# Synthesis and Evaluation of Fluoroalkyl Phosphonyl Analogs of 2-CMethylerythritol Phosphate as Substrates and Inhibitors of IspD from Human Pathogens 

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a)

b)







Figure S1. Regioselectivity of the acid-catalyzed ring-opening of epoxide 14. Epoxide 14 was treated with $\mathrm{H}_{2} \mathrm{SO}_{4}$ solution ( 5 M in $\mathrm{H}_{2} \mathrm{O}, \sim 98 \%{ }^{18} \mathrm{O}$ atom). a) The total ${ }^{18} \mathrm{O}$ incorporation was determined by LC-MS/MS to be $34 \%$. The diminished incorporation is likely due to ${ }^{16} \mathrm{O} /{ }^{18} \mathrm{O}$ exchange between the $\mathrm{H}_{2}{ }^{18} \mathrm{O}$ and the $\mathrm{H}_{2} \mathrm{~S}^{16} \mathrm{O}_{4}$. b) The regioselectivity of ${ }^{18} \mathrm{O}$ was determined by ${ }^{13} \mathrm{C}$ NMR. The ${ }^{18} \mathrm{O}$ induces an upfield $\Delta \delta$ of 30 ppb revealing $33 \%{ }^{18} \mathrm{O}$ incorporation at $\mathrm{C}_{2}$ and no discernible incorporation at either $\mathrm{C}_{1}$ or $\mathrm{C}_{3}$. The regioselectivity was determined to be $97 \%$ ( $94 \%$ ee) by taking the ratio of the ${ }^{18} \mathrm{O}$ incorporation at $\mathrm{C}_{2}$ and the total ${ }^{18} \mathrm{O}$ incorporation into 15.


21a


Figure S2. Full spectrum of the ${ }^{\mathbf{1}} \mathbf{H}-{ }^{\mathbf{1}} \mathbf{H}$ NOE NMR experiment with 21a. The full ${ }^{1} \mathrm{H}$ NMR spectrum of 21a can be found on page S76.



Figure S3. Full spectrum of the $\mathbf{1}^{\mathbf{1}} \mathbf{H}-\mathbf{}^{\mathbf{H}} \mathbf{H}$ NOE NMR experiment with $\mathbf{2 1 b}$. The full ${ }^{1} \mathrm{H}$ NMR spectrum of $\mathbf{2 1 b}$ can be found on page S 78 .


Figure S4. HPLC analysis of MEP turnover by E. coli IspD



Figure S5. HPLC analysis of 1 turnover by E. coli IspD


Figure S6. HPLC analysis of 2 turnover by E. coli IspD


Figure S7. HPLC analysis of 3 turnover by E. coli IspD

CTP


Figure S8. HPLC analysis of 4 turnover by E. coli IspD


Figure S9. HPLC analysis of 5a turnover by E. coli IspD


Figure S10. HPLC analysis of 5b turnover by E. coli IspD




Figure S11. LC-MS analysis of MEP turnover. Chromatograms display the total ion count (TIC) for scans containing an ion with an $m / z$ of 520 corresponding to CDPME. The mass spectrum is the scan at the center of the peak in the 30 min sample.



Figure S12. LC-MS analysis of $\mathbf{1}$ turnover. Chromatograms display the total ion count (TIC) for scans containing an ion with an $m / z$ of 518 corresponding to the $\mathbf{C D P}-1$ product. The mass spectrum is the scan at the center of the peak in the 30 min sample.




Figure S13. LC-MS analysis of 2 turnover. Chromatograms display the total ion count (TIC) for scans containing an ion with an $m / z$ of 516 corresponding to the CDP- 2 product. The mass spectrum is the scan at the center of the peak in the 30 min sample.




Figure S14. LC-MS analysis of $\mathbf{1}$ turnover. Chromatograms display the total ion count (TIC) for scans
containing an ion with an $m / z$ of 534 corresponding to the CDP-3 product. The mass spectrum is the scan at the center of the peak in the 30 min sample.




Figure S15. LC-MS analysis of 4 turnover. Chromatograms display the total ion count (TIC) for scans containing an ion with an $m / z$ of 554 corresponding to the CDP-4 product. The mass spectrum is the scan at 1.21 min (indicated by the red line) in the 30 min sample.




