

## **Supporting Information**

### **Identification of immune-responsive gene 1 (IRG1) as a novel target of A20**

Emmy Van Quickelberghe<sup>1,2</sup>, Arne Martens<sup>3,4</sup>, Ludger J.E. Goeminne<sup>1,2,5,6</sup>, Lieven Clement<sup>5,6</sup>, Geert van Loo<sup>3,4</sup>, Kris Gevaert<sup>1,2,\$</sup>

<sup>1</sup>VIB-UGent Center for Medical Biotechnology, B-9000 Ghent, Belgium

<sup>2</sup>Department of Biochemistry, Ghent University, B-9000 Ghent, Belgium

<sup>3</sup>VIB-UGent Center for Inflammation Research, B-9052 Ghent, Belgium

<sup>4</sup>Department of Biomedical Molecular Biology, Ghent University, B-9052 Ghent, Belgium

<sup>5</sup>Department of Applied Mathematics, Computer Science and Statistics, Ghent University, B-9000 Ghent, Belgium

<sup>6</sup>Bioinformatics Institute Ghent, Ghent University, B-9000 Ghent, Belgium

<sup>\$</sup> Correspondence to Kris Gevaert, VIB-UGent Center for Medical Biotechnology, Albert Baertsoenkaai 3, B-9000 Ghent, Belgium, Tel: +32-9-264.92.74, Fax: +32-9-264.94.90, Email: [kris.gevaert@vib-ugent.be](mailto:kris.gevaert@vib-ugent.be).

## **Table of Content**

**Figure S-1.** Completely annotated volcano plot visualizing protein regulation comparing the proteomes of wild-type (WT) and A20 deficient (KO) BMDMs following 6 h of TNF treatment.

**Figure S-2.** Uncropped Western blots showing expression of A20, IRG1 and Tubulin in WT and A20 KO BMDMs treated or not with 10 µg/ml LPS.

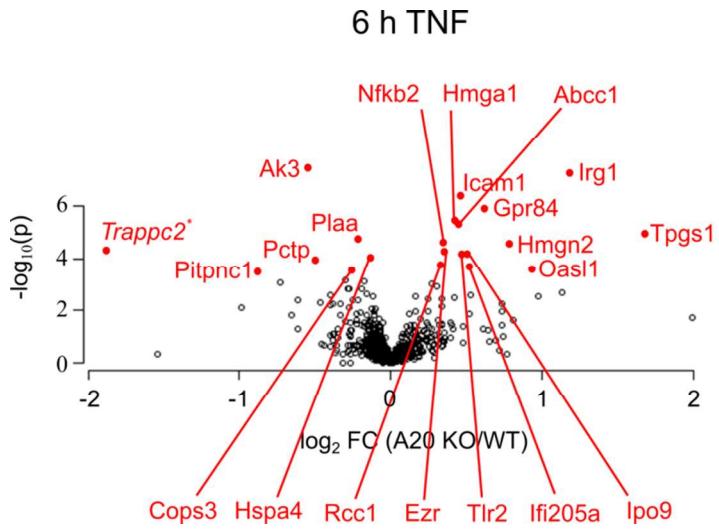
**Figure S-3.** Boxplots showing the  $\log_2$  peptide intensities of the significantly regulated proteins.

**Figure S-4.** Volcano plots visualizing protein regulation comparing the proteomes upon LPS or TNF treatment.

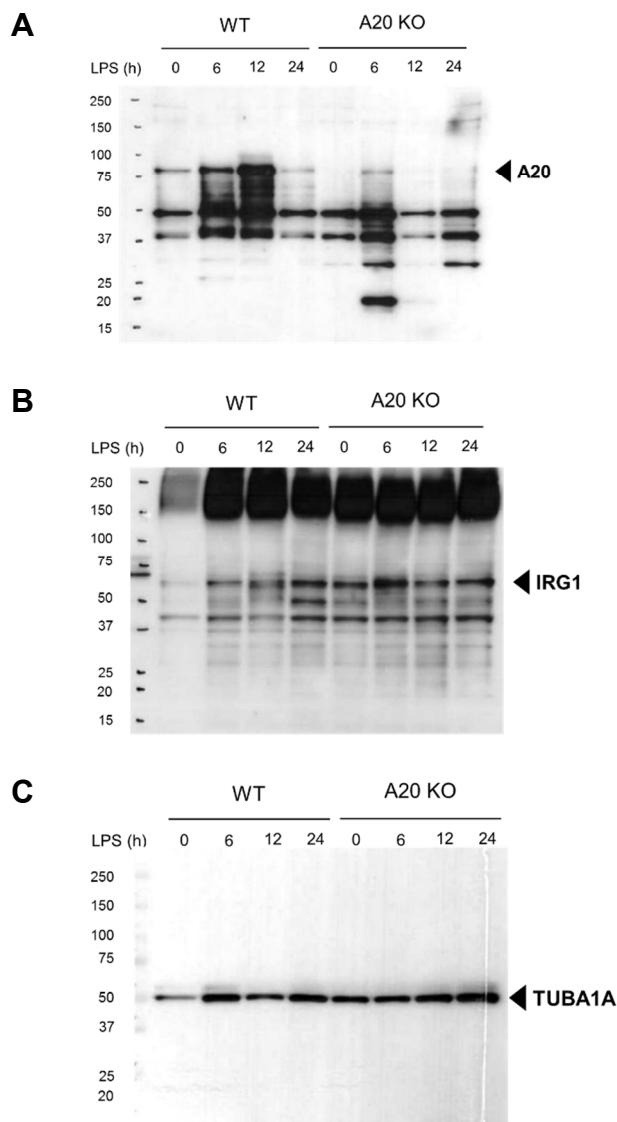
**Table S-1.** MSqRob Annotation file used for the MSqRob analysis.

