

Supporting Information

“Dissecting substrate specificities of the mitochondrial AFG3L2 protease”

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Figure S1.

Figure S2.

Figure S3.

Figure S4.

Figure S5.

Table S1.

Supporting Information References.

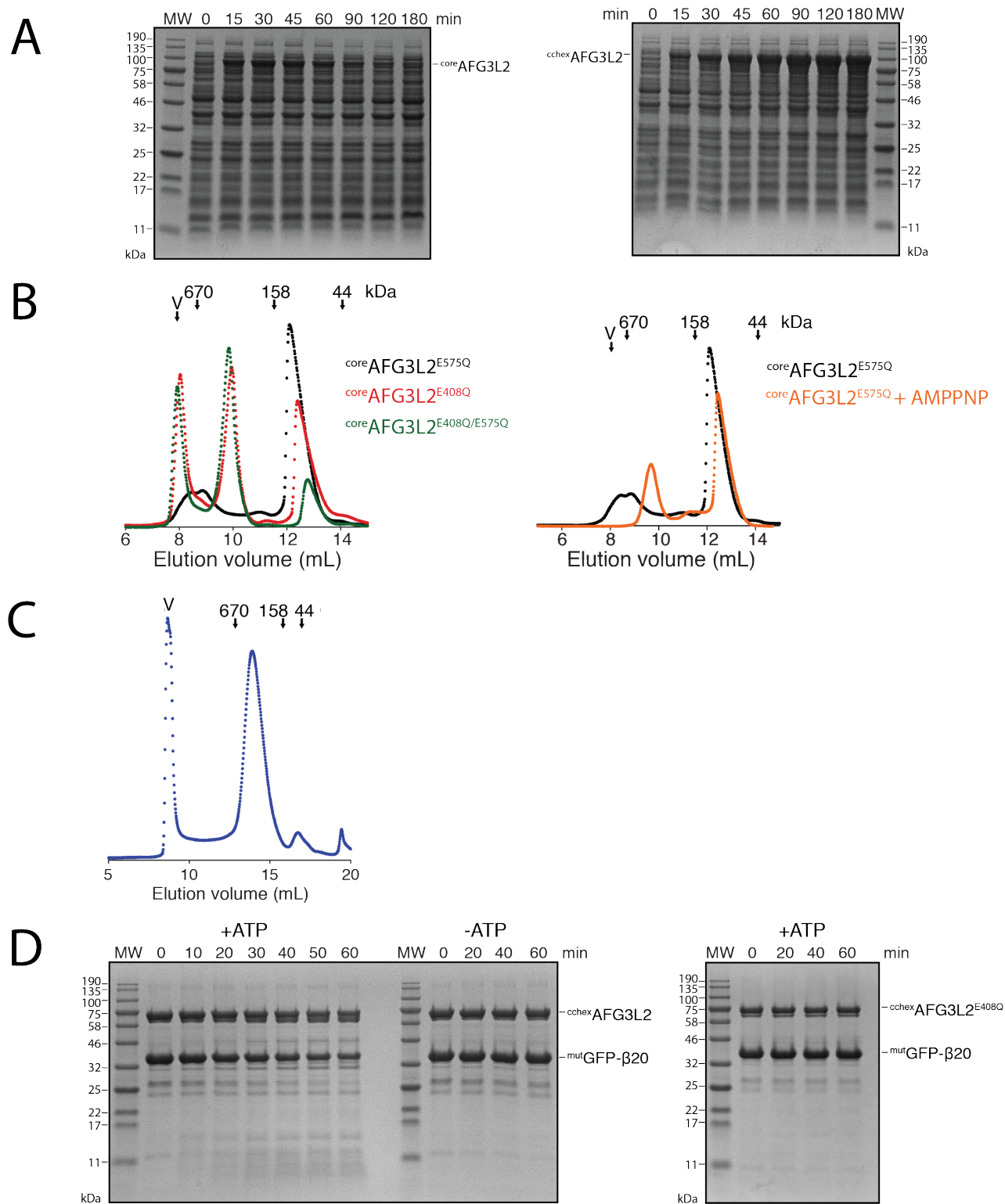
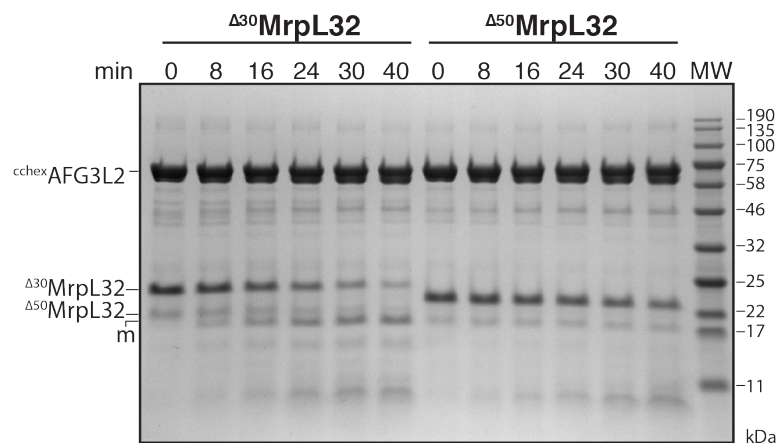
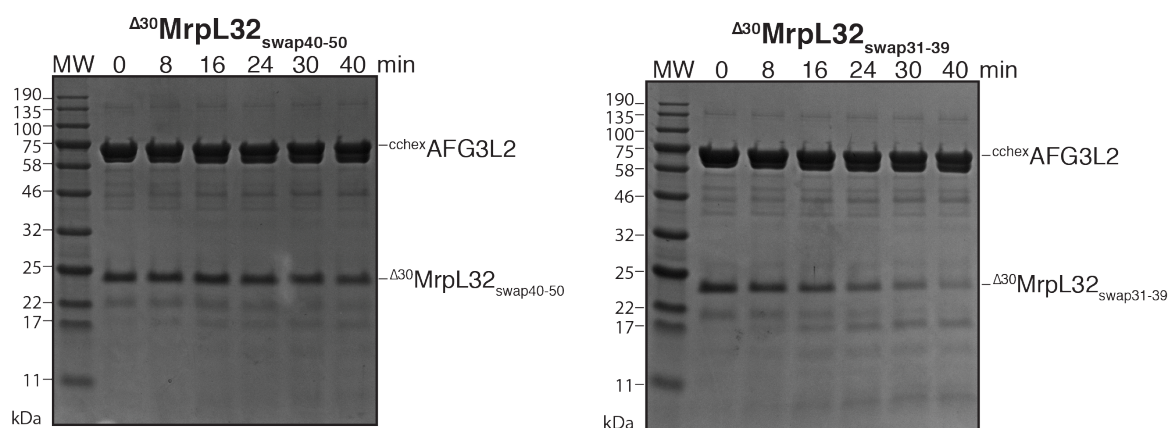


