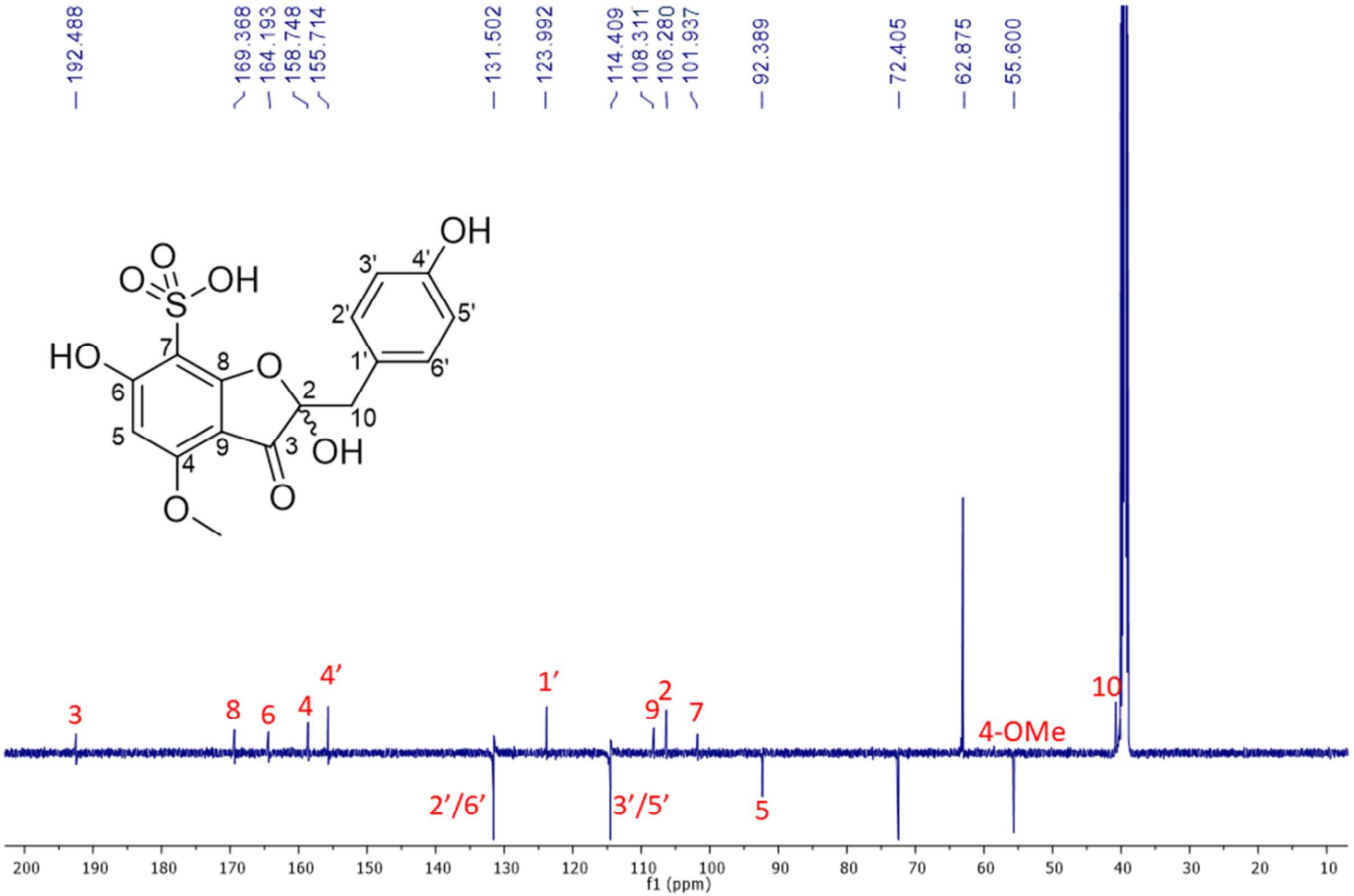
S6. HRESIMS spectrum of **2**



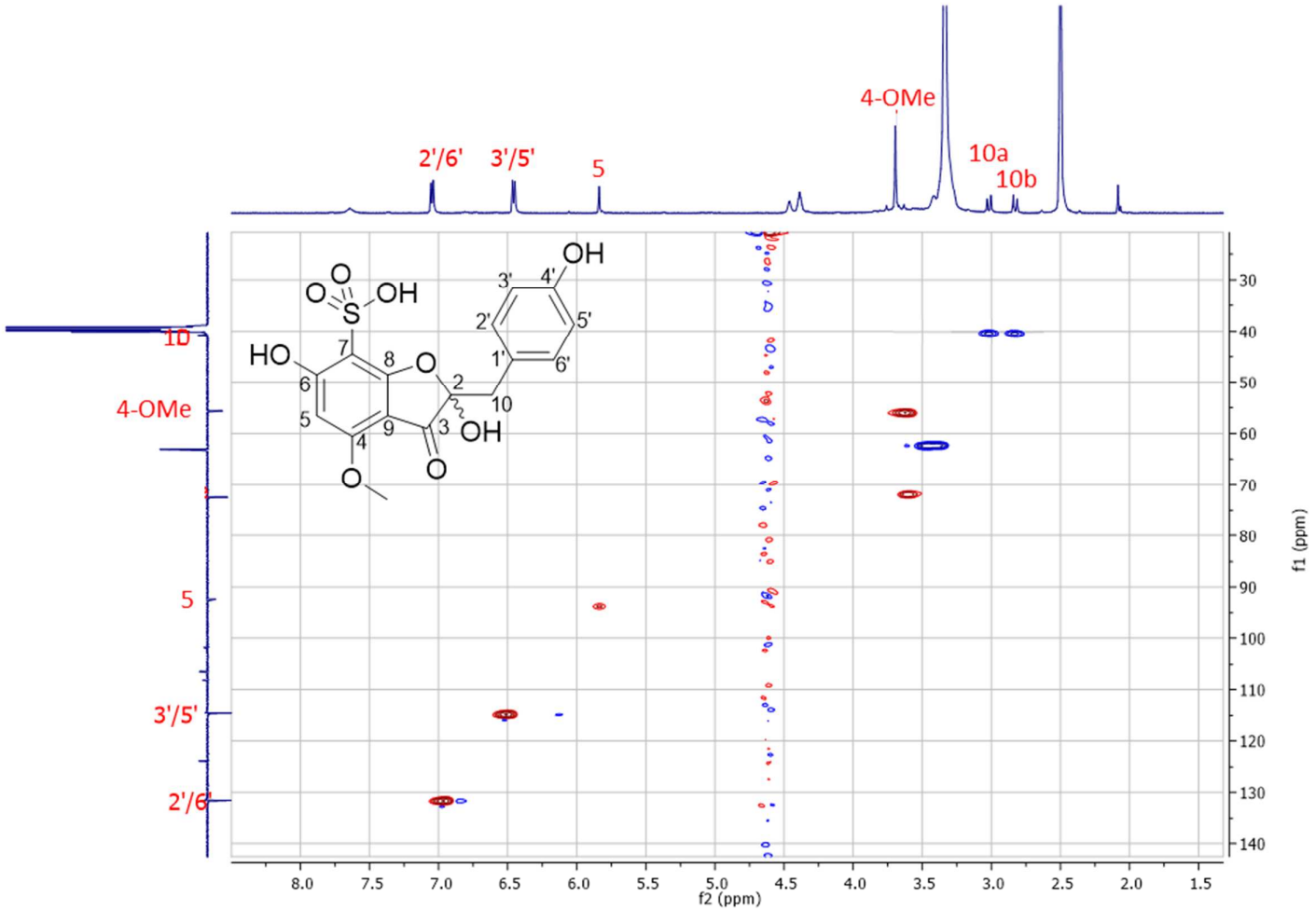
S7. ^{1}H NMR (DMSO-d_6, 500 MHz) spectrum of **2**



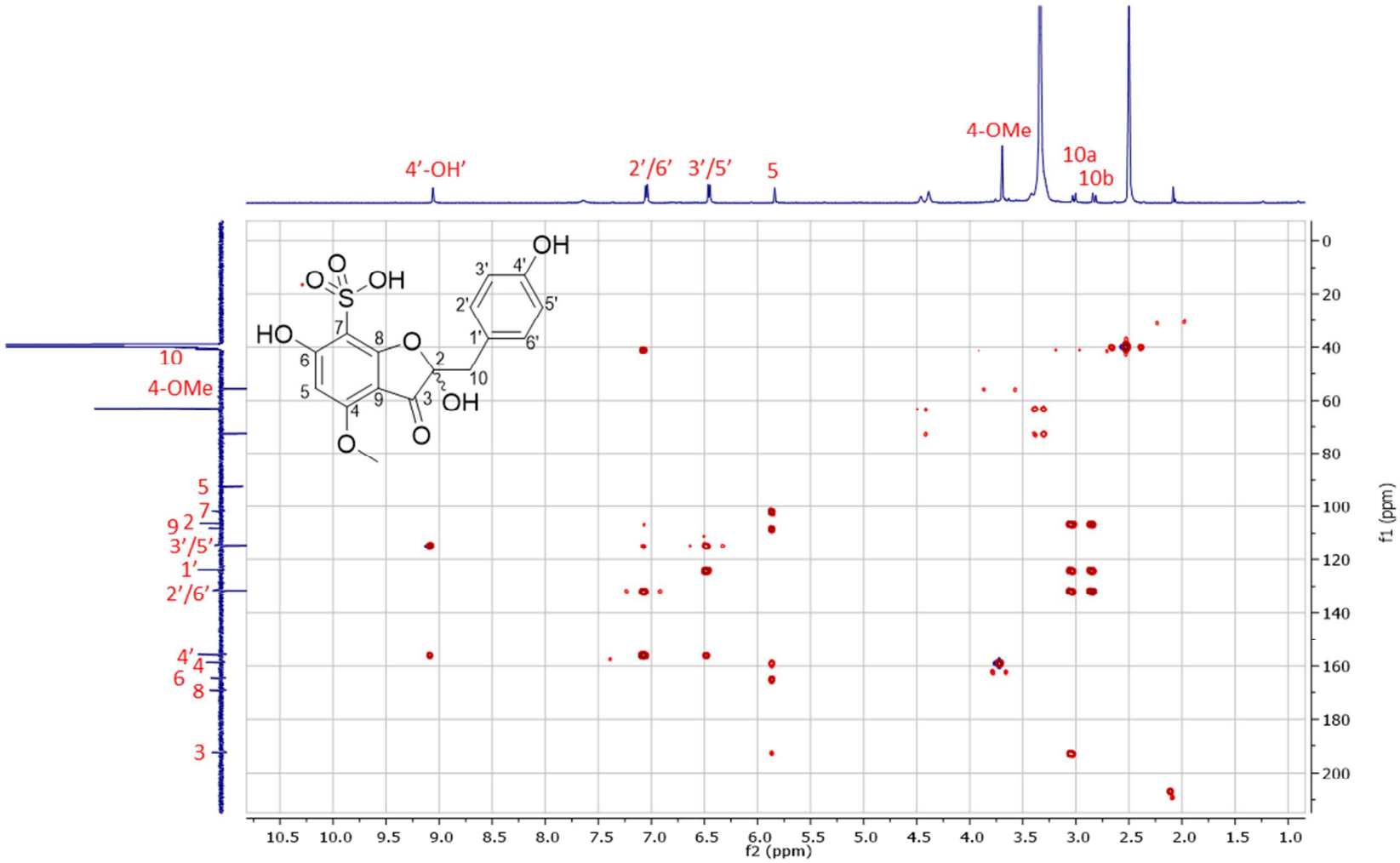
S8. JMOD (DMSO-d_6, 125 MHz) spectrum of **2**