**Table S-2.** MSqRob Results table.

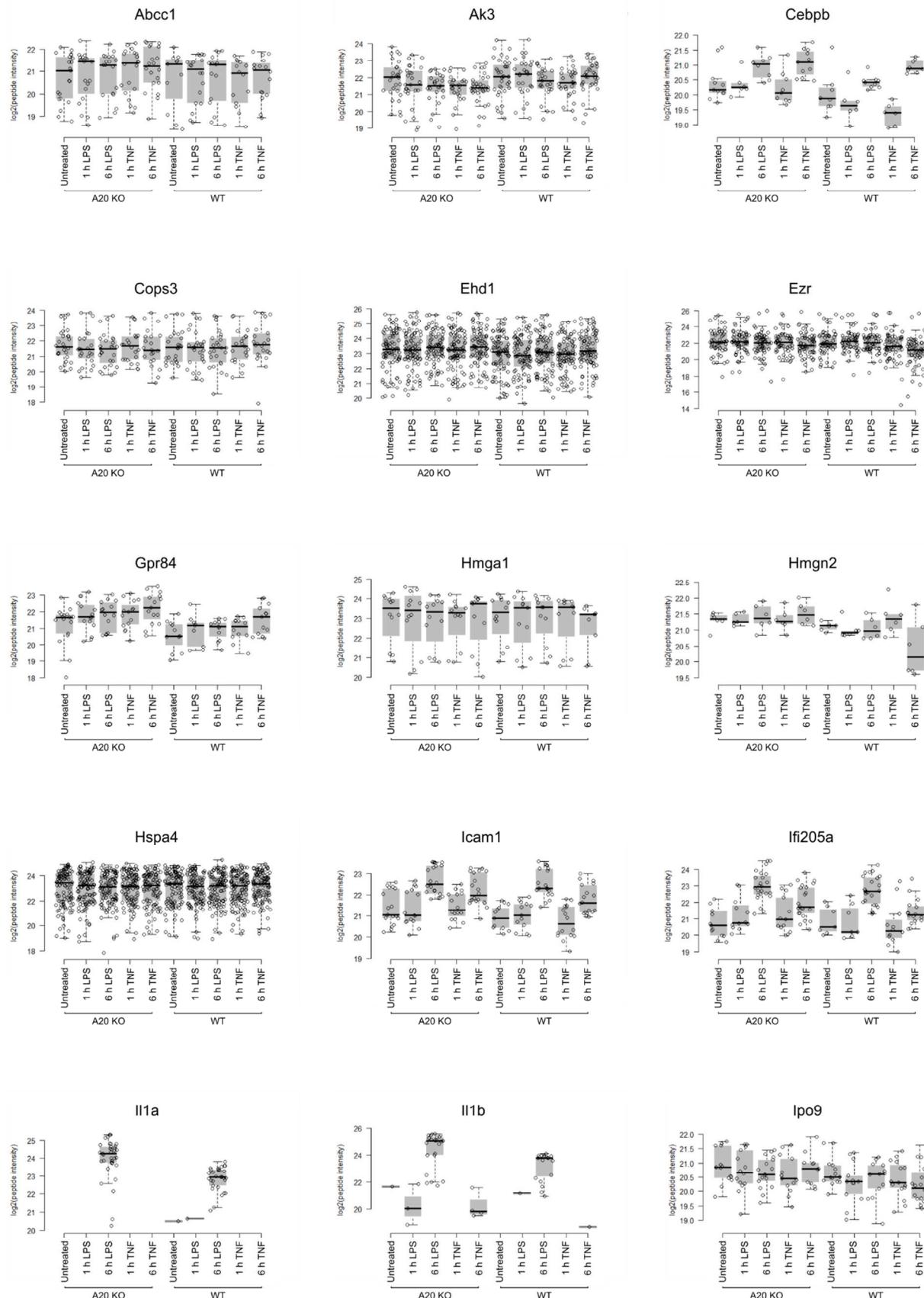
**Table S-3.** List of known substrates and associated proteins of A20.