Figure S1. (A) Full uncropped SDS-PAGE gels showing expression of core AFG3L2 and cchex AFG3L2. Levels of core AFG3L2 increase after induction followed by a steady decrease, whereas levels of cchex AFG3L2 do not decrease after induction. (B) Size exclusion chromatography profiles of core AFG3L2 constructs. core AFG3L2 E408Q and core AFG3L2 $^{E408Q/E575Q}$ migrate largely as hexamers whereas core AFG3L2 E575Q migrates largely as unassembled protomers. Incubation of core AFG3L2 E575Q with 30 μ M AMPPNP shifts the migration profile towards the hexameric species. (C) Size exclusion chromatography profile of cchex AFG3L2 showing the protein largely migrates as a hexamer (D) Full uncropped representative SDS-PAGE gels from Fig. 1C. showing degradation of mut GFP- β 20 by cchex AFG3L2 in the presence and absence of ATP and by cchex AFG3L2 E408Q in the presence of ATP.

A



B



C

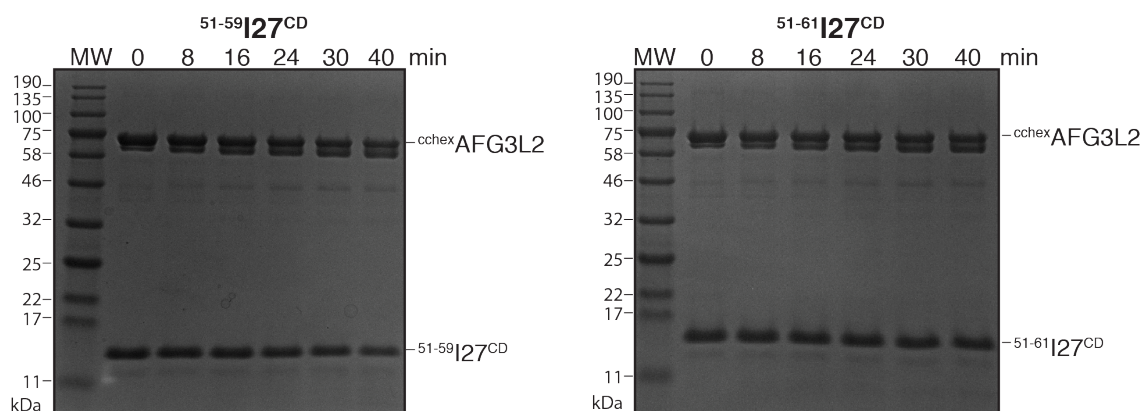


Figure S2. (A) Full uncropped representative SDS-PAGE gel showing proteolysis of Δ^{30} MrpL32 and Δ^{50} MrpL32 by $cchex$ AFG3L2 in the presence of ATP. Loss of Δ^{30} MrpL32 occurs rapidly and occurs concurrently with accumulation of a smaller molecular weight fragment (m). (B) Full uncropped representative SDS-PAGE gels showing proteolysis of Δ^{30} MrpL32_{swap40-50} and Δ^{30} MrpL32_{swap31-39} by $cchex$ AFG3L2. (C) Full uncropped representative SDS-PAGE gels showing proteolysis of $^{51-59}$ I27^{CD} and $^{51-61}$ I27^{CD} by $cchex$ AFG3L2.

