

Supporting Information

Development of a selective labeling probe for Bruton's tyrosine kinase quantification in live cells

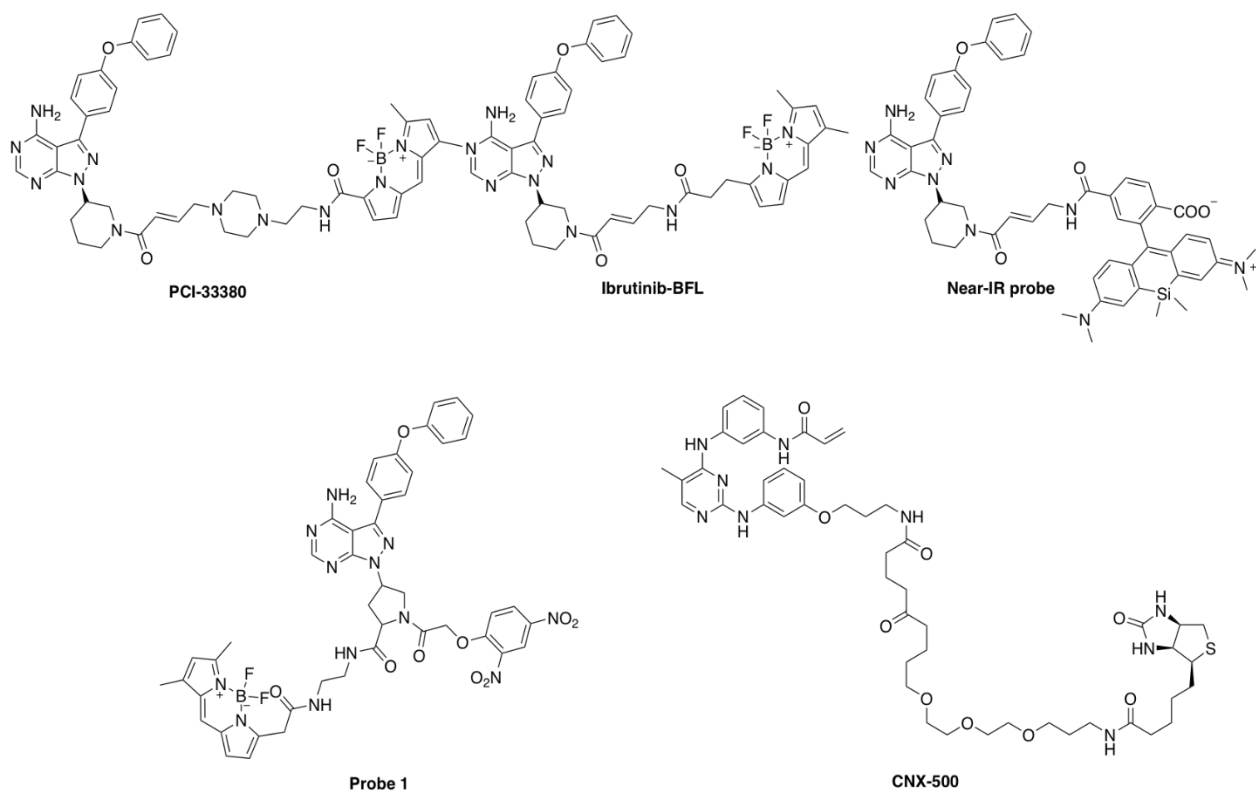
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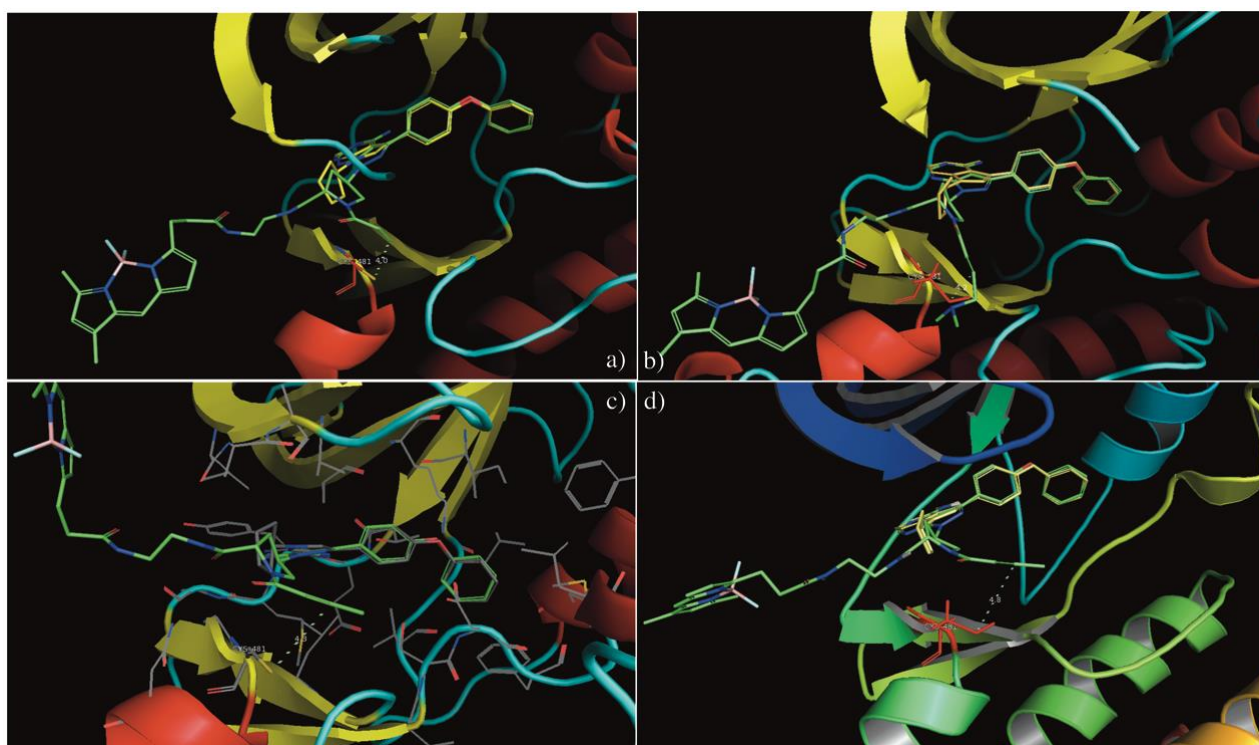
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I. Supporting Figures