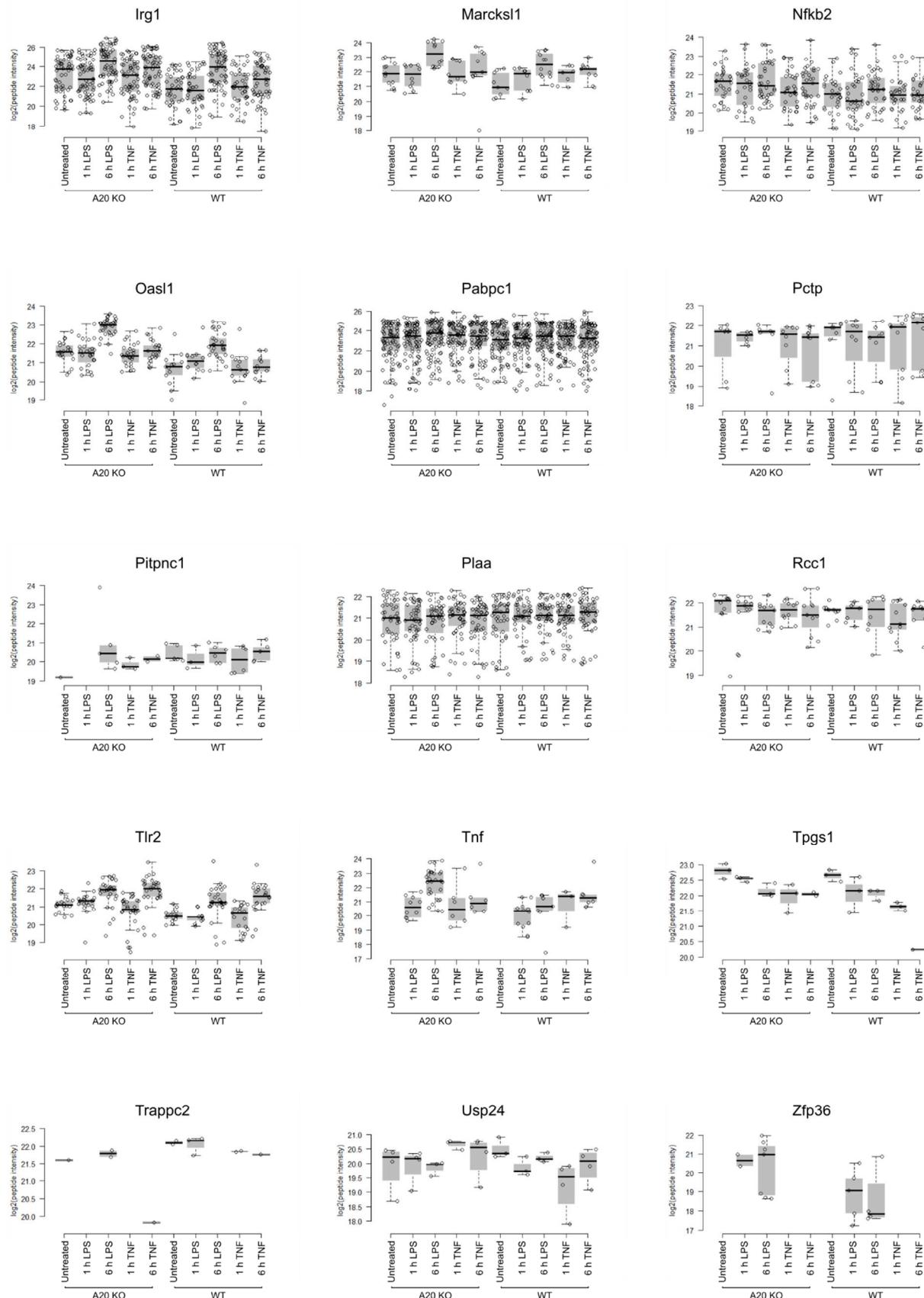


**Figure S-1.** Volcano plot visualizing protein regulation comparing the proteomes of wild-type (WT) and A20 deficient (KO) BMDMs following 6 h of TNF treatment. The x-axis shows the  $\log_2$  fold change of the protein abundances between A20 KO and WT BMDMs and the y-axis shows the  $-\log_{10}$  of the p-value. Significantly regulated proteins (FDR  $< 0.05$ ) between the A20 KO and WT BMDMs are represented with red dots and indicated with their gene name. Gene names in italics and indicated with an asterisk point to unreliable estimates as the quantification was based on only one peptide in one or more conditions.

