Quantitative proteomics and cytology of rice pollen sterol-rich

membrane domains reveals pre-established cell polarity cues in

mature pollen

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This supporting information includes 8 figures and 3 tables.

NO.	CAPTION						
1	Figure S1. Flotillin-like protein antibody preparation						
2	Figure S2. Distribution of sterols in developing pollen						
3	Figure S3. Work flow used to identify sterol-rich membrane microdomain-associated proteins of mature rice pollen						
4	Figure S4. SDS-PAGE separation of proteins from rice pollen DRM-Ls						
5	Figure S5. Representative images for protein identification and quantification with ESI-Qq-TOF mass spectrometer						
6	Figure S6. Comparisons of the chromatograph profiles among these duplicates (from analysis group 1 to 10)						
7	Figure S7. MS/MS fragmentation spectra of peptides, which led to identification of a protein by only itself						
8	Figure S8. Volcano plot depicting the quantification results of label-free peptides						
9	Table S1. Quantification and annotation of all identified proteins						
10	Table S2. List of 237 proteins of the sterol dependent protein dataset						
11	Table S3. Statistical analysis of gold particle distribution in developing rice pollen						



MGQCLGLVQIDQSTVAIKENFGKFSEVLEPGCHFLPWCIGQQIAGYLSLRVKQLDVRCET KTKDNVFVTVVASVQYRALADKASDAFYKLSNTREQIQSYVFDVIRATVPKLNLDDAFEQ KNDIAKAVEDELEKAMSAYGYEIVQTLIIDIEPDVHVKRAMNEINAAARLRVAANEKAEAEK ILQIKKAEGEAESKYLAGVGIARQRQAIVDGLRDSVLAFSENVPGTTAKDIMDMVLVTQYF DTMKEIGASSKSTSVFIPHGPGAVKDVAAQIRDGLLQANAERND



Figure S1. Flotillin-like protein antibody preparation. (a) Sequence of the flotillin-like protein (band_7 protein, BAD23328) and synthetic antigen. **(b)** Antigen prediction results (the software 'Epitope Adviser Lite' was developed by MBL, Woburn, MA, USA). **(c)** Antibody specificity testing in differential centrifugation products from rice pollen lysate. EC, entire cell lysate; P, pellet; S, supernatant; M, protein molecular weight marker.



Figure S2. Distribution of sterols in developing pollen. Sterols were labeled with the fluorescent molecule, filipin. Pseudocolor images indicating the intensity of filipin-sterol fluorescence. Color bar represents the relative intensities from 0 to 255. PMC, pollen mother cell; MS, microspore; BCP, bicellular pollen; TCP, immature tricellular pollen. Arrow: aperture. Arrow head: pollen tube. Bars = 10µm.



Figure S3. Work flow used to identify sterol-rich membrane microdomain-associated proteins of mature rice pollen. Five replicates were carried out in parallel. EC, entire cell lysate; DRM-L, detergent insoluble membrane-low density fraction; DRM-H, detergent insoluble membrane-high density fraction. Gn (n=1-10), group number.



Figure S4. SDS-PAGE separation of proteins from rice pollen DRM-Ls. Proteins were resolved on 4.5% stacking gel and 12.5% separation gel with a gel thickness of 1mm. Proteins were stained in-gel with Coomassie blue. The whole gel was cut into 100 samples along the dotted line, then each sample was cut into 1 cubic millimeter (mm³) pieces for in-gel tryptic digestion. The 10 samples between the two red dotted lines belong to 1 independent analysis group. Gn (n=1-10), group number; C, control; T, treatment; M, protein molecular weight marker.



Figure S5. Representative images for protein identification and quantification with ESI-Qq-TOF mass spectrometer. (a) Basic Peak Chromatogram of group 6. (b) Extracted Ion Chromatogram of three peptides matched to gi|31126785. (c) A representative MS/MS fragmentation spectra. In (a) and (b), the blue line indicates signals in control samples; the red line indicates signals of MβCD treated samples.





















The control sample of replicate #4 shows an approximate 20 minute delay, so this replicate was excluded from quantitative analysis.





















Figure S6. Comparisons of the chromatograph profiles among these duplicates (from analysis group 1 to 10).



























































































































Cmpd 5791, +MSn(855.4486), 75.3 min [Cmpd 5791, +MSn(855.4486), 75.3 min]





Figure S7. MS/MS fragmentation spectra of peptides, which led to identification of a protein by only itself.



Figure S8. Volcano plot depicting the quantification results of label-free peptides. The red dotted line indicates the mean of lg(C/T); the black dotted line indicates 2 times the standard deviation (S.D.) of the lg(C/T) distance from the mean value. The calculation was based on RMS90 (root mean square, 90 percent confidence) peptides.

Table S3. Statistical analysis of gold particle distribution in developing rice pollen. A. Statistical analysis data from 5 independent experiments showing that lipid raft exist in rice pollen mother cells. B. Statistical analysis data from 5 independent experiments showing that lipid rafts migrate from vesicles to the plasma membrane as pollen develop. Duncan grouping shows significant analysis.

Α

	Total number of g	Number of clustered gold particles			Number of scattered gold particles				
	Mean ± S.D.	Duncan grouping			Duncan grouping			Duncan grouping	
		α = 0.05	α = 0.01	Mean ± S.D.	α = 0.05	α = 0.01	Mean ± S.D.	α = 0.05	α = 0.01
Mature Pollen (n=5)	167 ± 17.85	а	А	117 ±10.83	а	А	50 ± 7.37	а	А
Tetrad (n=5)	113 ± 8.88	b	В	67± 5.86	b	В	46 ± 5.29	ab	AB
Bicellular pollen (n=5)	88 ± 8.53	с	С	47 ±4.74	с	С	41 ± 5.68	bc	BC
Microspore (n=5)	60 ± 2.86	d	D	24 ±2.70	d	D	36 ± 1.30	С	С
Pollen mother cell (n=5)	20 ± 3.78	е	E	3 ± 0.45	е	E	17 ± 3.51	d	D

	Total number of clustered gold particles			Number of clustered gold particles in plasma membrane			Number of clustered gold particles in vesicles		
		Duncan grouping			Duncan grouping			Duncan grouping	
	Mean ± S.D.	α = 0.05	α = 0.01	Mean ± S.D.	α = 0.05	α = 0.01	Mean ± S.D.	α = 0.05	α=
									0.01
Mature Pollen (n=5)	117 ± 10.83	а	A	77 ± 10.94	а	А	40 ± 3.54	а	A
Tetrad (n=5)	67 ± 5.86	b	В	28 ± 3.19	b	В	39 ± 4.91	а	A
Bicellular pollen (n=5)	47 ± 4.74	с	С	9 ± 2.68	с	С	38 ± 5.17	а	A
Microspore n=5)	24 ± 2.70	d	D	0	d	С	24 ± 2.70	b	В
Pollen mother cell (n=5)	3 ± 0.45	е	E	0	d	С	3 ± 0.45	С	С