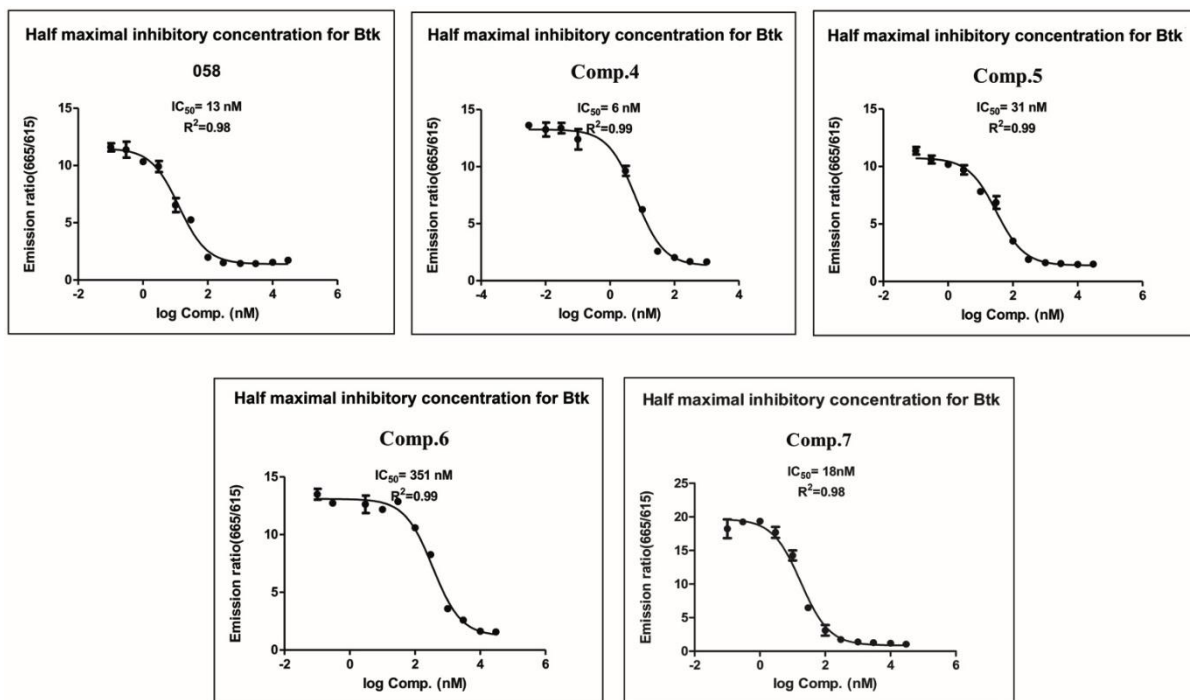


Supporting Figure 1. Structures of representative Btk probes.

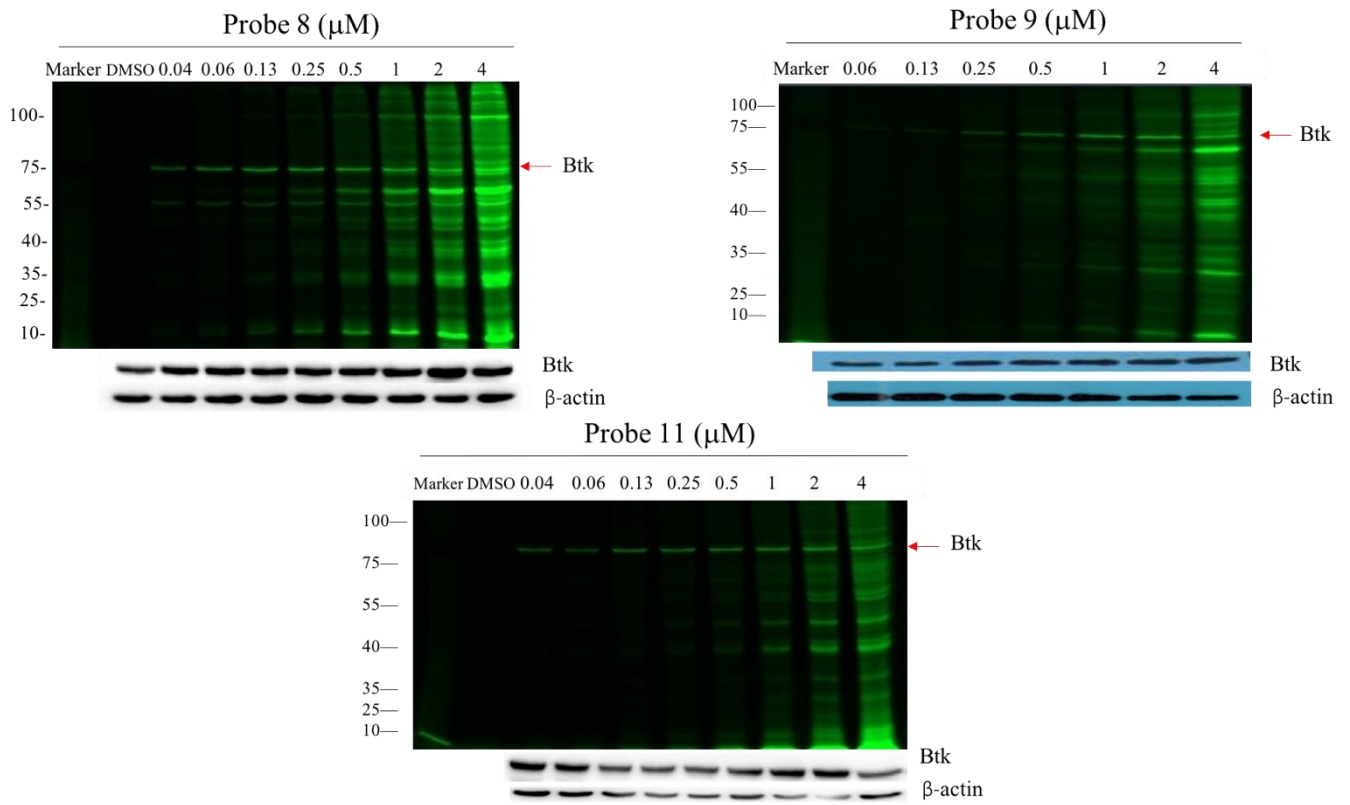


Supporting Figure 2. Docking models of probes in Btk's active site (carbon atoms in green, oxygen atom in red, nitrogen in blue). (Generated by MOE 2015)

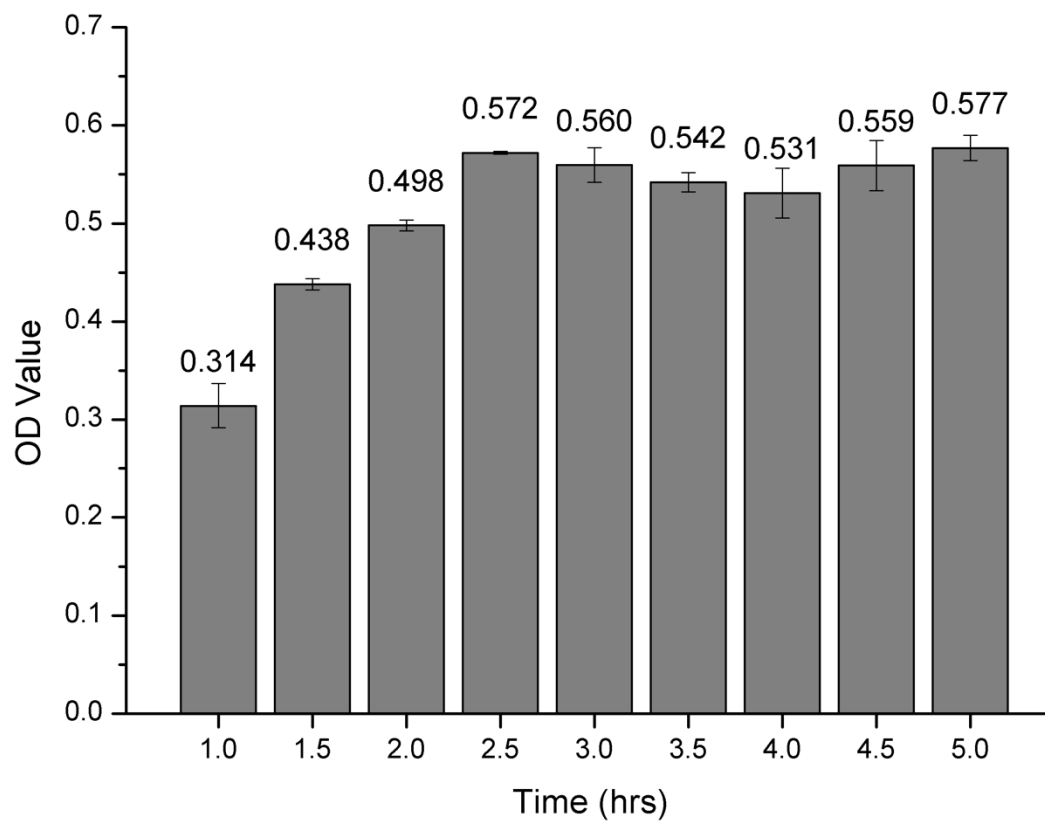
a) probe **8** b) probe **9** c) probe **11** d) probe **10** Cys481 is at the C-terminus of the hinge region and the distances between the warhead and the residue were show in red. All probes were predicted to overlap with the original ligand and the reactive groups are about 4 Å away from the cysteine residue.