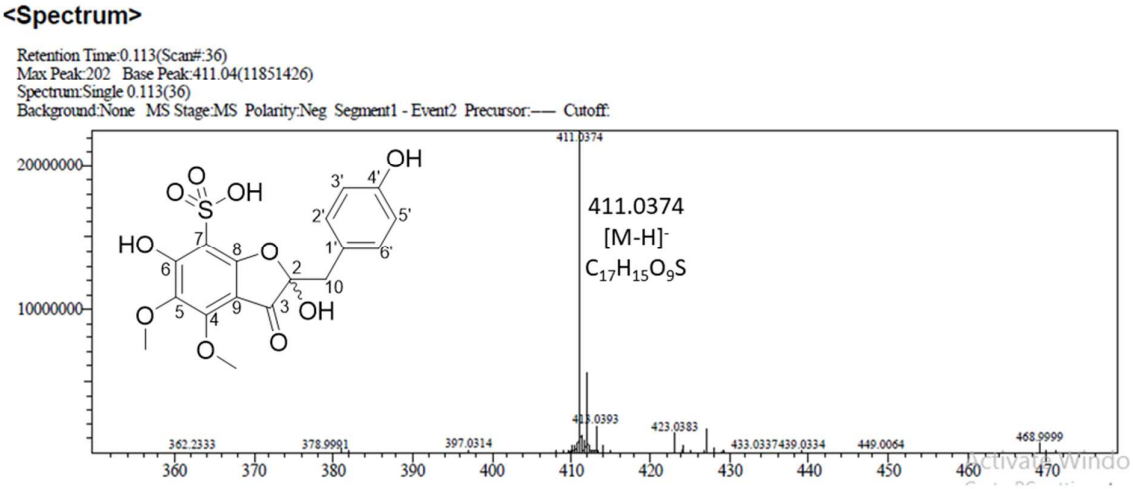
S9. HSQC (DMSO-d_6, 500 MHz, 125 MHz) spectrum of **2**



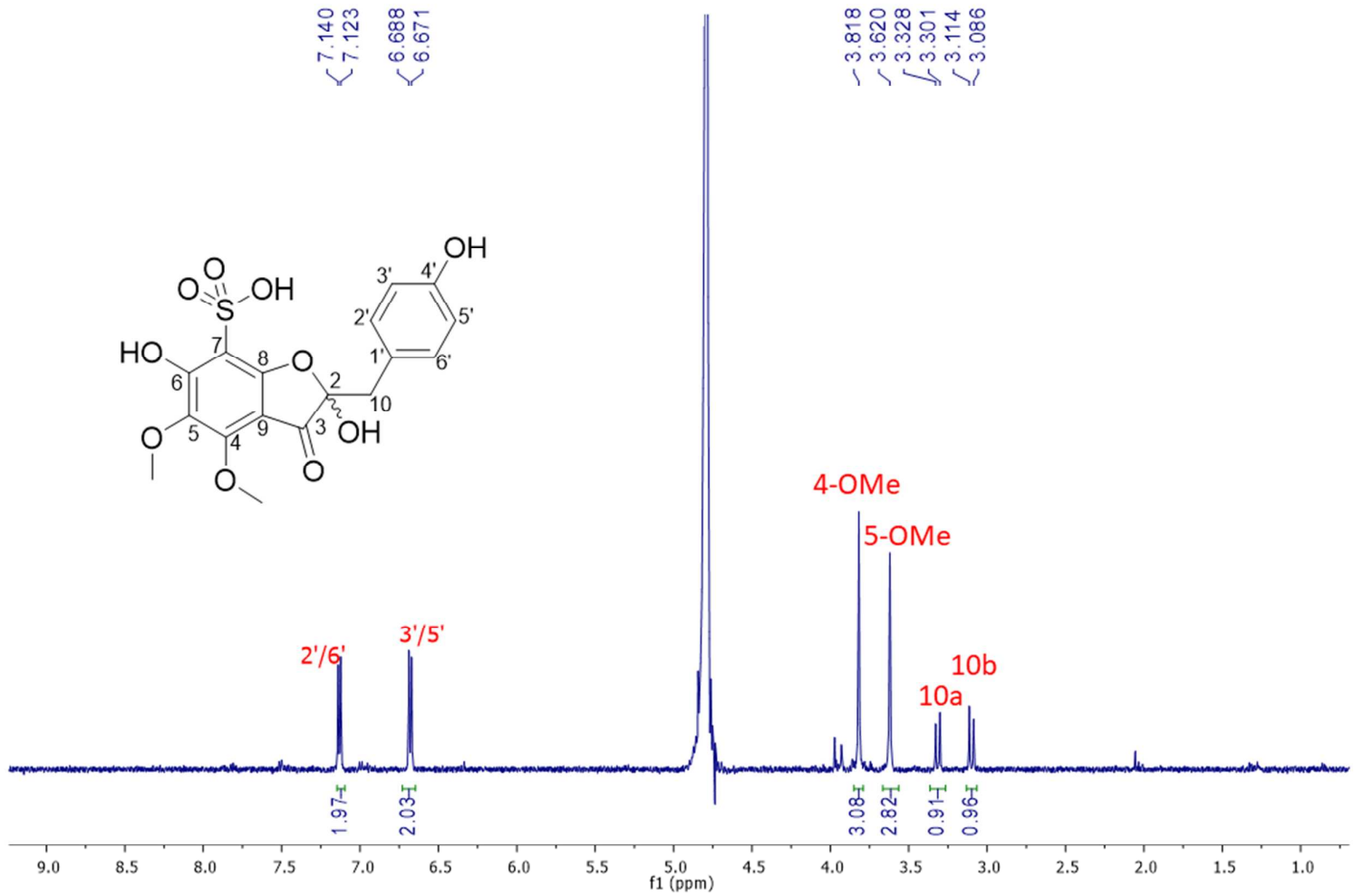
S10. HMBC (DMSO-d_6, 500 MHz, 125 MHz) spectrum of **2**



