

1 Diversity of Sporosarcina-Like Bacterial Strains Obtained from Meter-Scale Augmented and
2 Stimulated Bio-cementation Experiments

3
4 Charles M. R. Graddy¹, Michael G. Gomez², Lindsay M. Kline¹, Sydney R. Morrill¹, Jason T.
5 DeJong² and Douglas C. Nelson^{1*}

6
7
8 University of California, Davis, CA, 95616, USA

9
10 ¹Department of Microbiology and Molecular Genetics

11 ²Department of Civil and Environmental Engineering

12
13
14
15 Corresponding Author:

16 Douglas C. Nelson

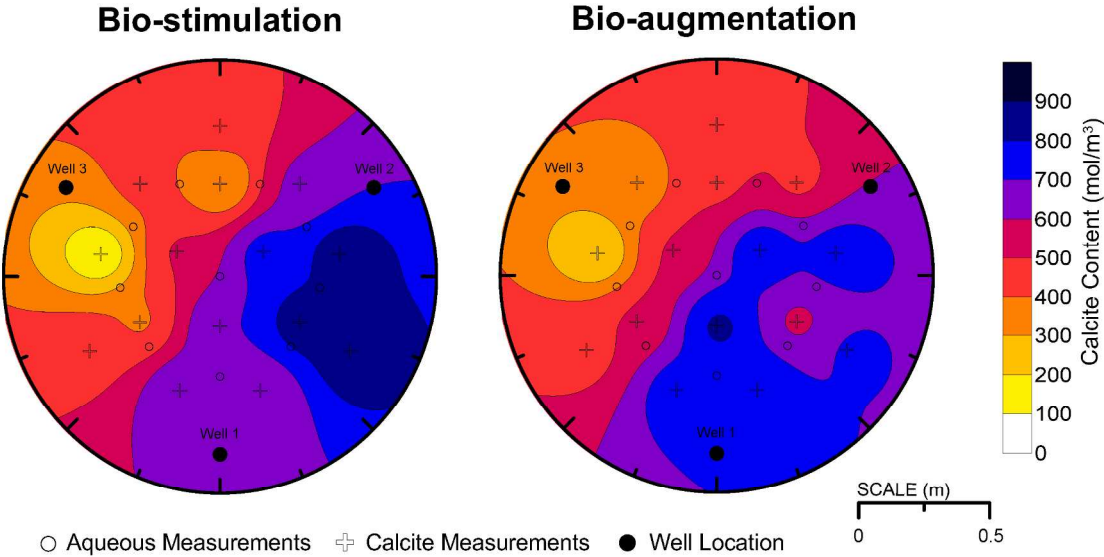
17 University of California, Department of Microbiology and Molecular Genetics; CA, 95616,
18 USA, email: dcnelson@ucdavis.edu

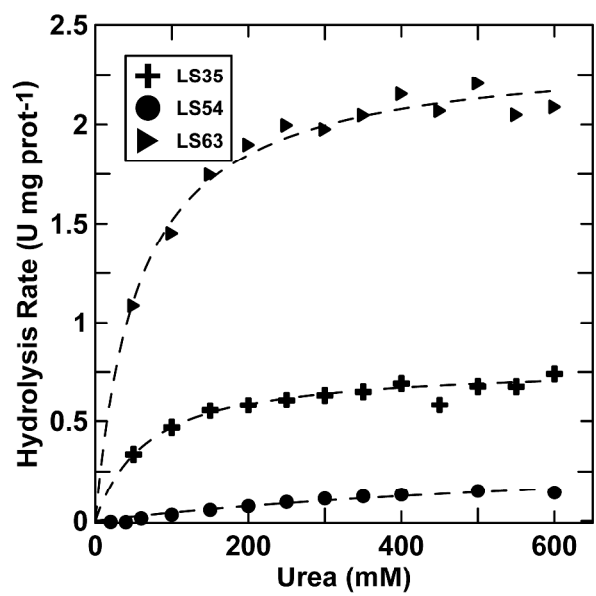
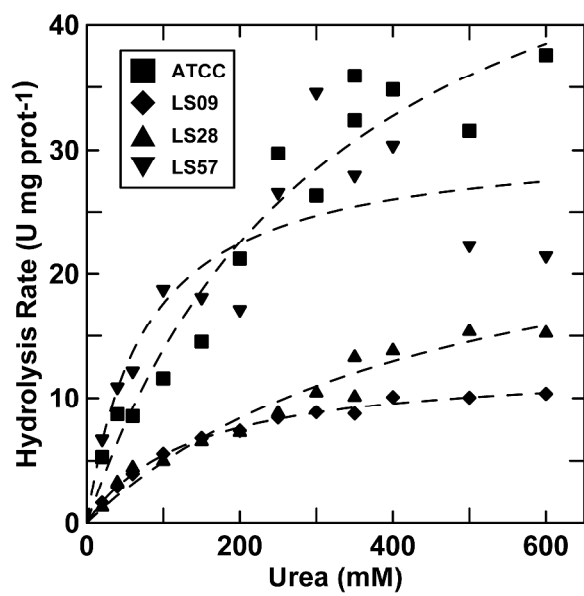
19
20 Pages: 6

21 Figures: 2

22 Tables: 2

Supplemental Figure 1.