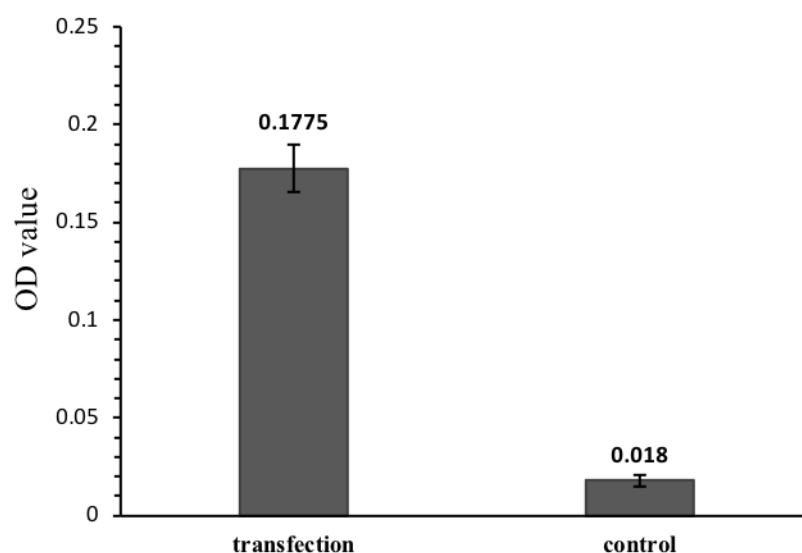
Supporting Figure 3. The IC₅₀ values of the four probe precursors. Compound **058** is a known Btk inhibitor with a reported IC₅₀ of 5 nM [compound **31** in *J Med Chem* **2014**, 57, 5112].



Supporting Figure 4. Btk in OCI-Ly7 cells were labelled by probes **8**, **9** and **11**.

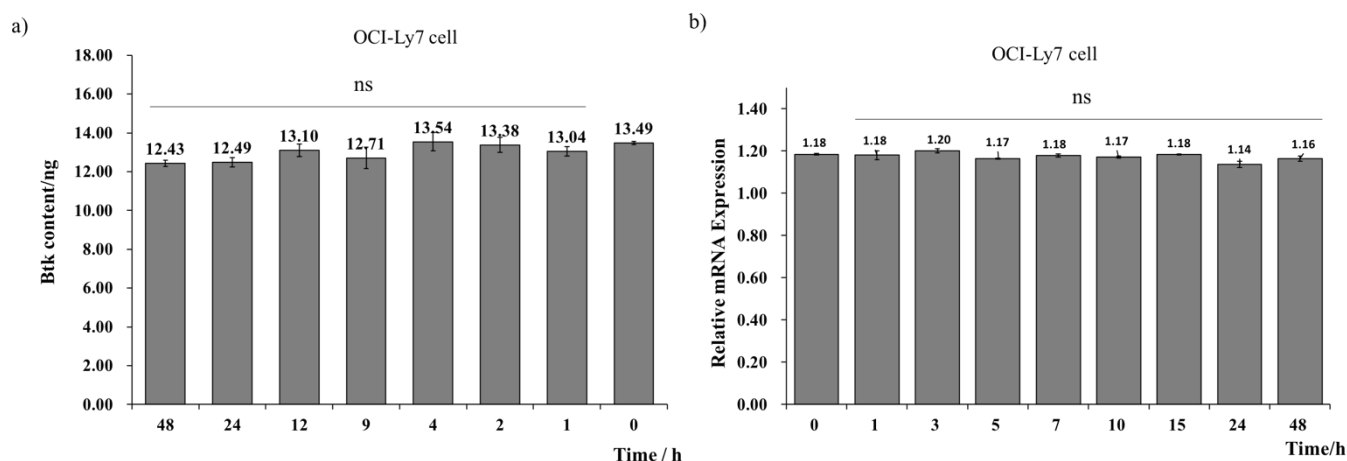


Supporting Figure 5. The relationship between sample incubation time and fluorescence readout. The cell lysates in each well were incubated for different time from 1 to 5 hours. It requires 2.5 hours for the antibody to fully capture Btk protein under our experiment conditions.



Supporting Figure 6. The detection result of Btk transfection in Jurkat cells by ELISA.

The human Btk plasmid were transferred into 100 thousand Jurkat cells by Electroporation technology. After 48 hours in 37°C, 5%CO₂, the cells were incubated with probe **10** (2 μM). After 1 hour, the cells were harvested and detected by ELISA method. The result showed significant difference between the two samples.



Supporting Figure 7. a) The Btk content in OCI-Ly7 cell after BCR activation for 0 to 48 hours. b) The relative mRNA expression levels in OCI-Ly7 BCR activation. Differences were evaluated using Student's *t* test comparing the value of *T*= 0 hour with the later time points; ns, non-significant. All the data are mean ± s.e.m. of two independent experiments measured in duplicate (n=2).

Btk/ng	OD value	Std. Error Mean
300.00	2.7434	0.02
150.00	1.5548	0.03
75.00	0.7894	0.09
37.50	0.3658	0.08
18.75	0.2319	0.02
9.38	0.1775	0.01
4.69	0.1128	0.00
2.34	0.0985	0.02
1.17	0.0935	0.01
0.59	0.0898	0.01

Supporting Figure 8. The raw data of standard curve.

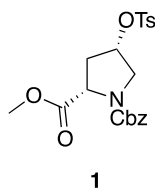
II. Chemistry

General Information

^1H NMR and ^{13}C NMR spectra were recorded on Bruker Advance 400 and 500 spectrometers at ambient temperature. Chemical shifts are reported in ppm relative to residual chloroform (δ_{H} 7.26 and δ_{C} 77.16) and dimethylsulfoxide (δ_{H} 2.50 and δ_{C} 39.52) as standards. Data are reported as follows: chemical shift, multiplicity (s = singlet, d = doublet, t = triplet, q = quartet, br = broad, m = multiplet), coupling constants and number of protons. Purity data were estimated by HPLC with UV detection at 254 nm. Mass spectrometric data were obtained using an AB Q-Star mass spectrometer.