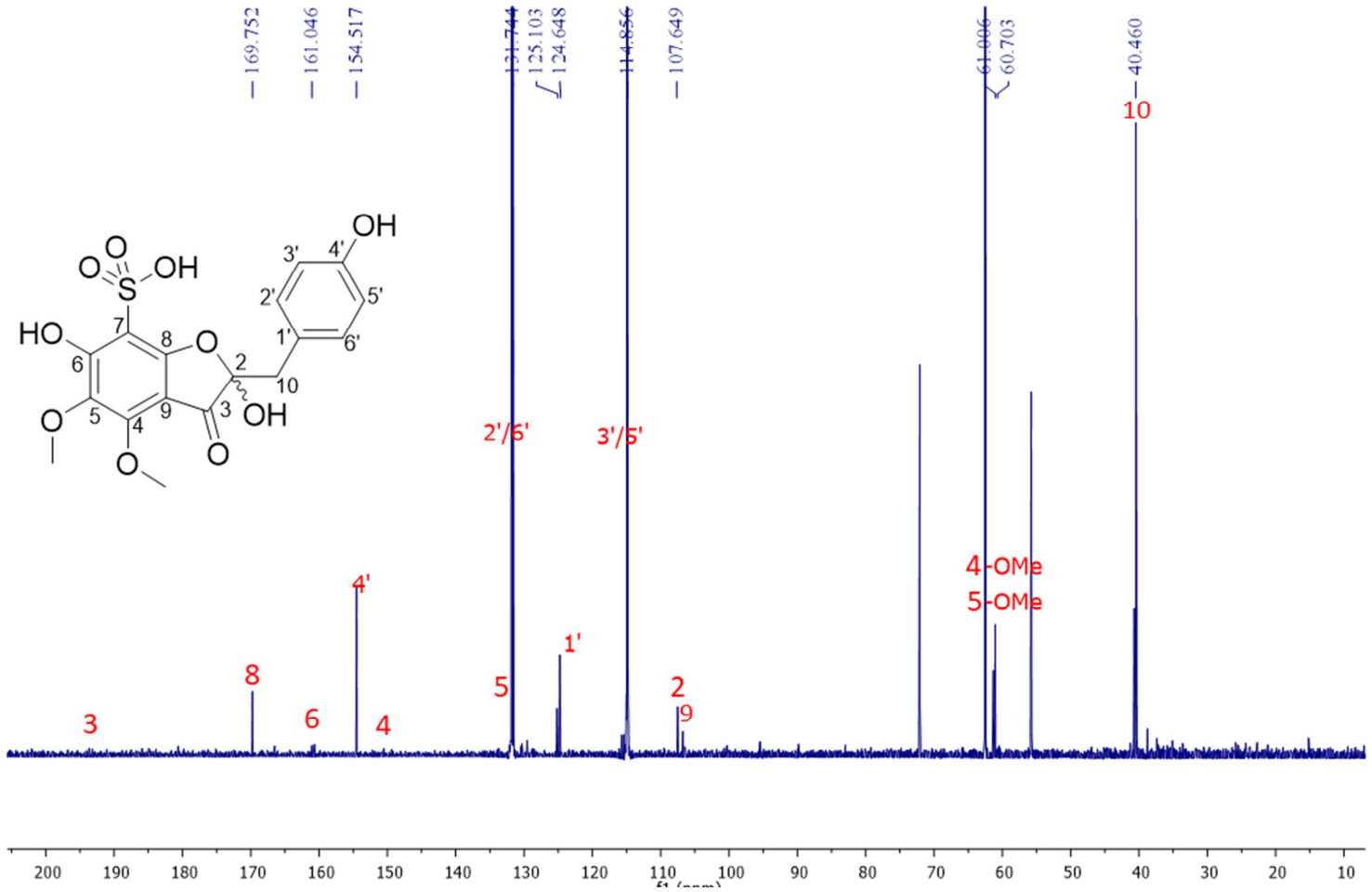
S11. HRESIMS spectrum of **3**



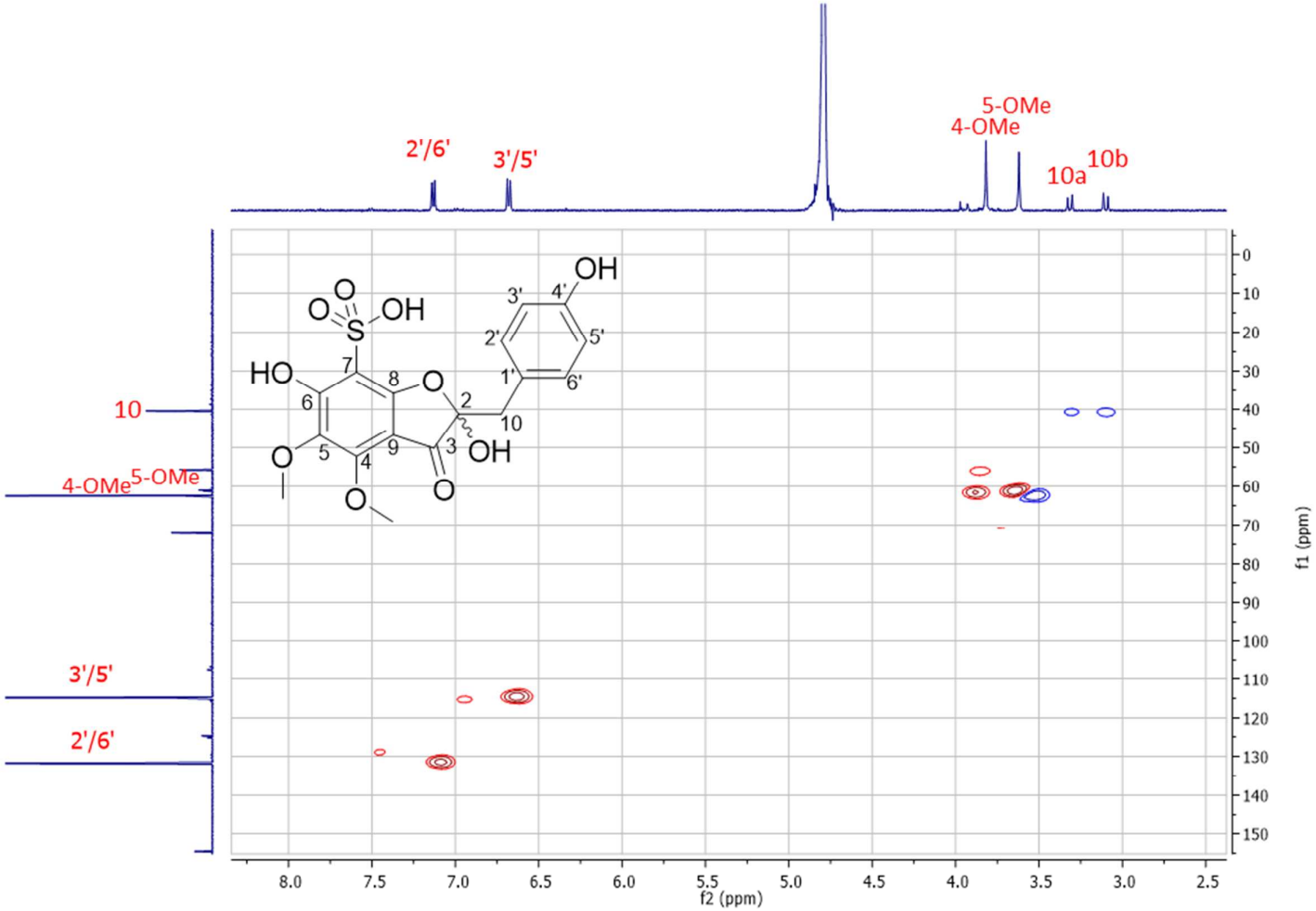
S12. ^{1}H NMR (D_2O, 500 MHz) spectrum of **3**



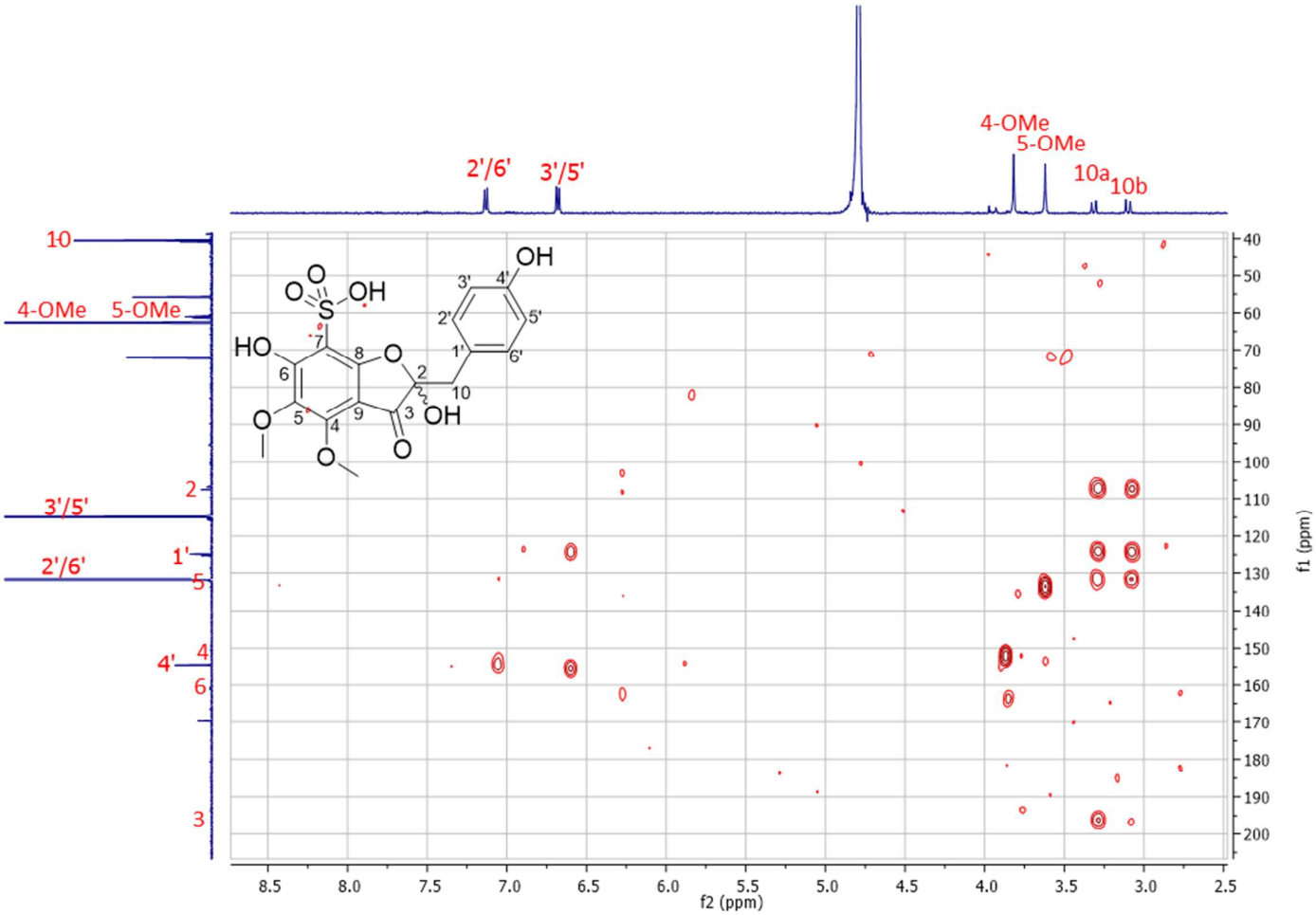
S13. ^{13}C NMR (D_2O, 125 MHz) spectrum of **3**



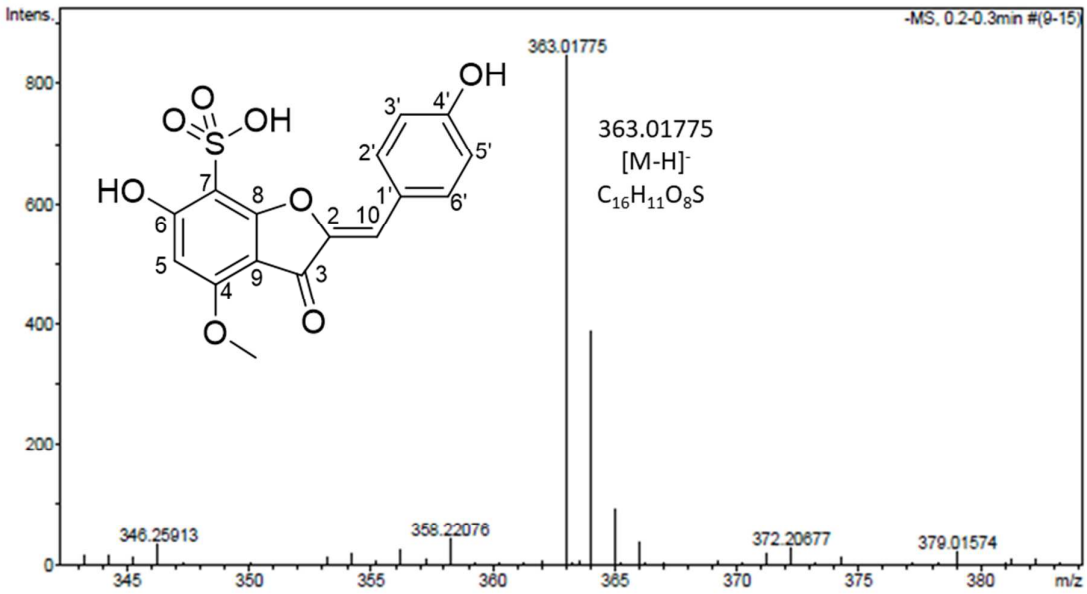
S14. HSQC (D_2O, 500 MHz, 125 MHz) spectrum of **3**



