

## Supporting Information

### Engineering Multifunctional Coatings on Nanoparticles Based on Oxidative

### Coupling Assembly of Polyphenols for Stimuli-Responsive Drug Delivery

*Hongshan Liang<sup>a,c</sup>, Bin Zhou<sup>b</sup>, Jing Li<sup>a,c</sup>, Xingnian Liu<sup>a,c</sup>, Ziyu Deng<sup>a,c</sup>, Bin Li<sup>a,c,d\*</sup>*

<sup>a</sup> College of Food Science and Technology, Huazhong Agricultural University, Wuhan 430070, China

<sup>b</sup> School of Food and Biological Engineering, Hubei University of Technology, Wuhan 430068, China

<sup>c</sup> Key Laboratory of Environment Correlative Dietology (Huazhong Agricultural University), Ministry of Education, China

<sup>d</sup> Functional Food Engineering & Technology Research Center of Hubei Province, China

\*Corresponding author: Bin Li

E-mail address: libinfood@mail.hzau.edu.cn

## **Experimental**

### **1. Materials**

Zein (Z0001) was purchased from Tokyo Chemistry Industry, Co., Ltd. (Tokyo, Japan). Tannin acid (TA) was purchased from Aladdin Chemistry Co., Ltd. (-)-epigallocatechin-3-gallate (EGCG) was purchased from Xi'an Natural Field Bio-Technique Co., Ltd. (Xi'an China). Persimmon tannin (PT, 98.7% purity) was kindly provided by Shanghai Ocean University composed of polymers ranging from 7 to 20 kDa. 3-(N-morpholino)-propanesulfonic acid (MOPS) was obtained from the Sinopharm Chemical Reagents Co., Ltd. (Shanghai, China).

### **2. Preparation and characterization of polyphenol-coated zein NPs**

All solutions were freshly prepared for immediate use. The standard preparation process was described as follows: zein was dissolved in aqueous ethanol solutions (75% v/v) to obtain a stock solution with final concentration of 10 mg/mL. Then 0.5 mL zein solution was added to 9.5 mL of MOPS buffer (10 mM, pH 7.8). Next, different volume (40  $\mu$ L, 60  $\mu$ L, 80  $\mu$ L or 100  $\mu$ L) of TA solution (24 mM) was added and the dispersion was under vigorous stirring for 1 h at room temperature. The product was then purified by successive dialysis (MWCO 3500) against deionized water for 48 h to remove the free TA. Then 200  $\mu$ L of H<sub>2</sub>AuCl<sub>4</sub> aqueous solutions (10 mM) was added in the zein-TA solution to observe the metallization of gold.

**UV-vis spectroscopy:** The UV-vis absorption spectra of polyphenol-coated zein NPs solutions were measured on a spectrophotometer (UV-1100, MAPADA).

Table S1. Characterization of NPs (pH 7.4), the results were displayed as the mean  $\pm$  standard deviation (n=3).

Samples	size, nm	PDI	zeta potential, (mV)
zein	89.2 $\pm$ 1.2	0.15 $\pm$ 0.04	-25.2 $\pm$ 1.6
zein/TA <sub>1</sub>	66.9 $\pm$ 0.6	0.21 $\pm$ 0.01	-35.1 $\pm$ 0.5
zein/TA <sub>2</sub>	76.0 $\pm$ 1.5	0.22 $\pm$ 0.02	-37.9 $\pm$ 0.7
zein/TA <sub>3</sub>	85.4 $\pm$ 1.4	0.25 $\pm$ 0.01	-38.6 $\pm$ 0.3
zein/TA <sub>4</sub>	85.2 $\pm$ 1.8	0.24 $\pm$ 0.02	-40.4 $\pm$ 1.6

zein represented zein nanoparticles; zein/TA<sub>1</sub>, zein/TA<sub>2</sub>, zein/TA<sub>3</sub> and zein/TA<sub>4</sub> represented TA-coated zein nanoparticles with TA volume of 40, 60, 80 and 100  $\mu$ L respectively.

Table S2. IC<sub>50</sub> Values for DOX or DOX-loaded NPs against HeLa cells.

Samples	Cell	IC <sub>50</sub> (μg/mL)
DOX	HeLa	0.918±0.05
DOX-zein	HeLa	0.939±0.04
DOX-zein/TA	HeLa	1.087±0.02**
DOX- zein/EGCG	HeLa	0.941±0.02
DOX-zein/PT	HeLa	1.192±0.03**

DOX represented free doxorubicin; DOX-zein represented DOX loaded zein nanoparticles; DOX-zein/TA, DOX-zein/EGCG and DOX-zein/PT represented DOX-loaded TA/EGCG/PT-coated zein nanoparticles with TA/EGCG/PT volume of 60 μL. Data displayed as mean±SD (n=3). \*p < 0.05; \*\*p < 0.01; versus DOX group.

Table S3. Characterization of NPs or DOX-loaded NPs (pH 7.4), the results were displayed as the mean  $\pm$  standard deviation (n=3).