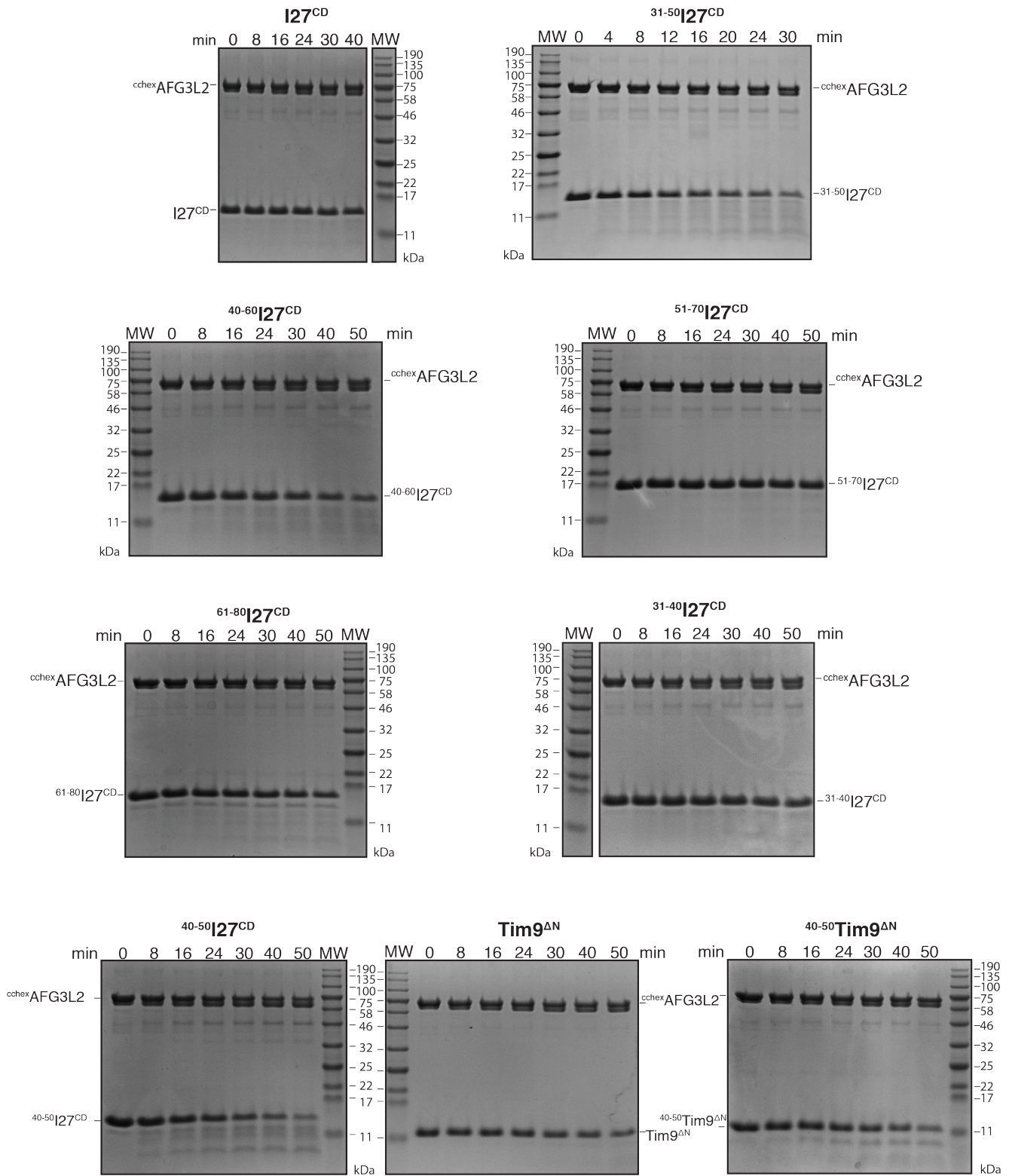


Figure S3. Full uncropped representative SDS-PAGE gels showing degradation of I27^{CD} and Tim9^{ΔN} constructs bearing sequences derived from MrpL32 at the N-terminus.