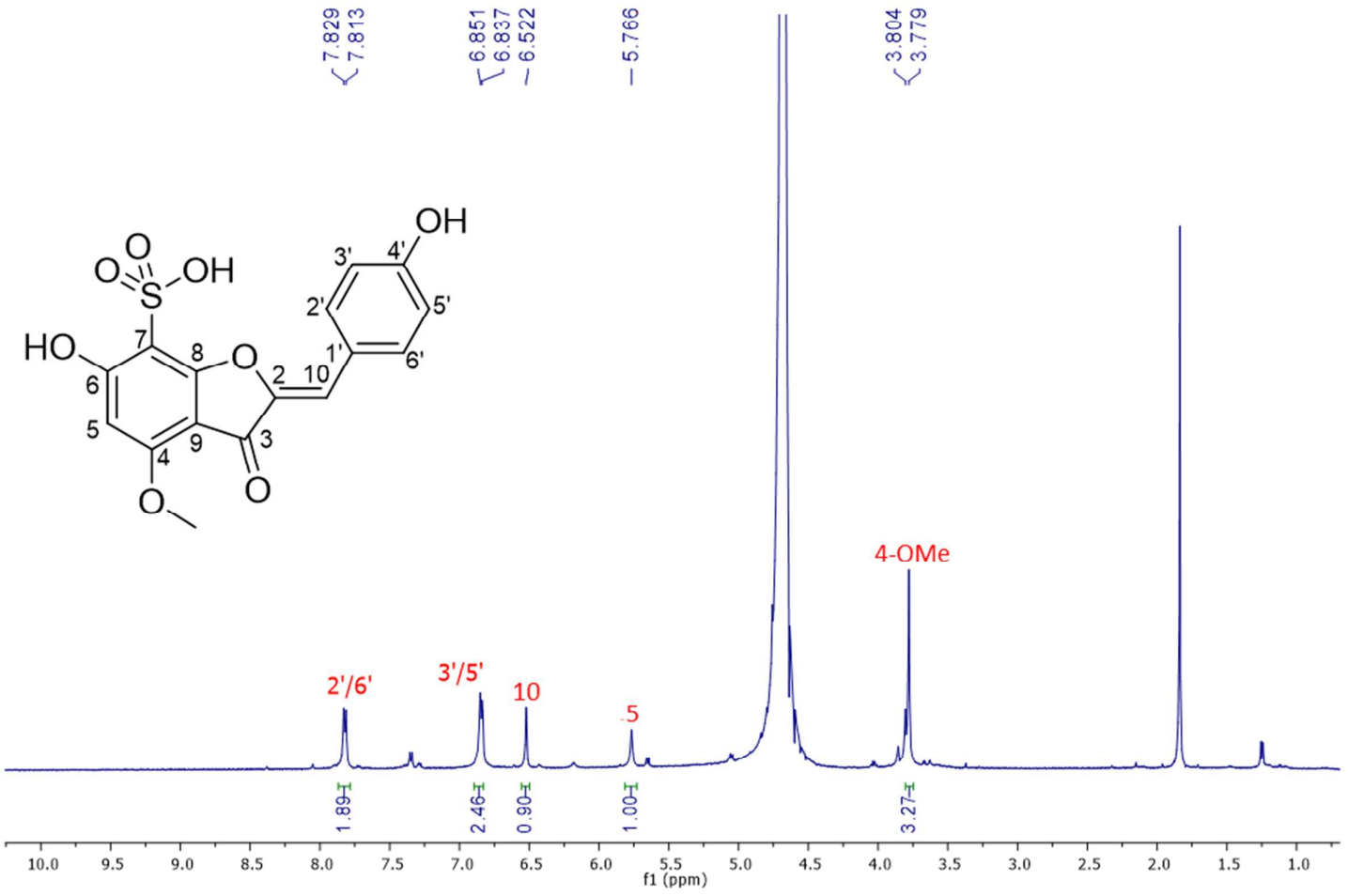
S15. HMBC (D_2O, 500 MHz, 125 MHz) spectrum of **3**



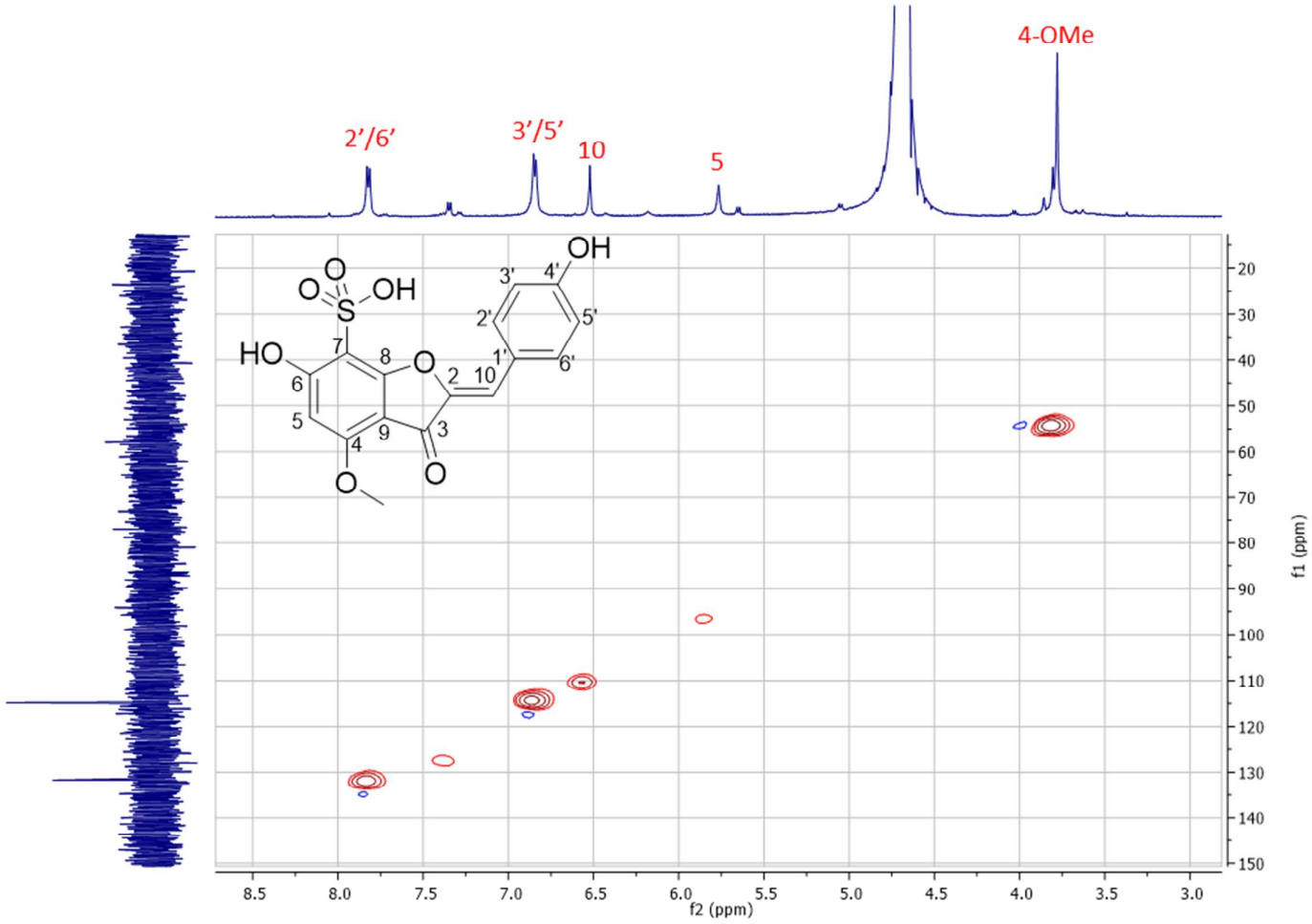
S16. HRESIMS spectrum of **4**



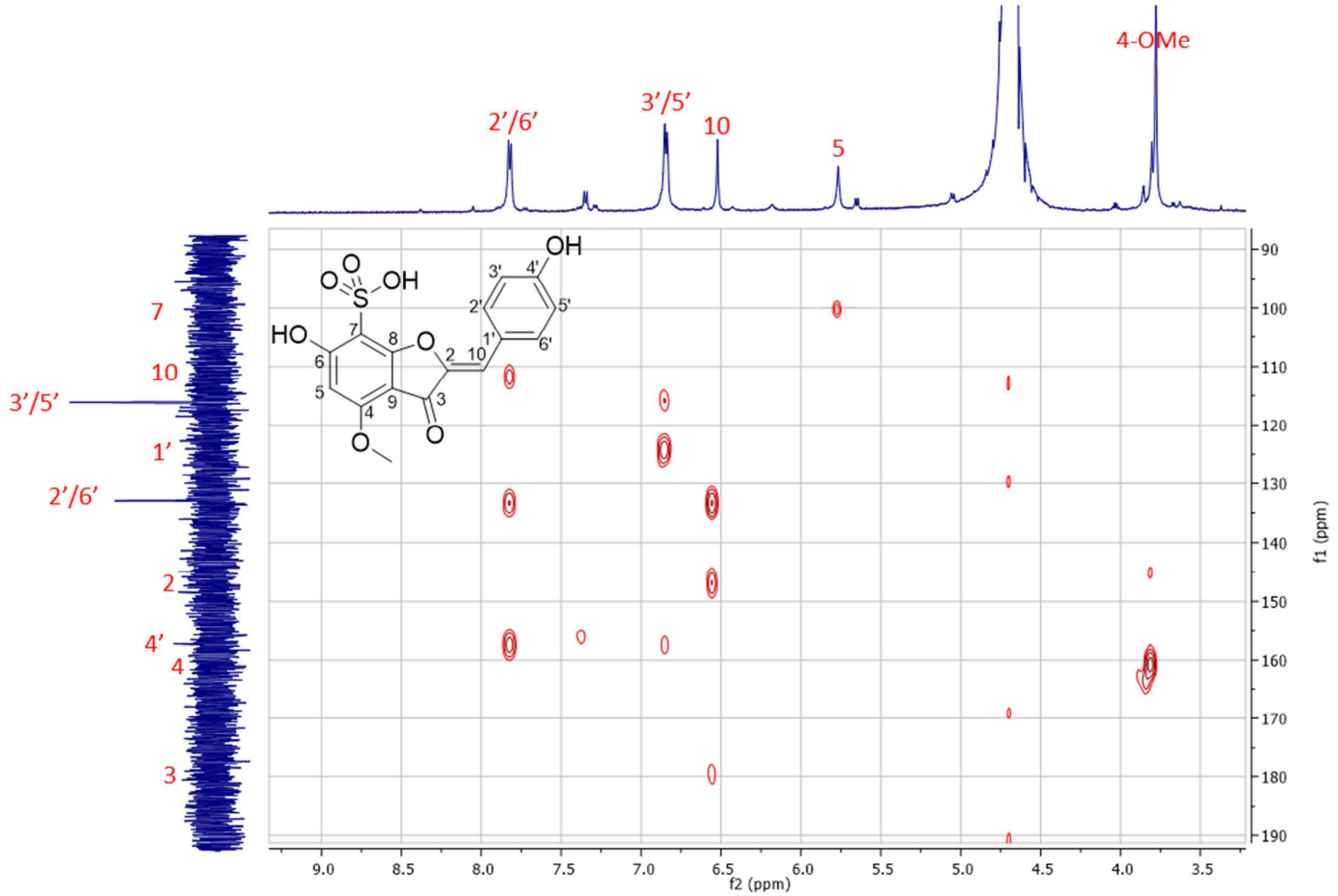
S17. ^{1}H NMR (D_2O, 500 MHz) spectrum of **4**