Synthesis of Compounds

Compound 1

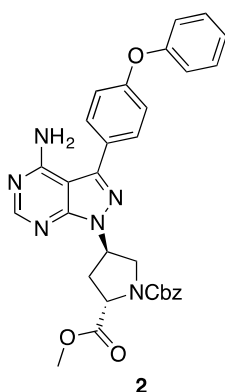


Thionyl chloride (9.5 ml, 133.5 mmol) was added to (4*S*)-4-Hydroxy-L-proline (5 g, 38.15 mmol) dissolved in 150 ml methanol for 1h. The reaction was stirred for 4h at room temperature. The mixture was concentrated and dissolved in 150 ml DCM. Cbz-OSU (11 g, 46 mmol) and Et₃N (9 ml, 76 mmol) were added to the mixture. The reaction was stirred for 4 hours at room temperature. The mixture was concentrated and extracted with ethyl acetate and water three times. The ethyl acetate layers were collected and washed with saturated aq. NaCl solution, dried over anhydrous Na₂SO₄, concentrated in vacuo and purified by flash column chromatography (EtOAc : Hexanes = 1:3) to give an intermediate as white solids (yield 98%).

P-TsCl (0.4 g, 2.2 mmol), DMAP (6 mg, 0.05 mmol) and Et₃N (0.3 ml, 2 mmol) were added to the intermediate (0.3 g, 1 mmol). The reaction was stirred for 4 hours at room temperature. The mixture was concentrated and extracted with ethyl acetate and water three times. The ethyl acetate layers were collected and washed with brine, dried over anhydrous Na₂SO₄, concentrated in vacuo and purified by flash column chromatography (EtOAc : Hexanes = 1:1) to give compound **1** as white oil (yield 74%).

¹HNMR (400 MHz, CDCl₃) δ 7.73 (d, *J* = 8.3 Hz, 2H), 7.37 – 7.20 (m, 7H), 5.20 – 4.94 (m, 3H), 4.47 (ddd, *J* = 24.8, 7.8, 3.9 Hz, 1H), 3.75 – 3.52 (m, 5H), 2.40 (qd, *J* = 8.5, 4.6 Hz, 5H). ¹³CNMR (101 MHz, CDCl₃) δ 171.56, 171.30, 154.39, 154.03, 145.27, 136.27, 133.56, 130.04, 128.53, 128.09, 127.75, 78.72, 77.83, 77.48, 77.16, 76.84, 67.38, 57.58, 57.33, 52.36, 52.00 – 51.72, 37.21, 36.12, 21.71. HRMS (*m/z*): [M+H]⁺ calcd. for C₂₁H₂₄NO₇S⁺, 434.1273; found: 434.1278.

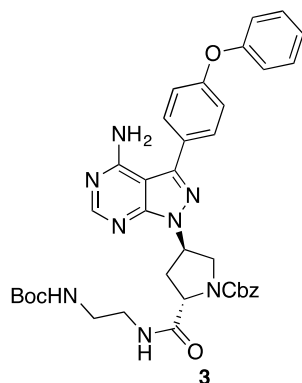
Compound 2



The 3-(4-Phenoxyphenyl)-1*H*-pyrazolo (3,4 d) pyridine-4-amine (1 g, 3.3 mmol) was dissolved in 25 ml DMF and stirred with potassium carbonate (0.9 g, 6.6 mmol) for 30 min. Compound **1** obtained in the previous step dissolved in 5 ml DMF was added to the reaction system. The reaction mixture was heated to 80°C and stirred overnight under argon atmosphere. The mixture was concentrated and extracted with ethyl acetate and water three times. The ethyl acetate layers were collected and washed with brine, dried over anhydrous Na₂SO₄, concentrated in vacuo and purified by flash column chromatography (EtOAc : Hexanes = 3:1) to give compound **2** as white solids (yield 90%).

¹H NMR (400 MHz, CDCl₃) δ 8.28 (s, 1H), 7.61 – 7.52 (m, 2H), 7.34 (dd, *J* = 11.5, 4.3 Hz, 2H), 7.30 – 7.18 (m, 5H), 7.16 – 6.98 (m, 5H), 6.04 (s, 2H), 5.68 – 5.51 (m, 1H), 5.15 (dd, *J* = 12.4, 7.0 Hz, 1H), 5.05 (dd, *J* = 12.4, 9.2 Hz, 1H), 4.81 – 4.70 (m, 1H), 4.13 – 3.93 (m, 2H), 3.75 (s, 1.5H), 3.60 (s, 1.5H), 3.05 – 2.90 (m, 1H), 2.52 – 2.40 (m, 1H). ¹³C NMR (101 MHz, CDCl₃) δ 172.75, 158.70, 158.15, 156.34, 155.90, 154.84, 154.61, 154.24, 144.46, 136.45, 130.01, 128.49, 128.18 – 127.71, 124.16, 119.67, 119.12, 98.75, 77.48, 77.16, 76.84, 67.31, 58.72, 54.45, 53.87, 53.52, 52.52, 51.66 (s), 50.86, 35.75, 34.81, 31.64, 22.71, 14.19. HRMS (*m/z*): [M+H]⁺ calcd. for C₃₁H₂₉N₆O₅⁺, 565.2194; found: 565.2194.

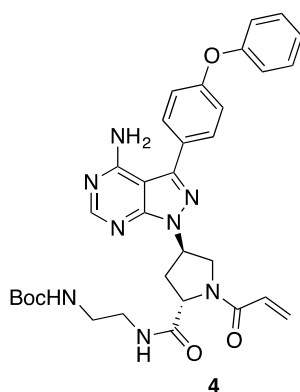
Compound 3.



Compound **2** (1 g, 1.8 mmol) was dissolved in 15 ml tetrahydrofuran : methanol : water = 9 : 3 : 3 at 0°C. 1N LiOH (0.1 g, 4.55 mmol) was added slowly to the reaction mixture. After 4 hours, the mixture was concentrated and extracted with ethyl acetate and water. The aqueous layers were combined and acidified to pH = 1 with 1N HCl at 0°C. The reaction mixture was extracted with ethyl acetate and water three times. The organic layers were collected and dried over anhydrous Na₂SO₄ to give the carboxylic acid. EDCI (1.7 g, 9 mmol), HOBt (0.5 g, 3.6 mmol), DIPEA (15 ml, 9 mmol) and N-(2-aminoethyl) carbamic acid *tert*-butyl ester (0.1 g, 4.55 mmol) were added to the solution of the acid under argon atmosphere. The mixture was stirred for 4 hours and then extracted with ethyl acetate and water. The organic layer was washed with saturated NaCl solution, dried over anhydrous Na₂SO₄, concentrated and purified by flash column chromatography (EtOAc : Hexanes = 5:1) to give compound **3** as white solids (yield 90 %).