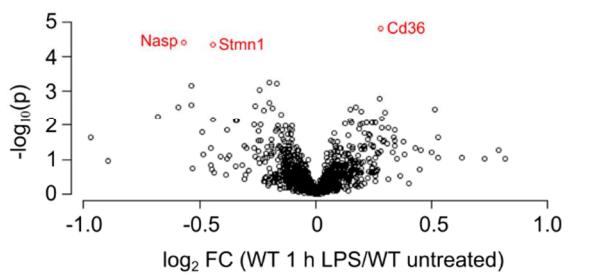
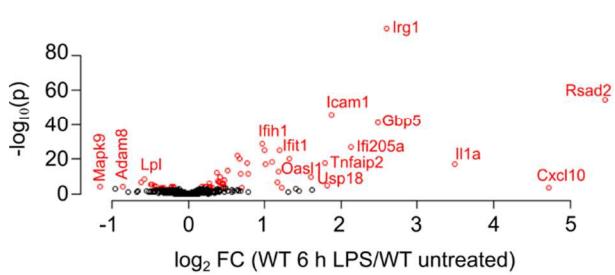
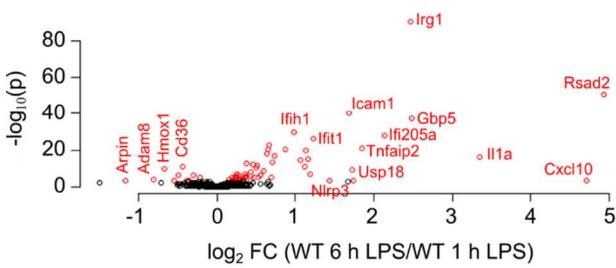
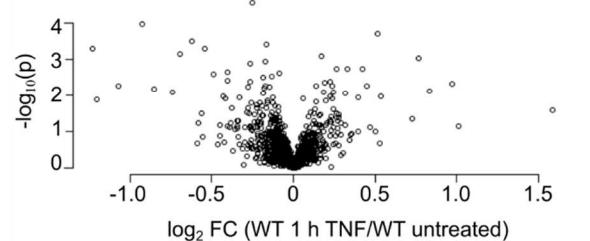
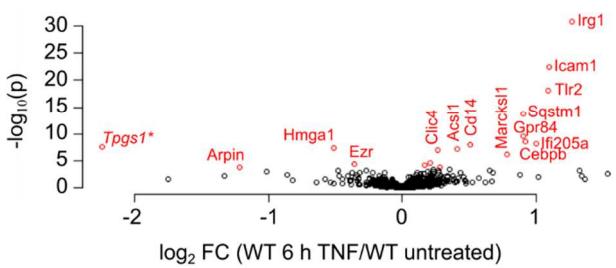
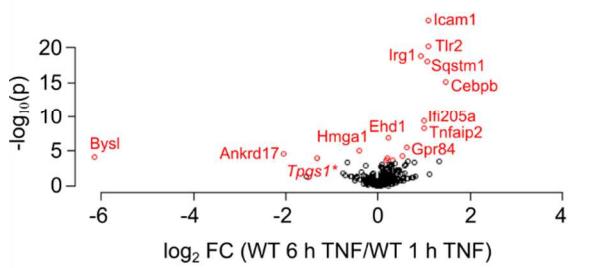
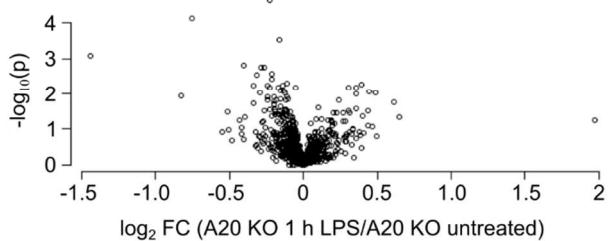
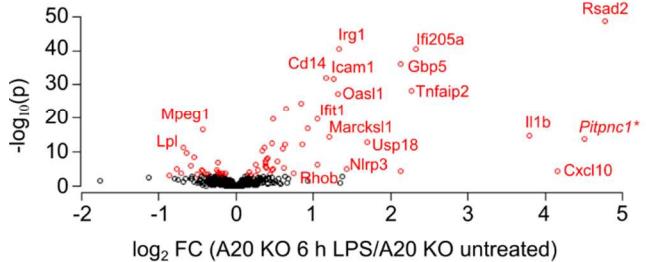