Samples	size, nm	PDI	zeta potential, (mV)	encapsulation efficiency (%)
zein/EGCG	67.9 $\pm$ 1.1	0.25 $\pm$ 0.02	-37.5 $\pm$ 1.6	-
zein/PT	96.4 $\pm$ 1.0	0.27 $\pm$ 0.01	-38.3 $\pm$ 0.9	-
DOX- zein/EGCG	95.3 $\pm$ 1.6	0.12 $\pm$ 0.03	-32.9 $\pm$ 0.7	85.3 $\pm$ 0.4
DOX-zein/PT	120.1 $\pm$ 3.1	0.18 $\pm$ 0.01	-32.3 $\pm$ 1.2	95.7 $\pm$ 0.6

zein/EGCG and zein/PT represented EGCG/PT-coated zein nanoparticles with EGCG/PT volume of 60  $\mu$ L. DOX-zein/EGCG and DOX-zein/PT represented DOX-loaded EGCG/PT-coated zein nanoparticles with EGCG/PT volume of 60  $\mu$ L.

Figure S1

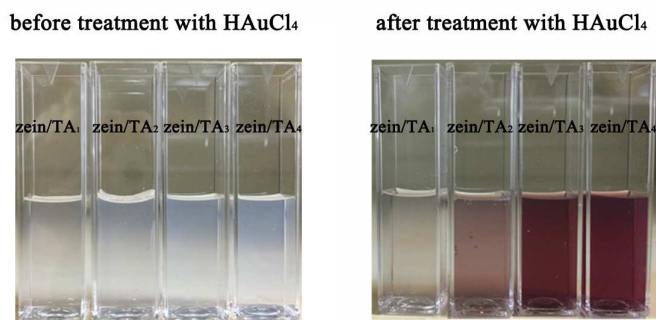


Figure S1. Photographs of zein/TA<sub>1</sub>, zein/TA<sub>2</sub>, zein/TA<sub>3</sub> and zein/TA<sub>4</sub> NPs before (left) and after (right) treatment with aqueous HAuCl<sub>4</sub>.

Figure S2

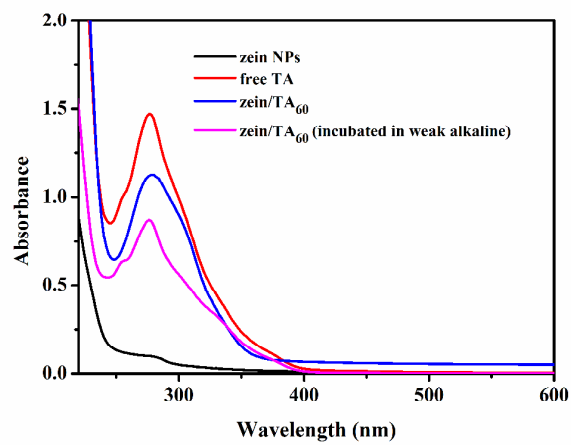


Figure S2. UV-vis spectra of zein NPs, free TA, zein/TA<sub>1</sub> NPs without incubation, zein/TA<sub>1</sub> NPs incubated in weak alkaline.

Figure S3

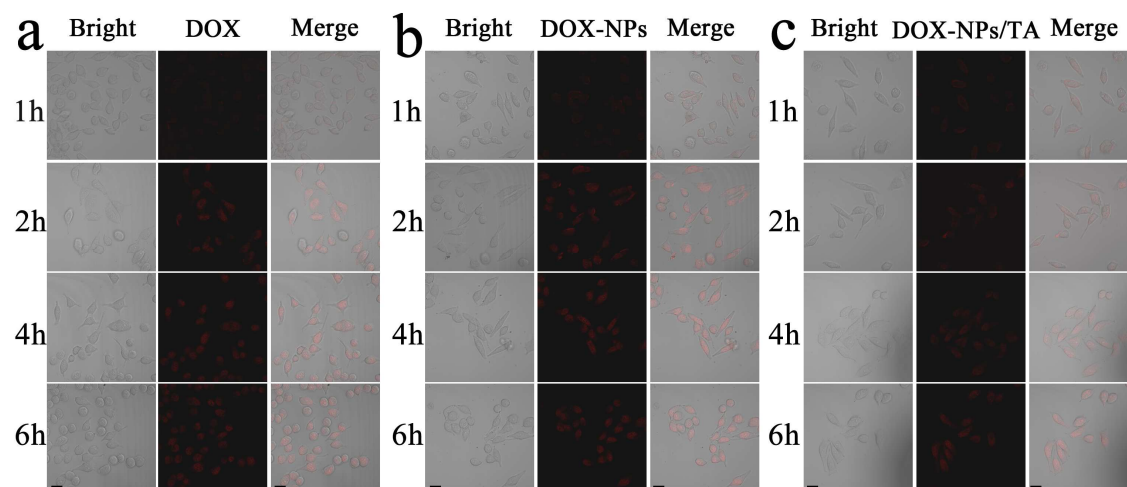


Figure S3. CLSM images of intracellular uptake of free DOX (a), DOX-loaded zein NPs (b) and DOX-loaded zein/TA NPs (c) by HeLa cells with DOX concentration of 1.0 μg/mL. The scale bars represented 20 μm.