¹H NMR (500 MHz, CDCl₃) δ 8.31 (s, 1H), 7.63 – 7.55 (m, 2H), 7.41 – 7.27 (m, 7H), 7.18 – 7.14 (m, 1H), 7.12 (d, *J* = 8.6 Hz, 2H), 7.07 (dd, *J* = 8.6, 1.0 Hz, 2H), 5.99 (s, 2H), 5.54 – 5.37 (m, 1H), 5.22 (d, *J* = 12.3 Hz, 1H), 5.12 (d, *J* = 12.3 Hz, 1H), 4.50 – 4.40 (m, 1H), 4.11 (s, 2H), 3.04 (dd, *J* = 197.6, 78.4 Hz, 6H), 1.36 (s, 9H). ¹³C NMR (126 MHz, CDCl₃) δ 158.95 – 158.66, 158.26 – 158.00, 156.40 – 156.38, 156.38 – 155.74, 154.85 – 154.41, 144.41 – 144.16, 136.22 – 135.93, 130.28 – 129.93, 128.80 – 128.51, 128.51 – 128.21, 128.21 – 128.02, 127.81 – 127.30, 124.43 – 124.15, 119.86 – 119.61, 119.17 – 118.97, 98.95 – 98.65, 77.41, 77.16, 76.91, 67.73, 60.40, 40.17, 28.46. HRMS (*m/z*): [M+H]⁺ calcd. for C₃₇H₄₁N₈O₆⁺, 693.3144; found: 693.3164.

Compound 4.

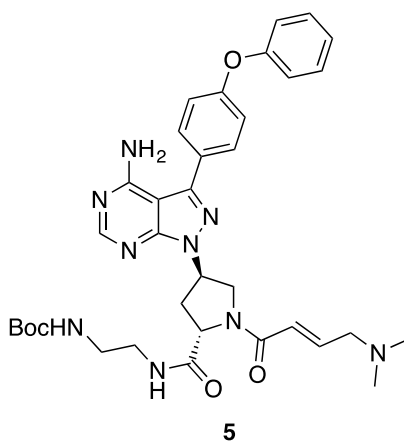


Compound **3** (220 mg, 0.32 mmol) was dissolved in 3 mL ethyl acetate and a few drops of methanol. A catalytic palladium hydroxide on carbon was added. The mixture was stirred under hydrogen at room temperature for 4 hours. The mixture was filtered with celatom and concentrated to give intermediate which was directly used in the next reaction without purification.

The reaction intermediate (60 mg, 0.1 mmol) was dissolved in a mixed solvent (1.5 ml tetrahydrofuran and two drops of water). Et₃N (28 mg, 40 μ l, 0.27 mmol), acryloyl chloride (9 mg, 0.1 mmol) were added at 0°C. The mixture was stirred for 2 hours in room temperature. The mixture was concentrated and extracted with ethyl acetate and saturated NaCl solution three times. The organic layers was dried over anhydrous Na₂SO₄, filtered and concentrated by rotary evaporator in vacuo. The concentrate was purified by flash column chromatography (EtOAc : Hexanes = 3:1) to give the compound **4** as white solid (yield 70%).

¹H NMR (400 MHz, CDCl₃) δ 8.36 (s, 1H), 7.62 (d, *J* = 8.6 Hz, 2H), 7.39 (t, *J* = 7.9 Hz, 2H), 7.21–7.11 (m, 3H), 7.08 (d, *J* = 7.7 Hz, 3H), 6.54–6.41 (m, 2H), 5.79 (dd, *J* = 8.2, 3.9 Hz, 1H), 5.66 (s, 2H), 5.49 (dt, *J* = 15.3, 7.5 Hz, 1H), 5.16 (s, 1H), 4.68 (t, *J* = 7.9 Hz, 1H), 4.32 (dd, *J* = 10.5, 7.5 Hz, 1H), 4.27–4.18 (m, 1H), 3.39–3.20 (m, 2H), 3.13–2.88 (m, 2H), 2.78–2.61 (m, 1H), 1.39 (s, 9H). ¹³C NMR (101 MHz, CDCl₃) δ 170.90 (s), 166.05 (s), 158.88 (s), 157.92 (s), 156.33 (s), 154.88 (s), 144.41 (s), 130.11 (d, *J* = 8.5 Hz), 127.95 (s), 127.48 (s), 119.77 (s), 119.14 (s), 99.00 (s), 79.30 (s), 77.48 (s), 77.16 (s), 76.84 (s), 59.78 (s), 54.68 (s), 51.05 (s), 40.40 (s), 40.11 (s), 33.25 (s), 28.53 (s). HRMS (*m/z*): [M+H]⁺ calcd. for C₃₂H₃₇N₈O₅⁺, 613.2881; found: 613.2897.

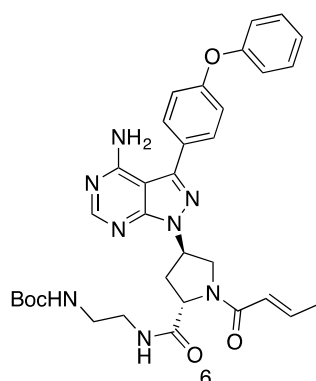
Compound 5.



Compound **5** was synthesized with a similar procedure as compound **4**. (2*E*)-4-(Dimethylamino)-2-butenoic acid (31 mg, 0.36 mmol), HOBt (25 mg, 0.18 mmol), EDCI (86 mg, 0.45 mmol) and Et₃N (45 mg, 0.45 mmol) were added to the intermediate in 3 ml DCM and stirred for 6 hours. The mixture was purified with a similar procedure to give compound **5** as white solids (yield 55%).

¹HNMR (400 MHz, CDCl₃) δ 8.29 (s, 4H), 7.58 (t, *J* = 9.7 Hz, 8H), 7.35 (t, *J* = 7.9 Hz, 10H), 7.17–7.00 (m, 20H), 6.93 (dt, *J* = 15.1, 5.8 Hz, 4H), 6.31 (d, *J* = 15.2 Hz, 4H), 5.49–5.39 (m, 4H), 4.64 (t, *J* = 8.1 Hz, 4H), 4.24 (dt, *J* = 17.6, 10.1 Hz, 7H), 3.26 (ddd, *J* = 19.5, 13.7, 7.2 Hz, 8H), 3.16 (d, *J* = 5.1 Hz, 7H), 2.99 (ddd, *J* = 21.9, 11.8, 5.6 Hz, 15H), 2.75–2.58 (m, 3H), 2.20 (s, 24H), 1.34 (s, 36H). ¹³CNMR (101 MHz, CDCl₃) δ 171.23, 165.81, 158.71, 158.16, 156.79–156.50, 156.10, 154.76, 144.44, 130.02, 127.47, 124.16, 122.48, 119.65, 119.06, 98.82, 79.15, 77.50, 77.18, 76.86, 60.61, 59.60, 54.49, 50.99, 45.59, 40.13, 31.62, 28.44, 22.69, 14.17. HRMS (*m/z*): [M+H]⁺ calcd. for C₃₅H₄₄N₉O₅⁺, 670.3460; found: 670.3460.

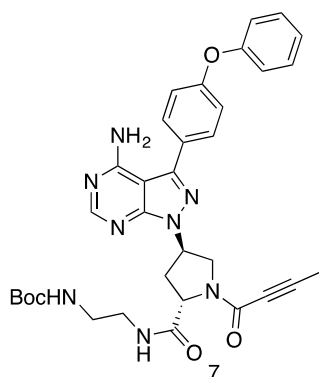
Compound 6.



Compound **6** was synthesized with a similar procedure as compound **4**. Crotonic acid (31 mg, 0.36 mmol), HOBT (25 mg, 0.18 mmol), EDCI (86 mg, 0.45 mmol) and Et₃N (45 mg, 0.45 mmol) were added to the intermediate in 3 ml DCM and stirred for 6 hours. The mixture was treated with similar procedure to give compound **6** as white solids (yield 76%).