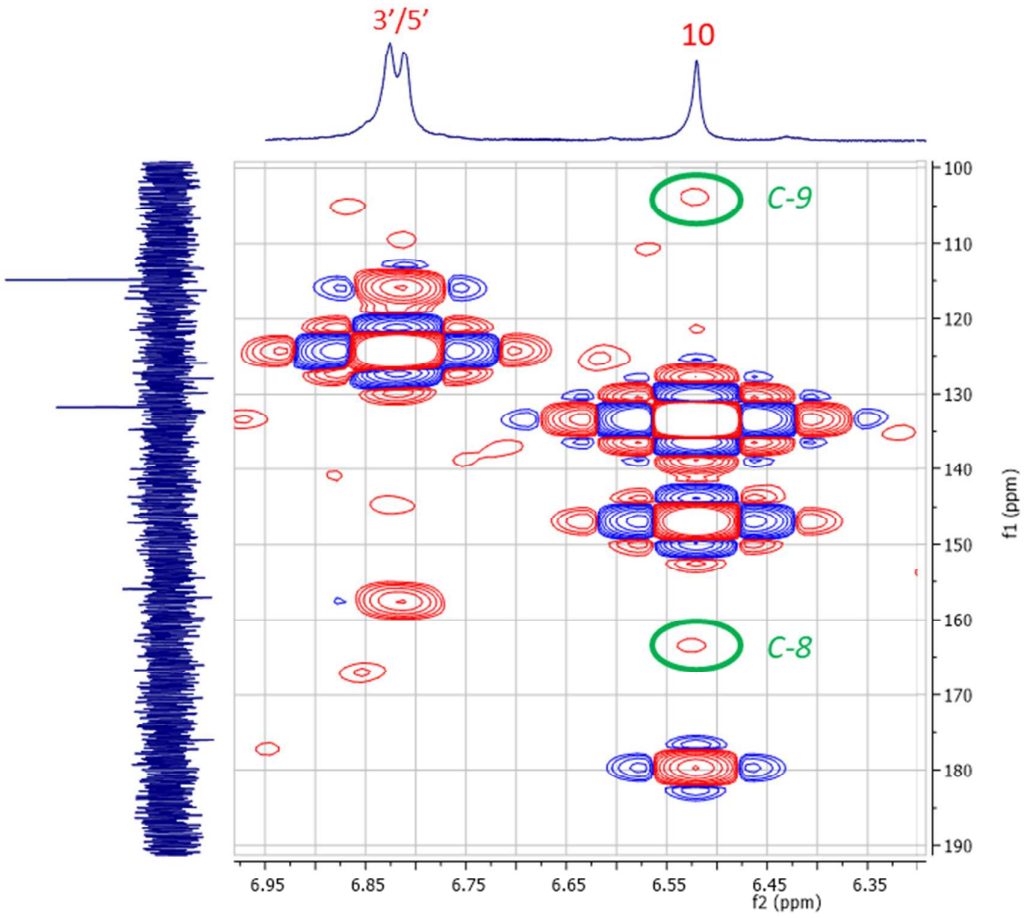
S18. HSQC (D_2O, 500 MHz, 125 MHz) spectrum of **4**

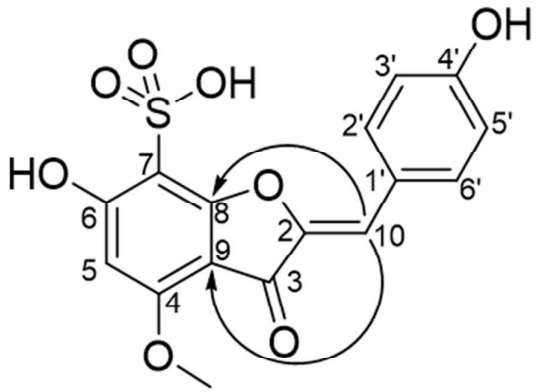


S19. HMBC (D_2O, 500 MHz, 125 MHz) spectrum of **4**

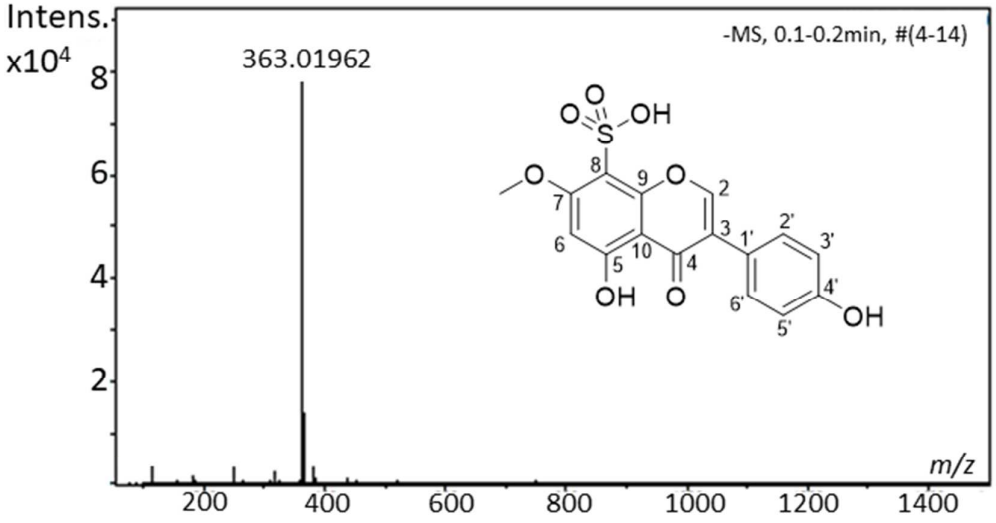


S20. Zoom into HMBC spectrum of **4** to reveal the ^{4}J correlations of olefinic proton H-10.