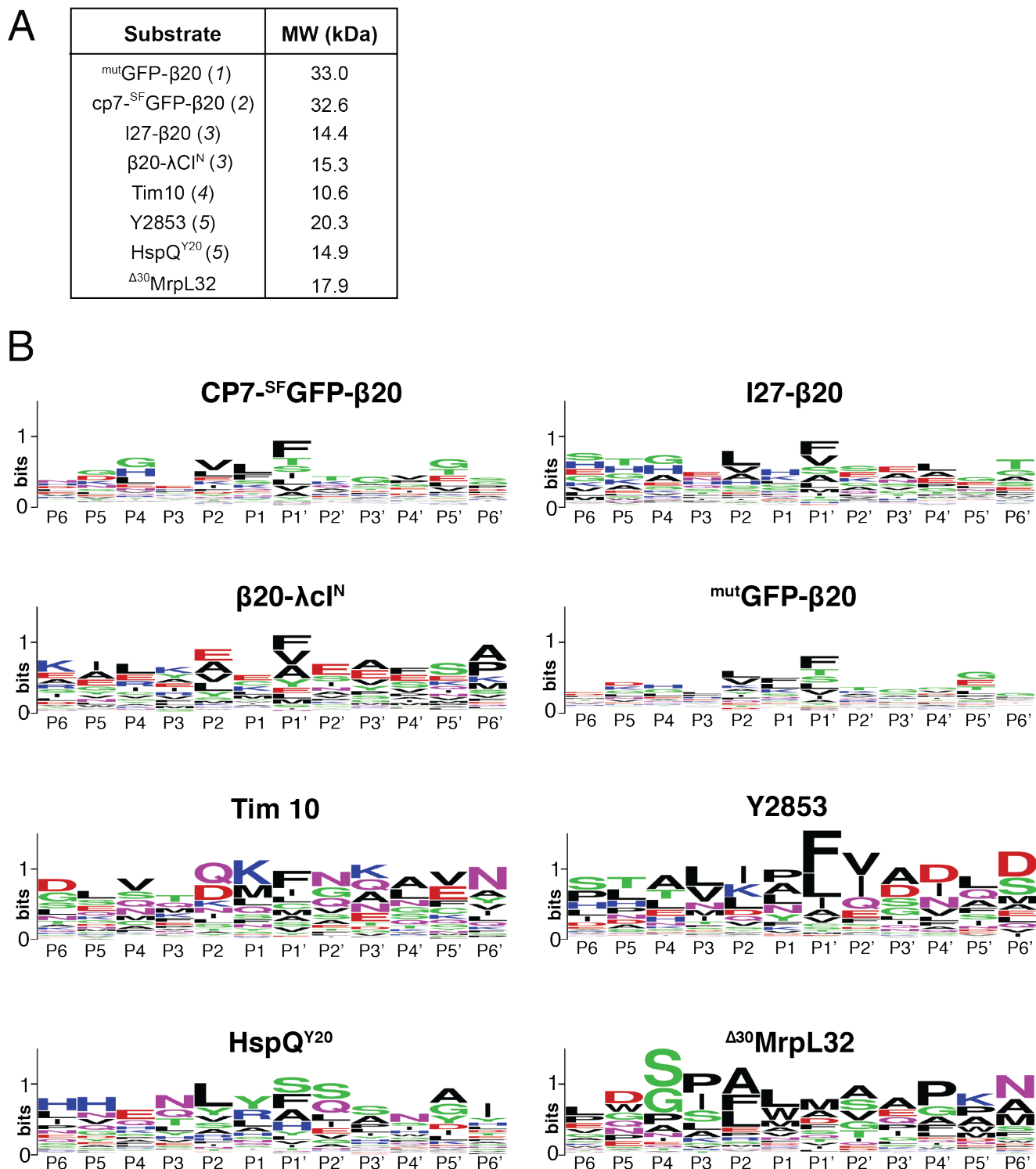


Figure S4. (A) Eight substrates used to generate the AFG3L2 peptidase specificity profile by degradation followed by LC/MS/MS. References for each substrate are given in parantheses. (B) Sequence logos showing individual peptidase specificity profiles from eight protein substrates as determined by LC/MS/MS.