Figure S16. LC-MS analysis of 5a turnover. Chromatograms display the total ion count (TIC) for scans containing an ion with an $\mathrm{m} / \mathrm{z}$ of 536 corresponding to the CDP-5a product. The mass spectrum is the scan at the center of the peak in the 30 min sample.



Figure S17. LC-MS analysis of 5a turnover. Chromatograms display the total ion count (TIC) for scans containing an ion with an $m / z$ of 536 corresponding to the CDP-5b product. The mass spectrum is the scan at the center of the peak in the 30 min sample.


Figure S18. Michaelis-Menten plots for MEP and analogs 1 - 5a,b with E. coli IspD.


Figure S19. Evaluation of inhibitory activity of analogs 1 - 5a,b against a) E. coli, b) P. falciparum, and c) M. tuberculosis IspD. The white (-) MEP bars indicate control experiments were MEP was excluded from the reaction mixture in order to determine if the observed rates were being significantly influenced by analog turnover. In the $P$. falciparum experiments testing analogs $\mathbf{1 , 5 a}$ and $\mathbf{5 b}$, analog turnover appeared to be nonnegligible prompting the evaluation of these analogs as substrates (see below, figure S27).


Figure S20. IC50 plots for analog 5b against a) E. coli IspD and b) P. falciparum IspD.


1
Exact Mass: 213.0533


Figure S21. LC-HRMS analysis of analog 1 demonstrating $\mathbf{> 9 5 \%}$ purity.


2
Exact Mass: 211.0377



Figure S22. LC-HRMS analysis of analog 2 demonstrating $>\mathbf{9 5 \%}$ purity.


Exact Mass: 229.0283


Figure S23. LC-HRMS analysis of analog $\mathbf{3}$ demonstrating $>\mathbf{9 5 \%}$ purity.


Exact Mass: 249.0345



Figure S24. LC-HRMS analysis of analog 4 demonstrating $>95 \%$ purity.


5a
Exact Mass: 231.0439



Figure S25. LC-HRMS analysis of analog 5a demonstrating $\mathbf{> 9 5 \%}$ purity. The mass observed at 211.0379 corresponds to the fluoride elimination product, vinyl phosphonate $\mathbf{2}$. This product appears to be an artifact of the ionization process as there are no observable peaks corresponding to $\mathbf{2}$ in the ${ }^{1} \mathrm{H}$ NMR spectrum (below).


5b
Exact Mass: 231.0439


Figure S26. LC-HRMS analysis of analog 5b demonstrating $\mathbf{> 9 5 \%}$ purity. The mass observed at 211.0379 corresponds to the fluoride elimination product, vinyl phosphonate 2. This product appears to be an artifact of the ionization process as there are no observable peaks corresponding to $\mathbf{2}$ in the ${ }^{1} \mathrm{H}$ NMR spectrum (below).


Figure S27. Michaelis-Menten plots for MEP and analogs 1 and 5a,b with P. falciparum IspD. The relatively high $K_{\mathrm{m}}$ values determined by these experiments demonstrate that the non-negligible (-) MEP control rates observed in Figure S19 are very unlikely to be masking potent inhibition.


Figure S28. Overlay of ${ }^{1} \mathrm{H}$ spectra for diastereomers of phosphonolactone 20a. The top and bottom spectra are assigned to be the $R$ - and $S$-configurations at the phosphonate, respectively.




Figure S29. Overlay of ${ }^{1} \mathrm{H}$ spectra for diastereomers of phosphonolactone 20b. The top and bottom spectra are assigned to be the $R$ - and $S$-configurations at the phosphonate, respectively.






10a



10a



9a



9a







2



2




8b



10b



10b



3



3







14



14



Mosher's Acid Derivative of 14



15






4



16a



16a











5b



5b





19b



19b



20a- $R_{P}$


$20 \mathrm{a}-\boldsymbol{R}_{\mathrm{P}}$


$20 a-S_{P}$


$20 \mathrm{a}-S_{P}$



20b- $R_{P}$


$20 \mathrm{~b}-R_{\mathrm{P}}$


$20 b-S_{P}$



21a



21a



21b



21b