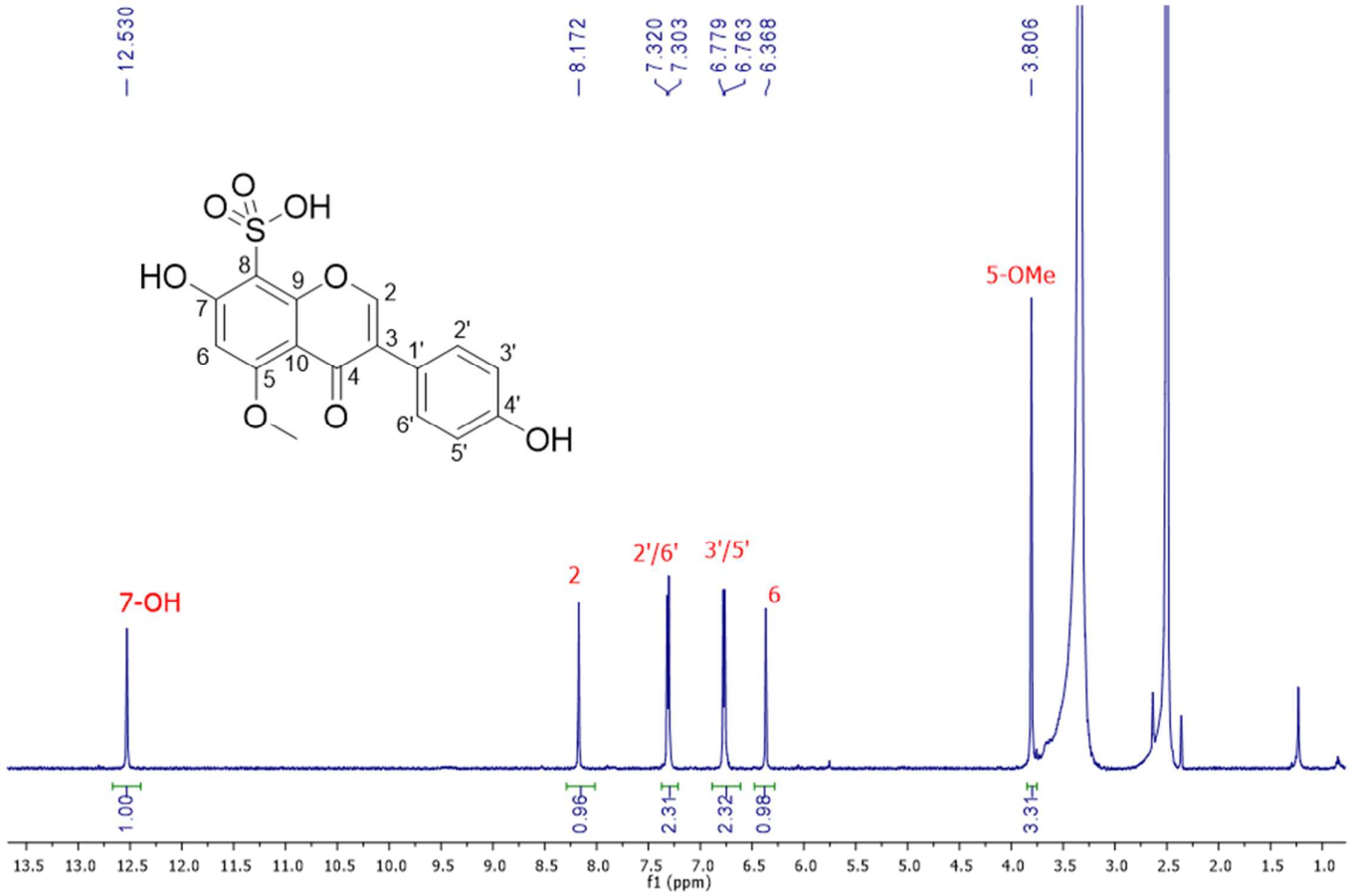




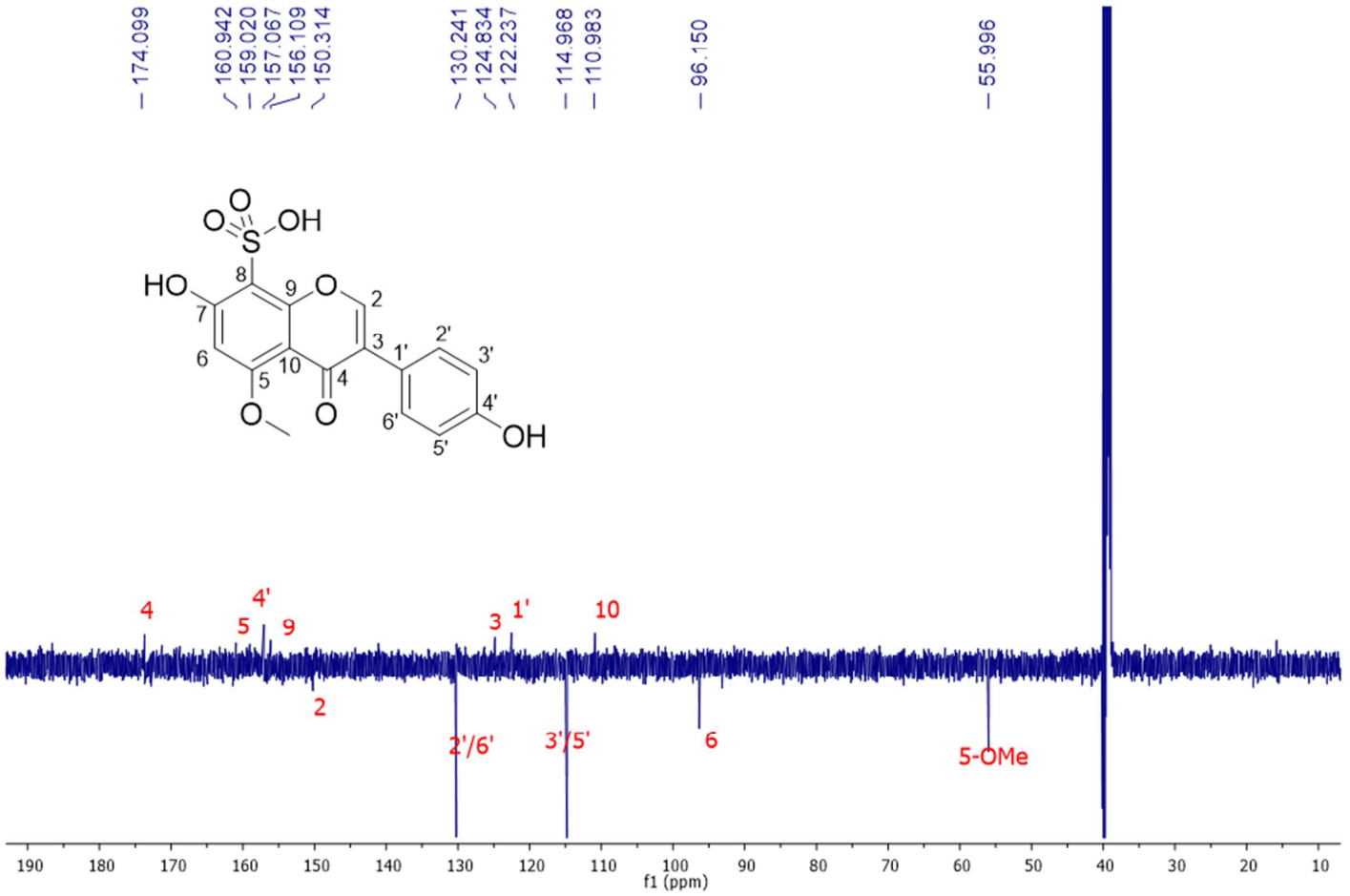
S21. HRESIMS spectrum of **5**



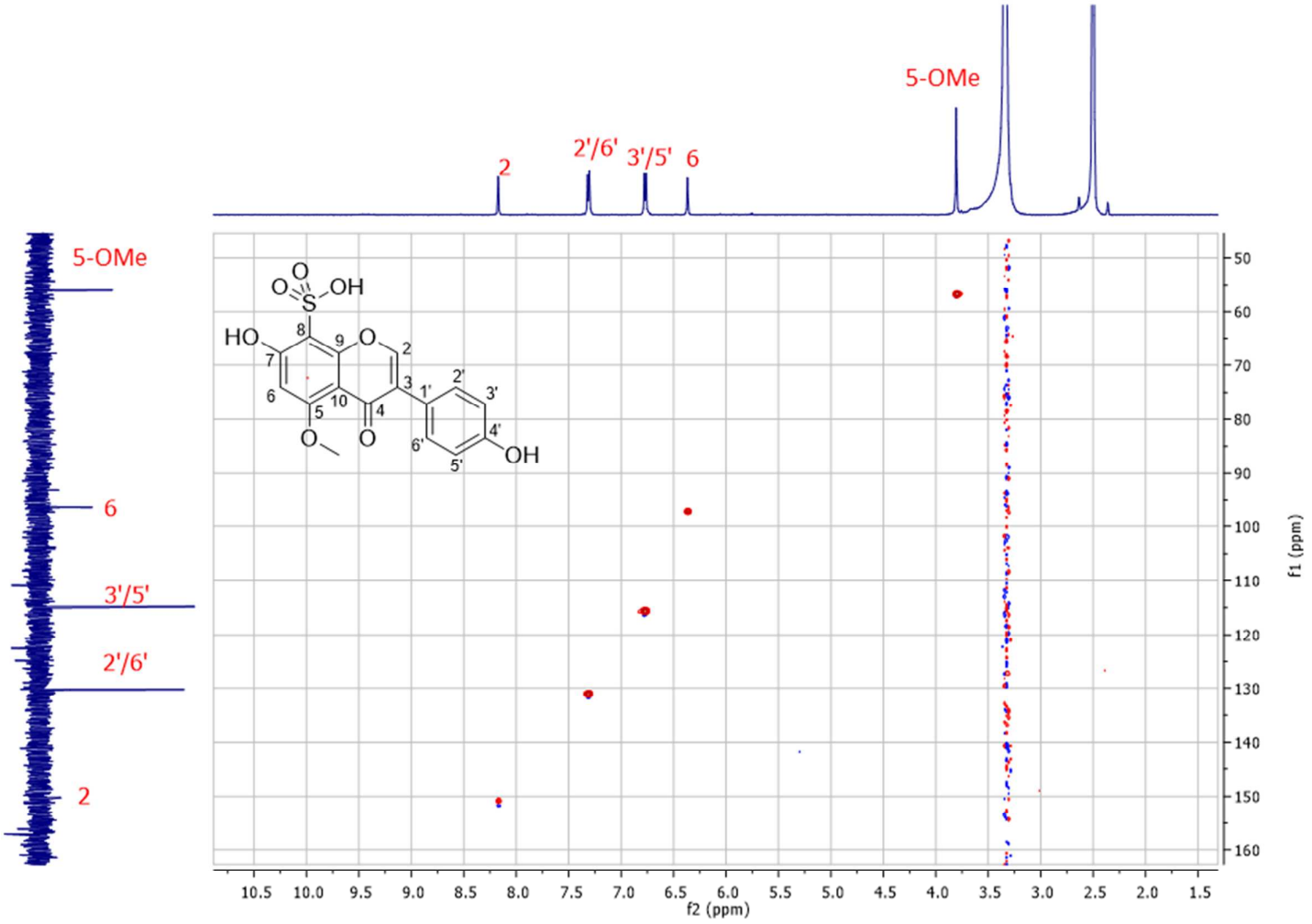
S22. ^{1}H NMR (DMSO-d_6, 500 MHz) spectrum of **5**



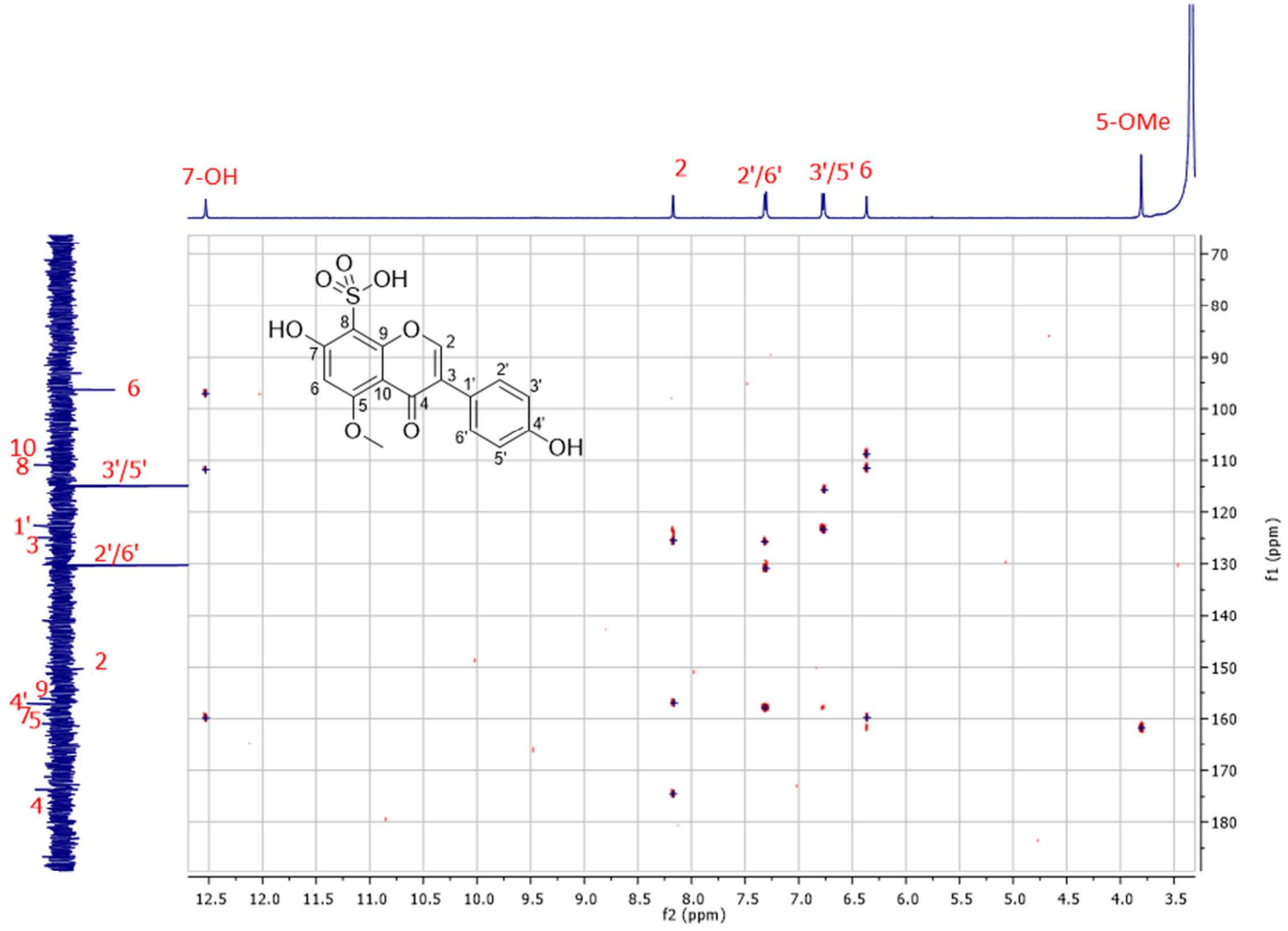
S23. JMOD (DMSO-d_6, 500 MHz) spectrum of **5**