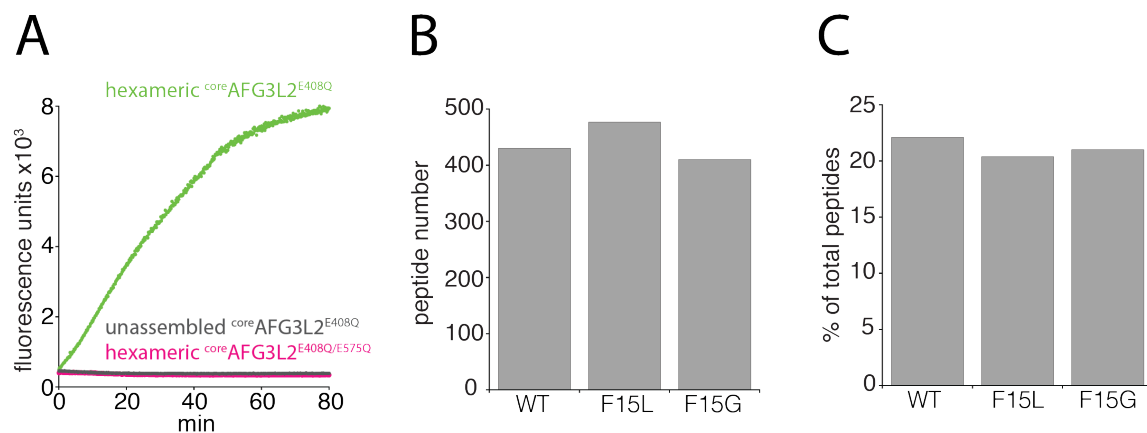


Figure S5. (A) Plot showing rapid cleavage of the reporter peptide Leu-(3-NO₂-Tyr)-Phe-Gln-(Lys-Abz) by hexameric *coreAFG3L2*^{E408Q}, whereas unassembled *coreAFG3L2*^{E408Q} does not cleave the peptide comparable to hexameric *coreAFG3L2*^{E408Q/575Q}. (B) Plot showing total number of peptides identified by LC/MS/MS resulting from cleavage from HspQ^{Y20} and HspQ^{Y20} variants bearing substitutions of Phe15 to Leu or Gly. (C) Plot showing percentage of total peptides identified by LC/MS/MS resulting from cleavage between residues 65 and 66 of HspQ^{Y20} and HspQ^{Y20} variants bearing substitutions of Phe15 to Leu or Gly.

Protein Substrates	
Substrate	Initial Degradation Rate ($\text{min}^{-1} \text{enz}_6^{-1}$)
Δ^{30} MrpL32	0.79 ± 0.04
Δ^{50} MrpL32	0.38 ± 0.04
Δ^{30} MrpL32 _{swap31-39}	0.78 ± 0.06
Δ^{30} MrpL32 _{swap40-50}	0.29 ± 0.02
I27 ^{CD}	0.14 ± 0.02
³¹⁻⁵⁰ I27 ^{CD}	0.76 ± 0.10
⁴⁰⁻⁶⁰ I27 ^{CD}	0.43 ± 0.07
⁵¹⁻⁷⁰ I27 ^{CD}	0.18 ± 0.03
⁶¹⁻⁸⁰ I27 ^{CD}	0.32 ± 0.03
³¹⁻⁴⁰ I27 ^{CD}	0.24 ± 0.02
⁴⁰⁻⁵⁰ I27 ^{CD}	0.65 ± 0.01
⁵¹⁻⁶¹ I27 ^{CD}	0.18 ± 0.03
⁵¹⁻⁵⁹ I27 ^{CD}	0.33 ± 0.06
Tim9 ^{ΔN}	0.24 ± 0.04
⁴⁰⁻⁵⁰ Tim9 ^{ΔN}	0.41 ± 0.03
Reporter Peptides	
Sequence	Initial Cleavage Rate ($\text{min}^{-1} \text{enz}_6^{-1}$)
LYFQK	0.99 ± 0.05
LYLQK	0.24 ± 0.02
LYSQK	0.098 ± 0.004
LYGQK	-0.0021 ± 0.002

Table S1. Initial rates for all degradation assays $\pm$ s.d.

Supporting Information References

- [1] Iosefson, O., Nager, A. R., Baker, T. A., and Sauer, R. T. (2015) Coordinated gripping of substrate by subunits of a AAA+ proteolytic machine, *Nat Chem Biol* 11, 201-206.
- [2] Wohlever, M. L., Nager, A. R., Baker, T. A., and Sauer, R. T. (2013) Engineering fluorescent protein substrates for the AAA+ Lon protease, *Protein engineering, design & selection : PEDS* 26, 299-305.
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- [4] Rampello, A. J., and Glynn, S. E. (2017) Identification of a Degradation Signal Sequence within Substrates of the Mitochondrial i-AAA Protease, *J Mol Biol* 429, 873-885.
- [5] Puri, N., and Karzai, A. W. (2017) HspQ Functions as a Unique Specificity-Enhancing Factor for the AAA+ Lon Protease, *Molecular cell* 66, 672-683 e674.