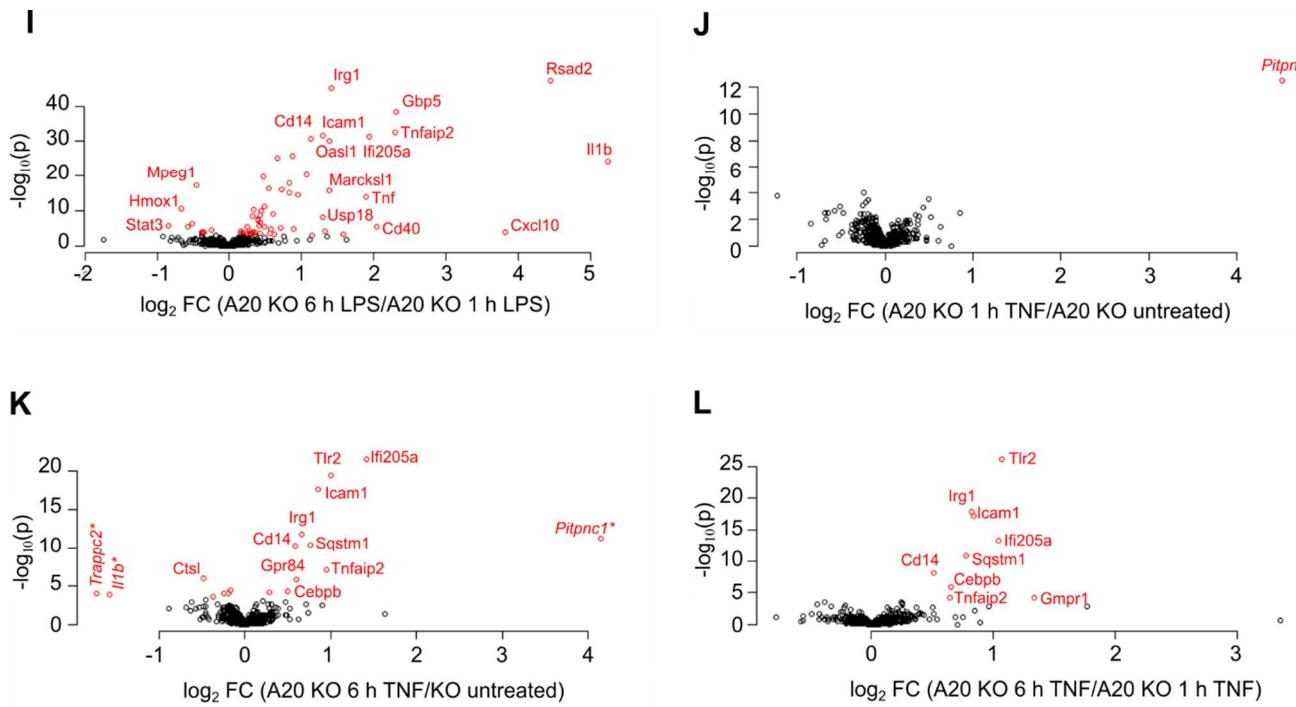
**Figure S-2.** Uncropped Western blots of Figure 3. Western blot analysis showing expression of A20 (A), IRG1 (B) and Tubulin (C) in WT and A20 KO BMDMs treated or not with 10 µg/ml LPS for the indicated time points.





**Figure S-3.** Boxplots showing the  $\log_2$  peptide intensities of the significantly regulated proteins. The y-axis shows the  $\log_2$  intensity of the peptide and the x-axis shows the different conditions studied. Each data point represents a single peptide, the median of the peptide intensity is represented by a thick black line, the box itself comprises the interquartile range and the whiskers extend to the most extreme data points that lies within 1.5 times the interquartile range on each side<sup>1</sup>. The gene name is given for every protein.

**A****B****C****D****E****F****G****H**



**Figure S-4.** Volcano plots visualizing protein regulation by comparing the proteomes upon LPS or TNF treatment. The x-axis shows the  $\log_2$  fold change of the protein abundances between two treatment conditions and the y-axis shows the  $-\log_{10}$  of the p-value. Significantly regulated proteins ( $\text{FDR} < 0.05$ ) are represented with red dots and indicated with their gene name. Gene names in italics and indicated with an asterisk point to unreliable estimates as the quantification was based on only one peptide in one or more conditions. For reasons of readability, not all significantly regulated proteins are indicated. However,  $\log_2$  fold changes and p-values for all proteins can be found in Table S-2.