Figure S4

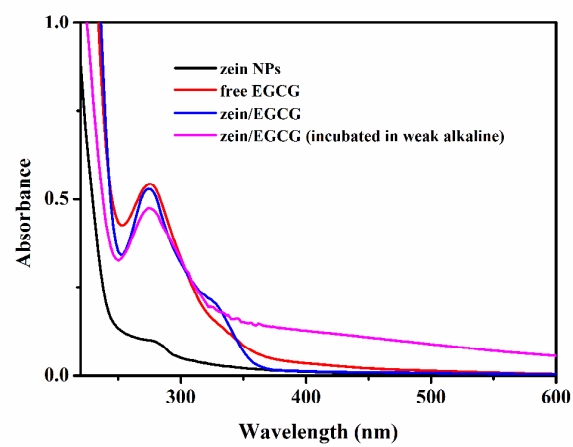


Figure S4. UV-vis spectra of zein NPs, free EGCG, zein/EGCG NPs without incubation, zein/EGCG NPs incubated in weak alkaline.

Figure S5

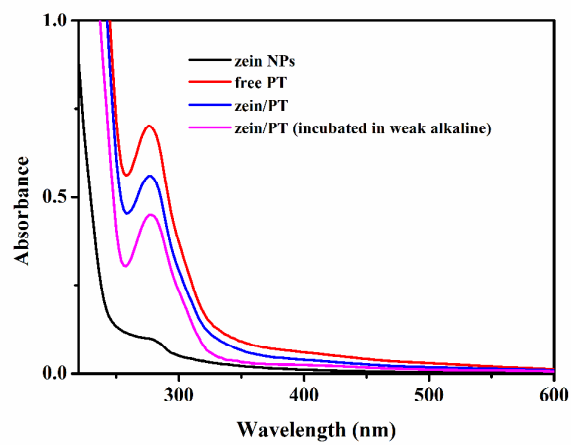


Figure S5. UV-vis spectra of zein NPs, free PT, zein/PT NPs without incubation, zein/PT NPs incubated in weak alkaline.

Figure S6

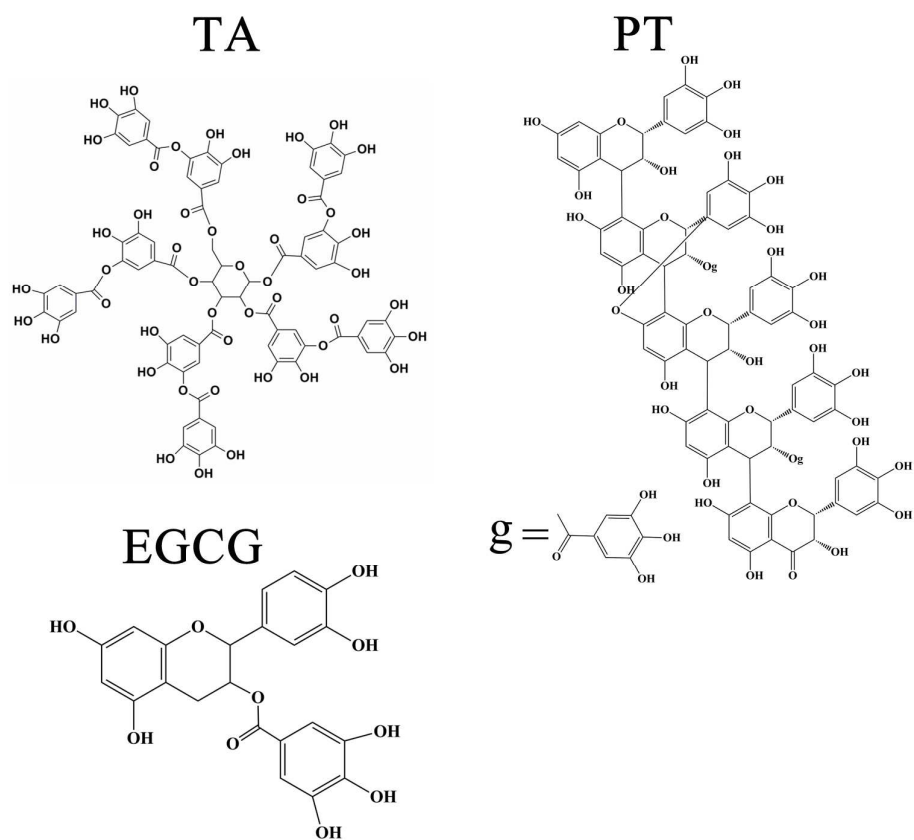


Figure S6. Chemical structure of TA, EGCG and PT.