Supplemental Figure 2.

Supplemental Table 1. Isolate List and Sample Source

Isolate	Tank	Day	Sample	Log		Urease	Closest Homologous Strain	Bases
		Isolated	Location	Dilution	Sequenced			
LS01	Stim	1	D	-2	+	Sporosarcina soli (T) I80	836	
LS02	Stim	1	D	-2	+	Sporosarcina soli (T) I80	1424	
LS03	Stim	1	D	-2	+	Sporosarcina soli (T) I80	1411	
LS07	Stim	1	G	-2	+	Oceanobacillus polygoni (T) SA9	902	
LS09	Stim	1	G	-2	+	Sporosarcina pasteurii WJ-5	1387	
LS10	Stim	1	J	-2	+	Bacillus lentus UR41	1405	
LS14	Stim	3	A	-4	+	Sporosarcina soli KNUC401	1421	
LS15	Stim	3	D	-4	+	Sporosarcina soli KNUC401	1421	
LS17	Stim	3	G	-5	+	Sporosarcina soli KNUC401	1416	
LS18	Stim	3	G	-5	+	Sporosarcina soli KNUC401	1415	
LS19	Stim	5	A	-3	+	Sporosarcina soli KNUC401	843	
LS20	Stim	5	B	-4	+	Sporosarcina soli KNUC401	1418	
LS21	Stim	5	B	-3	+	Sporosarcina soli KNUC401	1426	
LS22	Stim	5	D	-4	+	Sporosarcina soli KNUC401	897	
LS23	Stim	5	D	-3	+	Sporosarcina soli KNUC401	904	
LS24	Stim	5	E	-3	+	Sporosarcina soli KNUC401	1423	
LS25	Stim	5	E	-3	+	Sporosarcina soli KNUC401	1425	
LS26	Stim	5	G	-3	+	Sporosarcina soli KNUC401	1425	
LS27	Stim	5	J	-4	+	Sporosarcina soli KNUC401	912	
LS28	Stim	5	J	-3	+	Sporosarcina soli KNUC401	1324	
LS33	Stim	12	A	-3	+	Sporosarcina ginsengisoli Gsoil1433	839	
LS34	Stim	12	B	-3	+	Sporosarcina ginsengisoli Gsoil1433	995	
LS35	Stim	12	E	-1	+	Bacillus lentus (T) NCIMB8773	845	
LS36	Stim	12	E	-1	+	Sporosarcina soli KNUC401	1423	
LS37	Stim	12	G	-3	+	Bacillus lentus (T) NCIMB8773	948	
LS38	Stim	12	G	-2	neg.	Oceanobacillus luteolus WM-4	1437	
LS39	Stim	12	J	-1	+	Sporosarcina soli KNUC401	1446	
LS40	Stim	12	J	-3	+	Bacillus hackensackii AY14842	1289	
LS41	Stim	12	Well 2	-1	+	Sporosarcina soli KNUC401	1410	
LS45	Stim	12	Well 2	-1	+	Sporosarcina saromensis KUDC1822	915	
LS46	Stim	12	Well 2	-3	+	Sporosarcina luteola (T) Y1	955	
LS47	Stim	12	Well 2	-3	neg.	Sporosarcina aquimarina KUDC1821	921	
LS48	Stim	12	A	-2	+	Sporosarcina pasteurii IARI-J-21	1430	
LS49	Stim	13	B	-3	+	Oceanobacillus luteolus (T) WM-1	1396	
LS29	Aug	5	A	-3	+	Sporosarcina pasteurii (T) HQ676600	1095	
LS31	Aug	5	G	-3	+	Sporosarcina pasteurii (T) HQ676600	1099	
LS32	Aug	5	J	-3	+	Sporosarcina pasteurii (T) HQ676600	1096	
LS70	Aug	5	E	-3	+	Sporosarcina pasteurii (T) HQ676600	1095	
LS71	Aug	5	D	-4	+	Sporosarcina pasteurii (T) HQ676600	1089	
LS72	Aug	5	D	-4	+	Sporosarcina pasteurii (T) HQ676600	1093	
LS73	Aug	5	G	-4	+	Sporosarcina pasteurii (T) HQ676600	1093	
LS52	Aug	12	A	-3	+	Sporosarcina soli (T) I80	1426	
LS53	Aug	12	B	-3	neg.	Ornithinibacillus contaminans (T) CCUG 53201	1443	
LS54	Aug	12	B	-2	+	Sporosarcina aquimarina FT1	1425	
LS55	Aug	12	G	-1	+	Sporosarcina pasteurii WJ-5	1424	
LS56	Aug	12	D	-2	+	Sporosarcina soli KNUC401	1426	
LS57	Aug	12	E	-3	+	Sporosarcina soli KNUC401	1424	
LS58	Aug	12	E	-2	+	Sporosarcina soli KNUC401	1422	
LS61	Aug	12	G	-2	+	Sporosarcina soli KNUC401	1410	
LS62	Aug	12	J	-2	+	Sporosarcina soli (T) I80	1422	
LS63	Aug	13	A	-3	+	Sporosarcina soli (T) I80	1423	
LS64	Aug	13	A	-3	+	Sporosarcina soli (T) I80	1425	
LS65	Aug	13	B	-3	neg.	Oceanobacillus luteolus (T) WM-1	1435	
LS66	Aug	13	D	-3	+	Sporosarcina soli (T) I80	1426	
LS67	Aug	13	E	-3	+	Oceanobacillus luteolus WM-4	1438	
LS68	Aug	13	G	-3	neg.	Oceanobacillus luteolus WM-4	1434	
LS69	Aug	13	J	-3	+	Sporosarcina soli (T) I80	1424	