**Table S-1.** MSqRob Annotation file. (Separate Excel file)

This is the annotation file used for the MSqRob analysis. The first column (“sample”) contains the unique names of the samples, the second (“treat”) and third (“mouse”) column contains respectively the variables treatment and mouse.<sup>1</sup>

**Table S-2.** MSqRob Results table. (Separate Excel file)

The results table shows all proteins from the MsqRob analysis. Every sheet contains the results for the differential protein abundance between genotypes (A20 KO vs WT BMDMs) for a treatment condition (untreated, after 1 h of LPS treatment, after 6 h of LPS treatment, after 1 h of TNF treatment and after 6 h of TNF treatment) as well as the results for the differential protein abundances within treatments for each genotype. The first, second and third column respectively show the protein accession, the gene name and the protein name for each protein. The column “Log2 fold change” indicates the estimated log2 fold change between the proteomes of WT and KO A20 BMDMs. “Standard error” denotes the standard error on this log2 fold change. Degrees of freedom, T- and p-values are given for the corresponding moderated t-tests in their respective columns. “q value” is the smallest Benjamini-Hochberg false discovery rate (FDR) threshold at which the protein would be significant. The column “significant” indicates whether the protein is significant at the default 5% FDR threshold.<sup>1</sup>

**Table S-3.** List of known substrates and associated proteins of A20. Presented values are the A20/WT ratio values of protein abundances in this differential proteomics study. For every protein, the cell type and treatment used to validate the substrate or interaction are presented, as well as if it leads to degradation of the protein.

		A20 KO/WT ratio					Literature				
Protein	Gene Name	-	1 h LPS	6 h LPS	1 h TNF	6 h TNF	Cell type *	Treatment *	Degradation	Ref.	
<b>E3 ligase activity</b>											
RIP1	RIPK1	1.01	1.00	0.98	0.99	0.95	MEF	TNF	yes	2	
ASK1	ASK1	N/A	N/A	N/A	N/A	N/A	HEK293T	TNF	yes	3	
<b>DUB activity</b>											
TRAF6	TRAF6	N/A	N/A	N/A	N/A	N/A	MEF	LPS, IL-17	no	4	
TBK1	TBK1	0.98	0.98	1.09	0.96	0.95	MEF	poly(I:C)	no	5	
IKKE	IKBKE	1.00	1.00	1.00	1.01	1.01	MEF	poly(I:C)	no	5	
Caspase 8	CASP8	1.00	1.00	1.00	1.00	1.00	HEK293T	-	no	6	
cIAP1	BIRC2	N/A	N/A	N/A	N/A	N/A	MEF	TNF, IL-1	no	7	
cIAP2	BIRC3	N/A	N/A	N/A	N/A	N/A	MEF	TNF, IL-1	no	7	
RIP3	RIPK3	0.98	0.99	1.02	1.02	1.00	MEF	TNF (and CHX and/or Z-VAD)	no	8	
RIP2	RIPK2	N/A	N/A	N/A	N/A	N/A	BMDM	MDP	no	9	
<b>Other</b>											
TRAF2	TRAF2	N/A	N/A	N/A	N/A	N/A	MEF	TNF, actinomycin D, TWEAK	yes	10	
UBC13	UBE2N	1.00	1.00	1.00	1.00	1.00	MEF	IL-1	no	11	
UBCH5C	UBE2D3	1.00	1.00	1.00	1.00	1.00	MEF	IL-1	no	11	
NEMO	IKBKG	1.00	1.00	1.00	1.00	1.00	MEF	TNF	no	12-14	
HOIP	RNF31	1.00	1.00	1.00	1.00	1.00	MEF	TNF	no	12-14	
HOIL-1	RBCK1	N/A	N/A	N/A	N/A	N/A	MEF	TNF	no	12-14	
SHARPIN	SHARPIN	N/A	N/A	N/A	N/A	N/A	MEF	TNF	no	12-14	
NLRP3	NLRP3	1.28	1.59	1.47	1.85	1.67	BMDM	LPS and ATP and nigericin	no	15	
STAT1	STAT1	1.04	0.99	1.01	0.98	1.03	BMDM	IFNY	no	16	