¹HNMR (400 MHz, CDCl₃) δ 8.33 (s, 1H), 7.60 (t, *J* = 8.2 Hz, 2H), 7.38 (t, *J* = 7.9 Hz, 2H), 7.19 – 7.05 (m, 5H), 6.99 (dd, *J* = 14.8, 7.0 Hz, 1H), 6.16 (d, *J* = 14.4 Hz, 1H), 5.48 (dd, *J* = 14.7, 7.4 Hz, 1H), 5.27 (d, *J* = 5.6 Hz, 1H), 4.66 (t, *J* = 7.9 Hz, 1H), 4.25 (dt, *J* = 17.8, 10.3 Hz, 2H), 3.34 – 3.19 (m, 2H), 3.10 – 2.92 (m, 2H), 2.89 – 2.54 (m, 3H), 1.88 (dd, *J* = 6.9, 1.4 Hz, 3H), 1.37 (s, 9H). ¹³CNMR (101 MHz, CDCl₃) δ 171.22, 166.34, 158.89, 157.71, 156.36, 155.28, 154.60, 145.40 – 145.22, 144.15, 130.07, 127.29, 124.28, 122.35, 119.74, 119.12, 98.83, 79.09, 77.48, 77.16, 76.84, 59.60, 54.73, 50.96, 40.16, 33.31, 28.49, 18.35. HRMS (*m/z*): [M+H]⁺ calcd. for C₃₃H₃₉N₈O₅⁺, 627.3038; found: 627.3071.

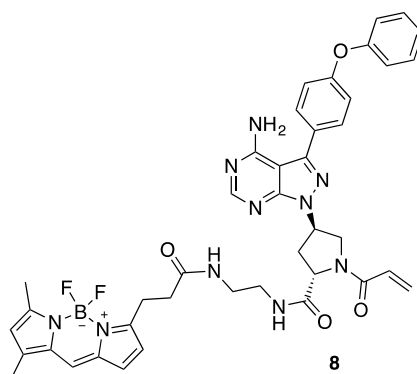
Compound 7.



Compound **7** was synthesized with a similar procedure as compound **4**. Butynoic acid (18.5 mg, 0.22 mmol) HOBt (25 mg, 0.18 mmol), EDCI (86 mg, 0.45 mmol) and Et₃N (45 mg, 0.45 mmol) were added to the intermediate in 3 ml DCM and stirred for 6 hours. The mixture was treated with similar procedure to give compound **7** as white solids (yield 85%).

¹HNMR (400 MHz, CDCl₃) δ 8.34 (s, 1H), 7.61 (dd, *J* = 12.2, 5.4 Hz, 2H), 7.43 – 7.36 (m, 2H), 7.21 – 7.11 (m, 3H), 7.11 – 7.04 (m, 2H), 6.21 (s, 2H), 5.53 – 5.42 (m, 1H), 5.05 (s, 1H), 4.61 (t, *J* = 7.9 Hz, 1H), 4.47 (dd, *J* = 11.1, 7.3 Hz, 1H), 4.27 (dd, *J* = 11.1, 8.1 Hz, 1H), 3.34 – 2.94 (m, 7H), 2.84 – 2.68 (m, 1H), 1.99 (s, 3H), 1.40 (s, 9H). ¹³CNMR (101 MHz, CDCl₃) δ 170.38, 159.09, 157.41, 156.32, 154.99, 154.41, 144.81, 130.29 – 129.90, 127.04, 124.39, 119.82, 119.14, 98.74, 90.44, 79.52, 77.48, 77.16, 76.84, 73.72, 61.91, 58.87, 53.96, 52.30, 50.07, 40.47, 40.21, 36.29, 33.58, 29.71, 29.40, 28.48, 22.81, 4.09. HRMS (*m/z*): [M+H]⁺ calcd. for C₃₃H₃₇N₈O₅⁺, 625.2881; found: 625.2918.

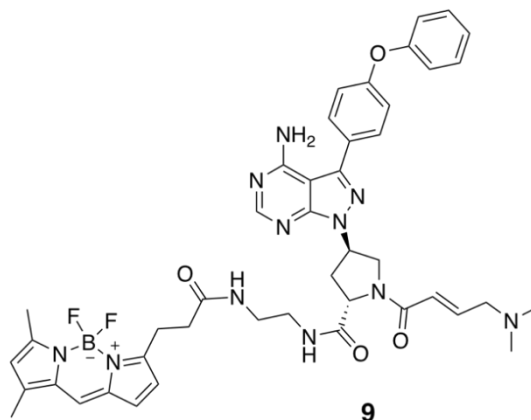
Compound 8.



Trifluoroacetic acid (35 mg, 0.31 mmol) was added to compound **4** (15 mg, 0.017 mmol) dissolved in 1 ml anhydrous DCM. The reaction was stirred at room temperature for 3 hours. The mixture was concentrated (theoretical: 8.8 mg, 0.017 mmol) and dissolved in 2 ml DMF which was directly used in the next reaction. HATU (19.4 mg, 0.051 mmol), BODIPY®FL (5 mg, 0.017 mmol) and DIPEA (11 mg, 15 μ l, 0.085 mmol) were added to the intermediate at 0°C under argon atmosphere. The reaction was stirred for 6 hours at room temperature. The mixture was concentrated in vacuo and extracted with ethyl acetate and saturated NaCl solution three times. The organic phase was dried over anhydrous Na₂SO₄, filtered and concentrated in vacuo. The mixture was purified by HPLC (gradient elution of acetonitrile from 50% to 95% in 15min) and lyophilized to give red solids (yield 42%, purity 93%).

¹H NMR (500 MHz, CDCl₃) δ 8.34 (s, 1H), 7.61 (d, J = 8.6 Hz, 2H), 7.42 – 7.34 (m, 2H), 7.18 (t, J = 7.4 Hz, 2H), 7.13 (d, J = 8.6 Hz, 2H), 7.08 (dd, J = 9.9, 2.2 Hz, 3H), 6.89 (d, J = 4.0 Hz, 1H), 6.46 (d, J = 14.2 Hz, 1H), 6.42 – 6.37 (m, 2H), 6.30 (d, J = 4.0 Hz, 1H), 6.08 (s, 1H), 5.80 – 5.73 (m, 1H), 5.70 (dd, J = 8.8, 3.4 Hz, 1H), 5.56 (s, 2H), 4.86 (t, J = 6.2 Hz, 1H), 4.17 (ddd, J = 16.6, 10.3, 6.8 Hz, 2H), 3.40 (dd, J = 12.7, 9.8 Hz, 3H), 3.27 (t, J = 7.4 Hz, 3H), 2.79 (t, J = 6.8 Hz, 2H), 2.72 – 2.59 (m, 2H), 2.52 (s, 3H), 2.23 (d, J = 8.3 Hz, 3H). ¹³C NMR (101 MHz, CDCl₃) δ 172.56, 171.56, 171.21, 165.98, 165.98, 160.25, 158.61, 157.78, 157.63, 156.03, 155.98, 154.34, 144.35, 143.68, 141.06, 138.74, 135.05, 133.40, 130.14, 130.14, 130.04, 129.50, 129.43, 128.46, 128.46, 127.95, 127.77, 127.10, 124.29, 124.11, 123.94, 123.79, 120.45, 119.76, 119.76, 119.23, 119.15, 117.50, 98.53, 77.48, 77.48, 77.37, 77.16, 77.16, 76.84, 76.84, 59.83, 59.37, 55.13, 54.73, 51.52, 51.41, 40.22, 39.79, 39.24, 38.80, 35.80, 35.75, 33.10, 32.81, 24.94, 24.87, 14.88, 11.43, 11.27, 1.28, 1.16, -0.04. HRMS (m/z): [M+H]⁺ calcd. for C₄₁H₄₂BF₂N₁₀O₄⁺, 787.3446; found: 787.3443.