Supplemental Table 2: Alkalinity and Carbon Dioxide Generating Microbial Processes*

Process	Substrate	Product	Representatives	Specific activity (substrate) (U mg prot ⁻¹)	OH- production** (μmol min ⁻¹ mg prot ⁻¹)	CO ₂ production** (μmol min ⁻¹ mg prot ⁻¹)
Denitrification ⁴¹	NO ₃ ⁻	0.5 N ₂	<i>Paracoccus denitrificans</i> <i>Pseudomonas stutzeri</i>	0.0764 0.1712	0.0764 - 0.171	0.096 - 0.214
Sulfate Reduction ⁴²	SO ₄ ²⁻	H ₂ S	<i>Desulfovibrio desulfuricans</i> CSN <i>D. vulgaris</i> Marburg <i>Desulfobulbus pripionicus</i> <i>Desulfobacter hydrogenophilus</i>	0.0285 0.041 0.061 0.036	0.057 - 0.122	0.057 - 0.122
Iron Reduction ⁴³	Fe(III)	Fe(II)	<i>Geobacter metallireducens</i> <i>Desulfuromonas acetoxidans</i> <i>Pseudomonas sp.</i> <i>Shewanella putrefaciens</i> MR-1 <i>Shewanella putrefaciens</i> MR-4 <i>Shewanella putrefaciens</i> sp200 strain BrY	0.210 - 0.870 0.414 - 0.570 0.175 0.0098 - 0.833 0.478 - 0.510 0.250 - 0.310 0.46	0.0196 - 1.74	0.0024 - 0.218
Ureolysis	urea	2NH ₄ ⁺ CO ₃ ²⁻	<i>Sporosarcina soli</i> LS28** <i>S. pasteurii</i> LS09** <i>Bacillus hackensackii</i> LS35** <i>S. aquimarina</i> LS54** <i>S. pasteurii</i> ATCC 11859**	12 9.26 0.662 0.115 30.7	0.230 - 24 61.4	0.115 - 12 30.7

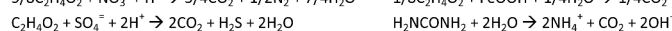
*For denitrifying, sulfate-reducing and iron-reducing bacteria, whole cell rates (μmol min⁻¹ mg prot⁻¹) are based on pure culture literature maxima and ranges. Literature rates for denitrification derived assuming protein = 50% of total dry weight. Whole cell ureolysis rates are based on kinetic parameters for Table 5 data at 350 mM urea, pH 8.5. Ranges shown are for 4 tank isolates; *S. pasteurii* ATCC 11859 data shown separately.

**Rates of OH⁻ production (or H⁺ consumption) and CO₂ production for tabulated processes are derived from balanced equations shown below, with acetate as electron donor for respirations, i.e. first 3 reactions. The derived ratios are:

OH⁻/substrate = 1:1 for denitrification; 2:1 for iron reduction, sulfate reduction, and ureolysis.

CO₂/substrate = 1.25:1 for denitrification; 2:1 for sulfate reduction; 0.25:1 for iron reduction; 1:1 for ureolysis.

Reaction Equations:⁴⁴



28 **Figure S1.** Mid-tank height calcite contours (mol/m^3) at mid-height in each tank at the end of
 29 cementation phase. Both tanks were dissected and core tubes were driven at 15 locations (shown
 30 here as + symbols) to collect soil samples for calcite quantification by a pressure chamber
 31 method that employed an acid dissolution reaction.⁵⁶ Resulting calcite values were inverse-
 32 distance interpolated to generate these spatial contours of final mid-depth calcite content.

33 **Figure S2.** Urea hydrolysis rates over a range of urea concentrations at pH 8.5 with Michaelis-
 34 Menten kinetic fits. Panel (a) shows higher rate strains: ATCC 11859 (type strain of *S. pasteurii*),
 35 LS09 (*S. pasteurii*), LS28 (*S. soli*) and LS57 (*S. soli*). Panel (b) shows data for LS35, LS54 and
 36 LS63 (*B. lentus*, *S. aquimarina* and *S. soli*, respectively).

37 **Table S1.** Isolate List and Sample Source for strains discussed within.

38 **Table S2.** Alkalinity and Carbon Dioxide Generating Microbial Processes