A20 ubiquitin-editing complex												
ABIN-1	TNIP1	N/A	N/A	N/A	N/A	N/A	HEK293T	-		no		17
ABIN-2	TNIP2	N/A	N/A	N/A	N/A	N/A	HEK293T	TNF, IL-1		no		18
ABIN-3	TNIP3	N/A	N/A	N/A	N/A	N/A	THP-1	TNF, LPS		no		19
TAX1BP1	TAX1BP1	N/A	N/A	N/A	N/A	N/A	MEF	TNF, IL-1		no		20
ITCH	ITCH	1.00	1.00	1.00	1.00	1.00	MEF	TNF, LPS		no		21
RNF11	RNF11	N/A	N/A	N/A	N/A	N/A	MEF	TNF, LPS		no		22

CHX: Cycloheximide; Z-VAD: pan-caspase inhibitor (carbobenzoxy-valyl-alanyl-aspartyl-[O-methyl]-fluoromethylketone); TNF–CHX–Z-VAD: 'cocktail' for caspase-independent necroptosis induction; MDP: Muramyl dipeptide; TWEAK: TNF-related weak inducer of apoptosis; ATP: Adenosine triphosphate; ATP and nigericin: NLRP3 inflammasome stimuli; (\*) If multiple cell lines and/or stimulation conditions were used in the referred study, the most relevant experiments for comparison with our study are represented in this table .

1. Goeminne, L. J. E.; Gevaert, K.; Clement, L., Experimental design and data-analysis in label-free quantitative LC/MS proteomics: A tutorial with MSqRob. *J Proteomics* **2017**.
2. Wertz, I. E.; O'Rourke, K. M.; Zhou, H.; Eby, M.; Aravind, L.; Seshagiri, S.; Wu, P.; Wiesmann, C.; Baker, R.; Boone, D. L.; Ma, A.; Koonin, E. V.; Dixit, V. M., De-ubiquitination and ubiquitin ligase domains of A20 downregulate NF-kappaB signalling. *Nature* **2004**, *430* (7000), 694-9.
3. Won, M.; Park, K. A.; Byun, H. S.; Sohn, K. C.; Kim, Y. R.; Jeon, J.; Hong, J. H.; Park, J.; Seok, J. H.; Kim, J. M.; Yoon, W. H.; Jang, I. S.; Shen, H. M.; Liu, Z. G.; Hur, G. M., Novel anti-apoptotic mechanism of A20 through targeting ASK1 to suppress TNF-induced JNK activation. *Cell Death Differ* **2010**, *17* (12), 1830-41.
4. Boone, D. L.; Turer, E. E.; Lee, E. G.; Ahmad, R. C.; Wheeler, M. T.; Tsui, C.; Hurley, P.; Chien, M.; Chai, S.; Hitotsumatsu, O.; McNally, E.; Pickart, C.; Ma, A., The ubiquitin-modifying enzyme A20 is required for termination of Toll-like receptor responses. *Nat Immunol* **2004**, *5* (10), 1052-60.
5. Parvatiyar, K.; Barber, G. N.; Harhaj, E. W., TAX1BP1 and A20 inhibit antiviral signaling by targeting TBK1-IKKi kinases. *J Biol Chem* **2010**, *285* (20), 14999-5009.
6. Jin, Z.; Li, Y.; Pitti, R.; Lawrence, D.; Pham, V. C.; Lill, J. R.; Ashkenazi, A., Cullin3-based polyubiquitination and p62-dependent aggregation of caspase-8 mediate extrinsic apoptosis signaling. *Cell* **2009**, *137* (4), 721-35.
7. Yamaguchi, N., The seventh zinc finger motif of A20 is required for the suppression of TNF-alpha-induced apoptosis. *FEBS Lett* **2015**, *589* (12), 1369-75.
8. Onizawa, M.; Oshima, S.; Schulze-Topphoff, U.; Osés-Prieto, J. A.; Lu, T.; Tavares, R.; Prodhomme, T.; Duong, B.; Whang, M. I.; Advincula, R.; Agelidis, A.; Barrera, J.; Wu, H.; Burlingame, A.; Malynn, B. A.; Zamvil, S. S.; Ma, A., The ubiquitin-modifying enzyme A20 restricts ubiquitination of the kinase RIPK3 and protects cells from necroptosis. *Nat Immunol* **2015**, *16* (6), 618-27.
9. Hitotsumatsu, O.; Ahmad, R. C.; Tavares, R.; Wang, M.; Philpott, D.; Turer, E. E.; Lee, B. L.; Shiffin, N.; Advincula, R.; Malynn, B. A.; Werts, C.; Ma, A., The ubiquitin-editing enzyme A20 restricts nucleotide-binding oligomerization domain containing 2-triggered signals. *Immunity* **2008**, *28* (3), 381-90.
10. Li, L.; Soetandyo, N.; Wang, Q.; Ye, Y., The zinc finger protein A20 targets TRAF2 to the lysosomes for degradation. *Biochim Biophys Acta* **2009**, *1793* (2), 346-53.
11. Shembade, N.; Harhaj, E., A20 inhibition of NFkappaB and inflammation: targeting E2:E3 ubiquitin enzyme complexes. *Cell Cycle* **2010**, *9* (13), 2481-2.
12. Skaug, B.; Chen, J.; Du, F.; He, J.; Ma, A.; Chen, Z. J., Direct, noncatalytic mechanism of IKK inhibition by A20. *Mol Cell* **2011**, *44* (4), 559-71.
13. Tokunaga, F.; Nishimasu, H.; Ishitani, R.; Goto, E.; Noguchi, T.; Mio, K.; Kamei, K.; Ma, A.; Iwai, K.; Nureki, O., Specific recognition of linear polyubiquitin by A20 zinc finger 7 is involved in NF-kappaB regulation. *EMBO J* **2012**, *31* (19), 3856-70.
14. Verhelst, K.; Carpentier, I.; Kreike, M.; Meloni, L.; Verstrepen, L.; Kensche, T.; Dikic, I.; Beyaert, R., A20 inhibits LUBAC-mediated NF-kappaB activation by binding linear polyubiquitin chains via its zinc finger 7. *EMBO J* **2012**, *31* (19), 3845-55.
15. Vande Walle, L.; Van Opdenbosch, N.; Jacques, P.; Fossoul, A.; Verheugen, E.; Vogel, P.; Beyaert, R.; Elewaut, D.; Kanneganti, T. D.; van Loo, G.; Lamkanfi, M., Negative regulation of the NLRP3 inflammasome by A20 protects against arthritis. *Nature* **2014**, *512* (7512), 69-73.