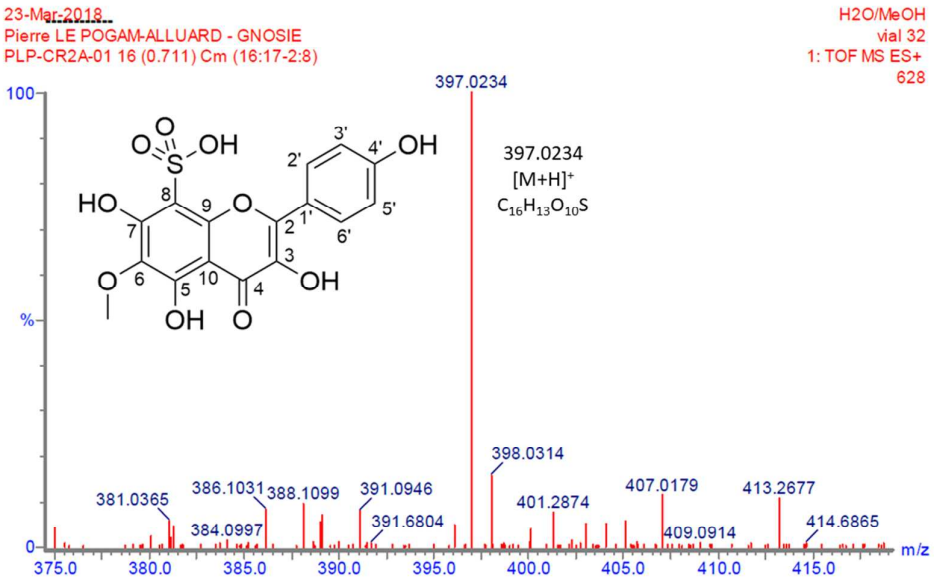
S24. HSQC (DMSO-d_6, 500 MHz, 125 MHz) spectrum of **5**



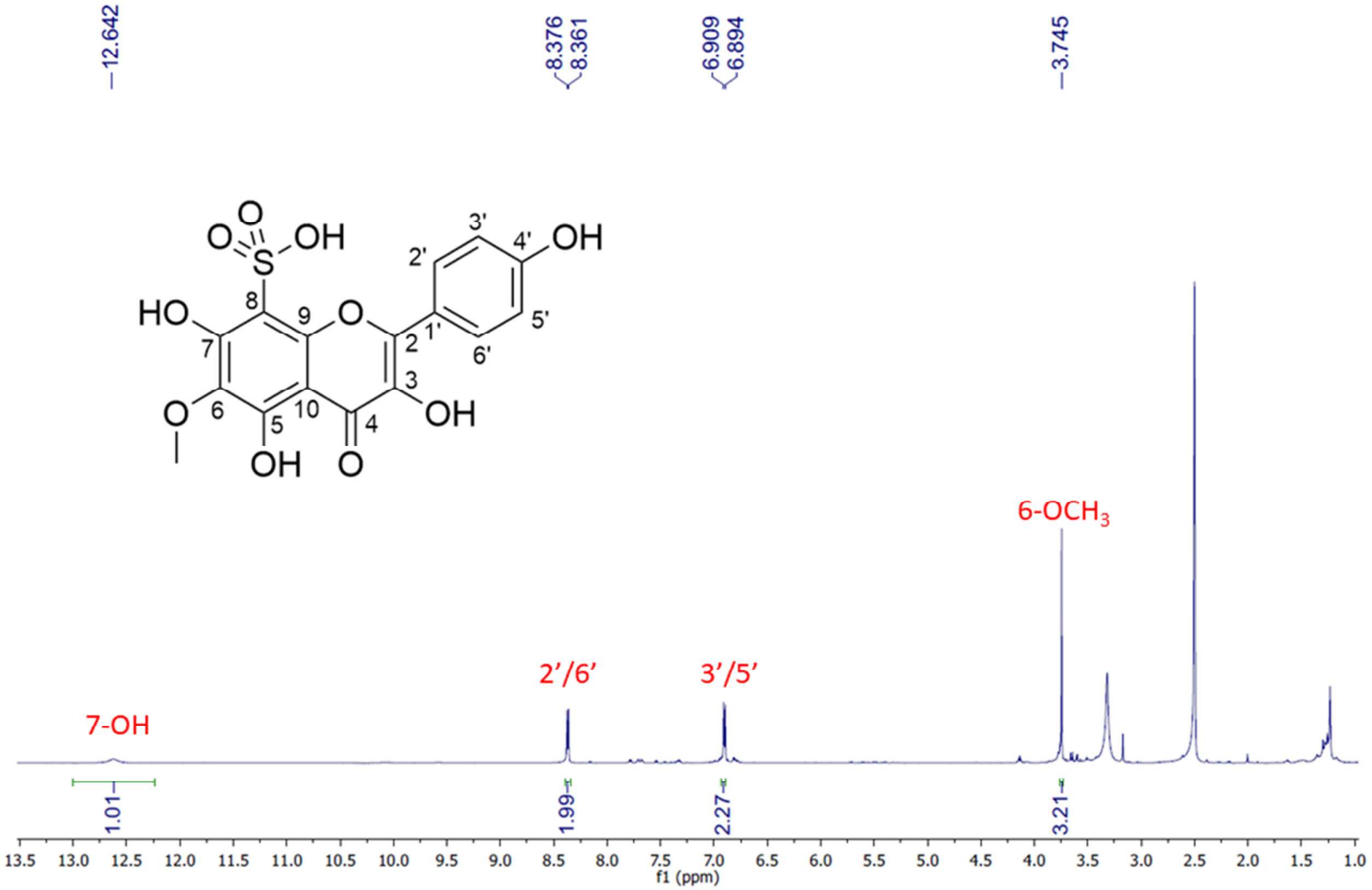
S25. HMBC (DMSO-d_6, 500 MHz, 125 MHz) spectrum of **5**