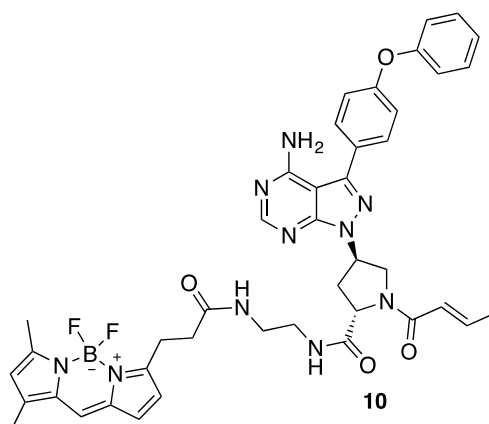
Compound 9.



Compound **9** was synthesized with a similar procedure as compound **8**. Red solids (yield 30%, purity 98.5%).

^1H NMR (500 MHz, CDCl_3) δ 11.69 (s, 2H), 8.20 (s, 2H), 7.53 (t, $J = 7.8$ Hz, 4H), 7.41 (t, $J = 7.8$ Hz, 4H), 7.21 (d, $J = 7.3$ Hz, 2H), 7.09 (dd, $J = 23.7, 7.6$ Hz, 9H), 6.86 (s, 4H), 6.71 (d, $J = 13.5$ Hz, 1H), 6.58 (s, 1H), 6.24 (d, $J = 21.2$ Hz, 3H), 6.11 (s, 2H), 5.49 (s, 1H), 4.66 (s, 3H), 4.30 (s, 5H), 3.98 (s, 2H), 3.81 (s, 3H), 3.71 (s, 2H), 3.43 (s, 2H), 3.30 (s, 6H), 3.14 (s, 5H), 2.91 (d, $J = 29.7$ Hz, 3H), 2.85 (s, 4H), 2.78 (s, 9H), 2.60 – 2.38 (m, 10H), 2.24 (s, 6H). ^{13}C NMR (126 MHz, CDCl_3) δ 160.53, 159.99, 157.33, 157.14, 155.90, 153.91, 152.39, 147.18, 146.51, 146.42, 144.57, 135.39, 133.38, 132.09, 131.73, 130.58, 130.30, 129.96, 128.49, 125.19, 124.74, 124.05, 120.81, 120.02, 119.32, 116.91, 97.78, 97.73, 77.41, 77.16, 76.91, 59.81, 59.29, 57.37, 55.52, 54.17, 51.04, 42.71, 42.66, 42.55, 40.34, 38.98, 36.12, 35.13, 34.95, 33.62, 24.65, 15.05, 11.40. HRMS (m/z): $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{44}\text{H}_{49}\text{BF}_2\text{N}_{11}\text{O}_4^+$, 844.4025; found: 844.4027.

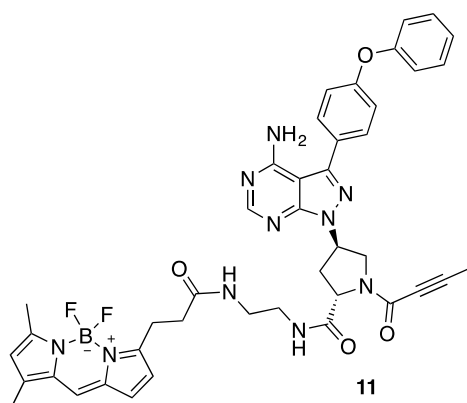
Compound 10.



Compound **10** was synthesized with a similar procedure as compound **8**. Red solids (yield 25%, purity 98%).

^1H NMR (400 MHz, CDCl_3) δ 11.66 (s, 1H), 8.10 (d, $J = 13.8$ Hz, 1H), 7.52 (t, $J = 9.1$ Hz, 2H), 7.42 (t, $J = 7.9$ Hz, 3H), 7.22 (t, $J = 7.4$ Hz, 1H), 7.15 (d, $J = 8.5$ Hz, 2H), 7.12 – 7.06 (m, 3H), 7.06 – 6.93 (m, 1H), 6.90 (d, $J = 3.8$ Hz, 1H), 6.70 (s, 1H), 6.27 (d, $J = 3.9$ Hz, 2H), 6.15 – 6.06 (m, 2H), 5.86 – 5.76 (m, 1H), 4.87 (s, 2H), 4.17 (ddd, $J = 27.7, 18.6, 11.7$ Hz, 3H), 3.42 (t, $J = 19.5$ Hz, 3H), 3.24 (d, $J = 25.3$ Hz, 3H), 2.82 – 2.60 (m, 4H), 2.48 (s, 3H), 2.24 (s, 3H), 1.85 (d, $J = 6.3$ Hz, 3H), 1.28 (dt, $J = 14.3, 5.5$ Hz, 1H). ^{13}C NMR (101 MHz, CDCl_3) δ 159.76, 155.43, 153.32, 151.92, 147.19, 145.95, 144.57, 144.16, 135.06, 133.04, 129.99, 129.46, 128.11, 124.63, 124.49, 123.82, 120.35, 119.78, 119.02, 116.74, 97.03, 77.16, 77.05, 76.85, 76.53, 59.13, 55.24, 50.72, 39.73, 39.35, 35.19, 32.91, 24.46, 18.12, 14.72, 11.11, -0.18. HRMS (m/z): $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{42}\text{H}_{44}\text{BF}_2\text{N}_{10}\text{O}_4^+$, 801.3603; found: 801.3657.

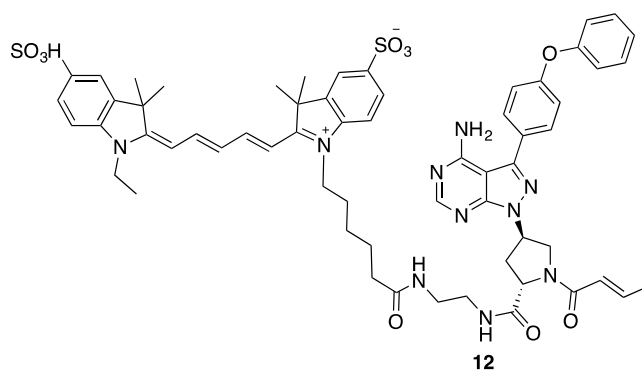
Compound 11.



Compound **11** was synthesized with a similar procedure as compound **8**. Red solids (yield 25%, purity 94%).