16. De Wilde, K.; Martens, A.; Lambrecht, S.; Jacques, P.; Drennan, M. B.; Debusschere, K.; Govindarajan, S.; Coudenys, J.; Verheugen, E.; Windels, F.; Catrysse, L.; Lories, R.; McGonagle, D.; Beyaert, R.; van Loo, G.; Elewaut, D., A20 inhibition of STAT1 expression in myeloid cells: a novel endogenous regulatory mechanism preventing development of enthesitis. *Ann Rheum Dis* **2017**, *76* (3), 585-592.
17. Mauro, C.; Pacifico, F.; Lavorgna, A.; Mellone, S.; Iannetti, A.; Acquaviva, R.; Formisano, S.; Vito, P.; Leonardi, A., ABIN-1 binds to NEMO/IKKgamma and co-operates with A20 in inhibiting NF-kappaB. *J Biol Chem* **2006**, *281* (27), 18482-8.
18. Van Huffel, S.; Delaei, F.; Heyninck, K.; De Valck, D.; Beyaert, R., Identification of a novel A20-binding inhibitor of nuclear factor-kappa B activation termed ABIN-2. *J Biol Chem* **2001**, *276* (32), 30216-23.
19. Wullaert, A.; Verstrepen, L.; Van Huffel, S.; Adib-Conquy, M.; Cornelis, S.; Kreike, M.; Haegman, M.; El Bakkouri, K.; Sanders, M.; Verhelst, K.; Carpentier, I.; Cavaillon, J. M.; Heyninck, K.; Beyaert, R., LIND/ABIN-3 is a novel lipopolysaccharide-inducible inhibitor of NF-kappaB activation. *J Biol Chem* **2007**, *282* (1), 81-90.
20. Shembade, N.; Harhaj, N. S.; Liebl, D. J.; Harhaj, E. W., Essential role for TAX1BP1 in the termination of TNF-alpha-, IL-1- and LPS-mediated NF-kappaB and JNK signaling. *EMBO J* **2007**, *26* (17), 3910-22.
21. Shembade, N.; Harhaj, N. S.; Parvatiyar, K.; Copeland, N. G.; Jenkins, N. A.; Matesic, L. E.; Harhaj, E. W., The E3 ligase Itch negatively regulates inflammatory signaling pathways by controlling the function of the ubiquitin-editing enzyme A20. *Nat Immunol* **2008**, *9* (3), 254-62.
22. Shembade, N.; Parvatiyar, K.; Harhaj, N. S.; Harhaj, E. W., The ubiquitin-editing enzyme A20 requires RNF11 to downregulate NF-kappaB signalling. *EMBO J* **2009**, *28* (5), 513-22.