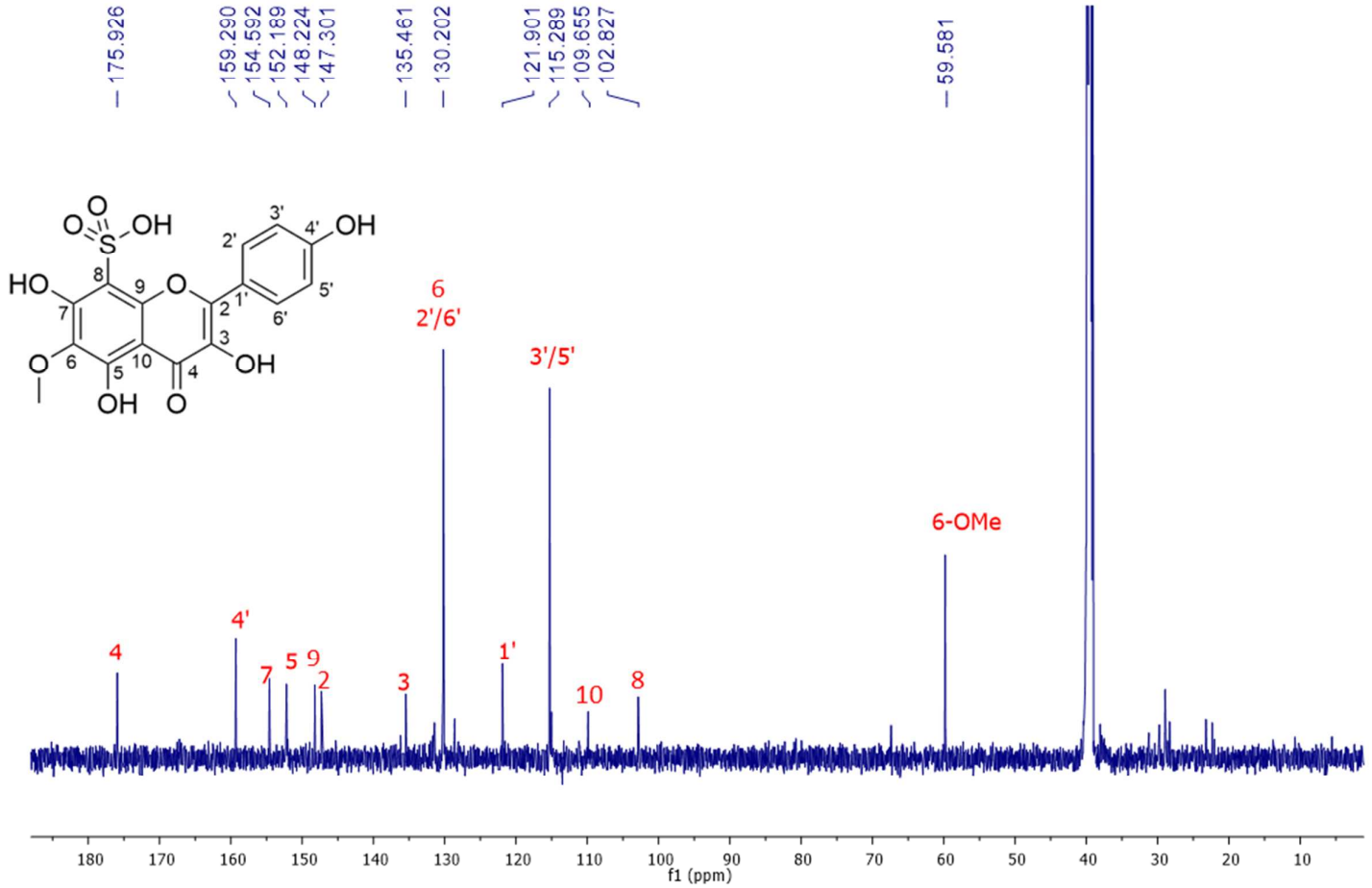
S26. HRESIMS spectrum of **6**



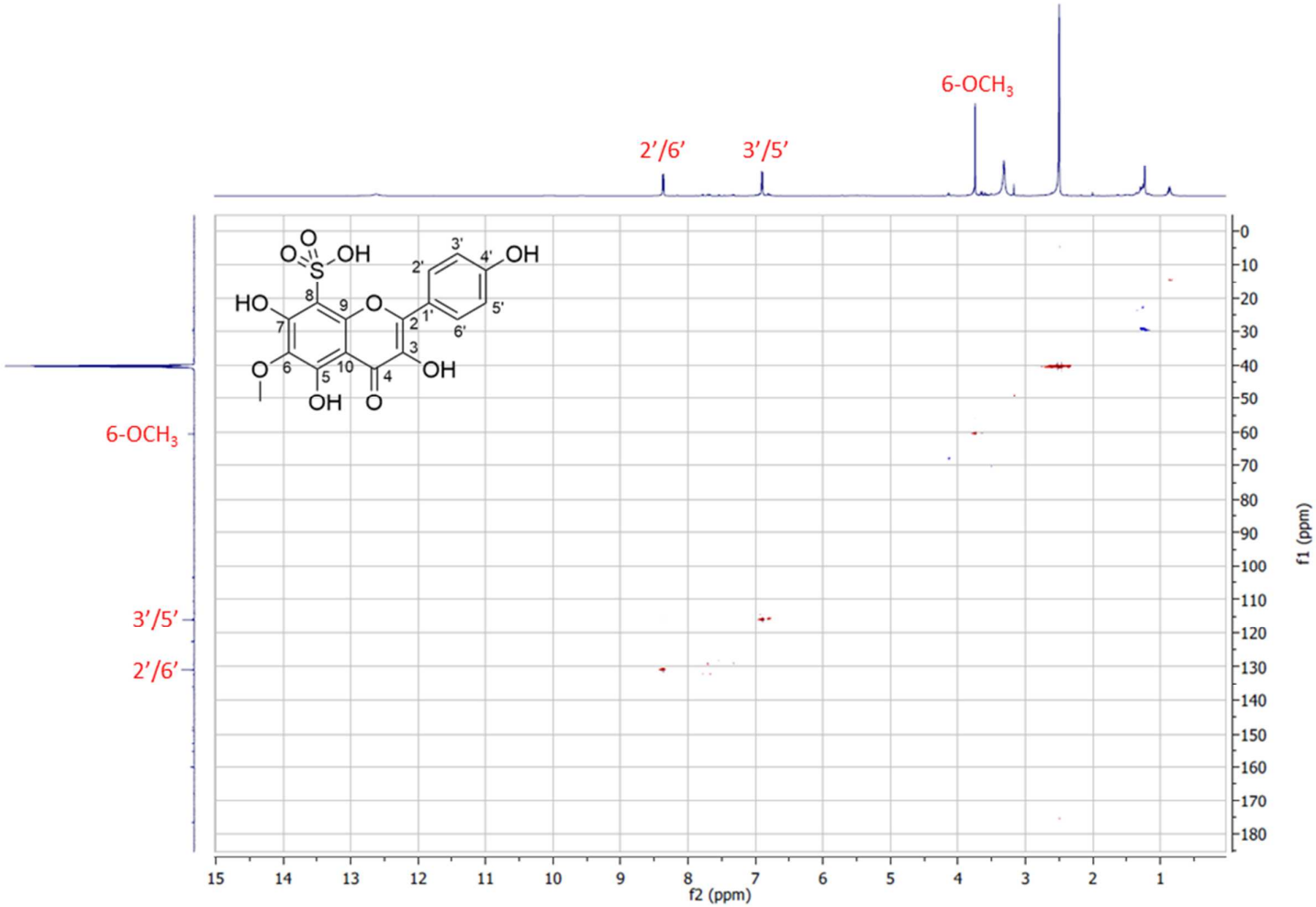
S27. ^{1}H NMR (DMSO-d_6, 600 MHz) spectrum of **6**



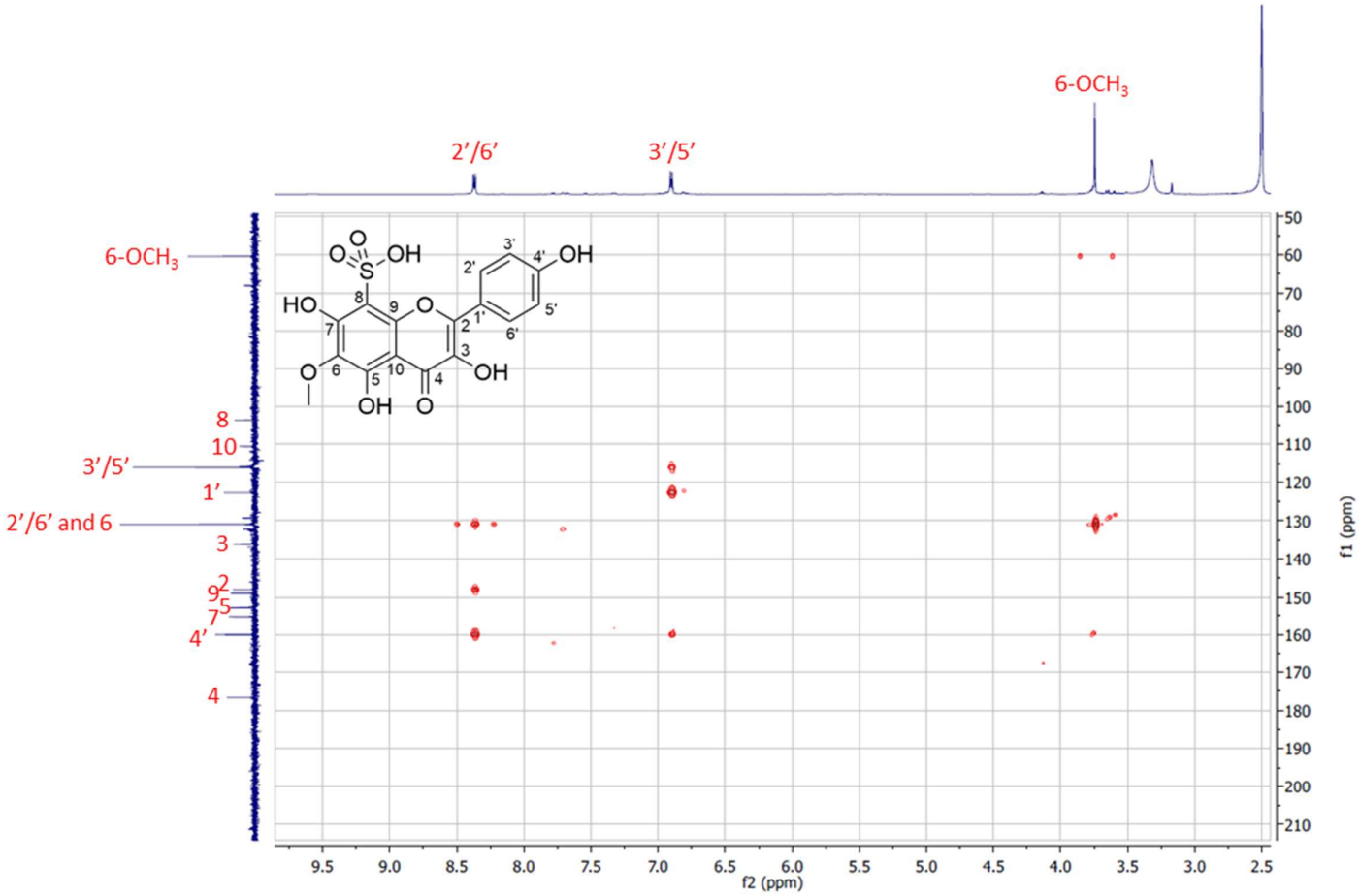
S28. ^{13}C NMR (DMSO-d_6, 150 MHz) spectrum of **6**



S29. HSQC (DMSO-d_6, 600 MHz, 150 MHz) spectrum of **6**



S30. HMBC (DMSO-d_6, 600 MHz, 150 MHz) spectrum of **6**



S31. Experimental details for X-ray diffraction analyses of **1**

(±) Acidoflavanone (**1**) crystallized as $NaC_{16}H_{13}O_9S$, in the monoclinic system, centrosymmetric space group $P2_1/c$ (no. 14) with unit cell constants a = 8.9866(4) Å, b = 10.5769(5) Å, c = 17.8065(8) Å, β = 101.942(2)°, V = 1655.9(1) Å^3, Z = 4, D_{calc} = 1.622 g cm^{-3}, MW= 404.31. A colorless rod-like single crystal of **1** with dimensions 0.22×0.32×0.36 mm was mounted on a MiTeGen micromount using epoxy glue. With the help of APEX2 Software Suite, a total of 21,429 reflections were collected at 296(2) K on a Bruker X8 APEXII Kappa CCD area-detector diffractometer with Mo Kα radiation (λ = 0.71073 Å). Data were integrated, reduced by SAINT+, corrected for absorption effects, scaled by SADABS, and merged by XPREP (Bruker AXS Inc.), yielding 5059 unique reflections (R_{int} = 0.0338). The structure was solved by intrinsic phasing method using SHELXTL XT (Bruker AXS Inc.) and refined by full-matrix least squares on F^2 with SHELXTL XLMP (Bruker AXS Inc.); final R_1 = 0.0486, wR_2 = 0.1286, $GooF$ = 1.057 for 3782 reflections with $F^2 > 2\sigma(F^2)$. Crystallographic data for the structure of **1** have been deposited within the Cambridge Crystallographic Data Center as supplementary publication (CCDC 1836646). Copies of these data can be obtained free of charge via www.ccdc.cam.ac.uk/conts.retrieving.html (or from the Cambridge Crystallographic Data Center, 12 Union Road, Cambridge CB21EZ, UK; fax: (+44) 1223-336-033; or deposit@ccdc.cam.ac.uk.