^1H NMR (400 MHz, CDCl_3) δ 8.14 (s, 14H), 7.59 – 7.50 (m, 42H), 7.42 (t, $J = 8.0$ Hz, 42H), 7.22 (s, 28H), 7.18 – 7.12 (m, 44H), 7.09 (d, $J = 7.3$ Hz, 55H), 6.90 (d, $J = 4.0$ Hz, 19H), 6.36 (s, 17H), 6.30 (d, $J = 3.8$ Hz, 19H), 6.26 – 6.16 (m, 25H), 6.09 (s, 13H), 5.82 – 5.69 (m, 19H), 4.81 (d, $J = 7.8$ Hz, 17H), 4.23 (d, $J = 5.8$ Hz, 40H), 3.40 (s, 62H), 3.31 – 3.12 (m, 93H), 2.87 (d, $J = 15.8$ Hz, 55H), 2.76 – 2.57 (m, 111H), 2.51 (d, $J = 12.1$ Hz, 75H), 2.24 (s, 68H), 1.97 (d, $J = 13.6$ Hz, 58H). ^{13}C NMR (101 MHz, CDCl_3) δ 160.06, 155.78, 153.86, 152.15, 147.46, 146.39, 144.04, 135.42, 135.05, 133.40, 122.14, 117.13, 97.38, 90.77, 73.58, 64.39, 58.93, 55.04, 52.88, 40.26, 39.43, 35.64, 33.76, 24.85, 21.17, 15.04, 11.44, 5.26, 4.16, 0.14. HRMS (m/z): $[\text{M}+\text{H}]^+$ calcd. for $\text{C}_{42}\text{H}_{42}\text{BF}_2\text{N}_{10}\text{O}_4^+$, 799.3446; found: 799.3449.

Compound 12.



Trifluoroacetic acid (35 mg, 0.31 mmol) was added to compound **6** (13 mg, 0.02 mmol) dissolved in 1 ml anhydrous DCM. The reaction was stirred at room temperature for 3 hours. The mixture was concentrated (theoretical: 9 mg, 0.02 mmol) and dissolved in 2 ml DMF which was directly used in the next reaction. TSTU (3.6 mg, 0.012 mmol), compound Cy5 (5 mg, 0.0075 mmol) and Et₃N (7 μ l, 0.051 mmol) were added to the intermediate at 0°C under argon atmosphere. The reaction was stirred overnight at room temperature. The reaction was monitored by HPLC. Petroleum ether (5 ml) was added to the reaction. The precipitate was dissolved in DMSO and purified by HPLC (gradient elution of acetonitrile from 30% to 95% in 30min) and lyophilized to give blue solids (yield 38%, purity 93%).

¹H NMR (500 MHz, DMSO) δ 8.35 (t, J = 12.9 Hz, 17H), 8.26 (t, J = 5.2 Hz, 11H), 7.81 (s, 22H), 7.64 (dd, J = 6.5, 2.1 Hz, 36H), 7.46 – 7.39 (m, 25H), 7.33 – 7.23 (m, 33H), 7.22 – 7.04 (m, 75H), 6.74 – 6.62 (m, 12H), 6.57 (t, J = 12.3 Hz, 9H), 6.30 (dd, J = 13.6, 9.6 Hz, 22H), 5.58 (dd, J = 21.4, 14.8 Hz, 8H), 4.62 (dd, J = 8.3, 4.8 Hz, 7H), 4.22 – 3.97 (m, 48H), 3.97 – 3.83 (m, 10H), 3.16 – 3.03 (m, 54H), 2.05 (dd, J = 12.7, 5.4 Hz, 19H), 1.84 – 1.73 (m, 32H), 1.68 (s, 107H), 1.52 (dd, J = 13.9, 6.5 Hz, 22H), 1.26 – 1.20 (m, 62H). ¹³C NMR (126 MHz, DMSO) δ 173.20, 172.38, 172.05, 171.23, 163.98, 157.88, 157.17, 156.17, 155.64, 154.21, 145.37, 143.54, 141.85, 141.38, 140.53, 140.38, 130.02, 129.94, 128.38, 127.68, 127.40, 126.03, 125.55, 123.72, 123.27, 122.46, 119.86, 118.90, 118.87, 110.02, 103.43, 103.06, 97.45, 59.11, 58.85, 54.31, 50.89, 48.82, 48.78, 45.66, 40.02, 39.85, 39.69, 39.52, 39.35, 39.19, 39.02, 38.58, 38.50, 38.38, 38.03, 36.16, 35.67, 35.10, 34.07, 31.03, 30.69, 28.97, 28.88, 28.84, 28.69, 28.54, 28.45, 27.85, 27.02, 26.91, 26.55, 26.45, 25.63, 24.99, 24.67, 21.94, 17.49, 13.79, 13.70, 11.96, 8.47, 0.68. HRMS (m/z): [M+H]⁺ calcd. for C₆₁H₆₉N₁₀O₁₀S₂⁺, 1165.4640; found: 1165.4637.

III. Biology

General Information

All the fluorescence intensity analysis was recorded by Perkin Elmer Envision 2104 Multilabel Reader (Em =450 nm) using striwellTM microplate (costar 42592).

All the fluorescence gel scanning was conducted with PharasFXTM Plus Molecular Imager (Bio-Rad, Ex = 480nm and Em = 530nm, Ex = 635nm and Em =695nm) using Quantity OneTM software.

Experimental Procedures

The detection efficiency of probe in cell samples.

Sample A and B of Figure 3b: 3×10^6 /ml OCI-Ly7 cells were grown in six-well plates with probe **10** (1 μ M) or probe **12** (1 μ M) for 1 hour. Then DMSO (5 μ l) was added into each well for 1 hour. All the cells were harvested.

Sample C and D of Figure 3b: 3×10^6 /ml OCI-Ly7 cells were grown in six-well plates with probe **10** (1 μ M) or DMSO for 1 hour. Then the lysate labelled by probe 12 (16 μ M) for 1 hour either.

All the samples were heated at 100°C for 10 min and loaded on SDS-page gels (10%), and the gels were treated with the same fluorescence scanning and western blot steps as the probe labelling experiment. (Figure 3b)

Detection of the residual Btk in cell sample which have undergone ELISA detection.

The cell samples were harvested after the ELISA detection. After heating at 100°C for 10 minutes, the denatured samples were loaded on SDS-page gels (10%), and was scanned for fluorescence signals, and also by western blotting. (Figure 3c)

RT-qPCR experiment

20×10^3 Ramos and OCI-Ly7 cells were incubated in 1ml of RPMI 1640 medium containing 9% fetal bovine serum and activated with 200 μ g/ml IgM (R&D G-105-C) for 0h, 1h, 2h, 4h, 9h, 12h and 48h. Total RNA was extracted from each group using TRIzol reagent according to the manufacturer's instructions (ambion 15596026). The RNA quality was assessed by determining the ratio of spectrophotometric absorbance of the sample at 260nm to that of 280nm using Nanodrop 2000. All samples had an A260/A280 absorption ratio > 1.6. cDNA was reverse transcribed from RNA using Revert Aid First Strand cDNA Synthesis kit (Thermo scientific #K1621). Primers for RT-qPCR assays of Btk were from sanggo biotech (9406063704 forward and 9406063705 reverse). Primers of β -actin used were from BBI life sciences (BN61102-0001 forward and BN61102-0002 reverse). All RT-qPCR reactions containing 2 μ l cDNA were performed using the FastStar Universal SYBR Green Master (Rox) (Roche 04913914001) on BIO-RAD CFX Connect™ real time system which was programmed as 50°C for 2min, 95°C for 10min, followed by 40 cycles at 90°C

for 10s, 60°C for 30s, finally subjecting to the melting temperature to check the amplification curve.
(Supporting Figure 7)

IV. ^1H NMR, ^{13}C NMR and HRMS Spectra of Compounds

