SUPPORTING INFORMATION

Magnetic Oleosome as a Functional Lipophilic Drug Carrier for Cancer Therapy

Hyeon-Yeol Cho,^{†,‡,} # Taek Lee,^{†,§,} # Jinho Yoon,[†] Zhenlin Han,[#] Hudifah Rabie,[‡] Ki-Bum Lee,^{‡,⊥} Wei Wen Su,*[#] and Jeong-Woo Choi*[†]

[†]Department of Chemical & Biomolecular Engineering, Sogang University, Seoul, 04107, Korea
[‡]Department of Chemistry and Chemical Biology, Rutgers, The State University of New Jersey,
Piscataway, New Jersey, 08854, United States
[§]Department of Chemical Engineering, Kwangwoon University, Seoul 01897, Korea
^{II}Department of Molecular Biosciences and Bioengineering, University of Hawaii at Manoa,
Honolulu, Hawaii 96822, United States
^LCollege of Pharmacy, Kyung Hee University, Seoul 02447, Korea

[#] Equal contribution

Prof. Jeong-Woo Choi Email: <u>jwchoi@sogang.ac.kr</u> Tel: (+82) 2-705-8480, Fax: (+82) 2-3273-0331

Prof. Wei Wen Su Email: <u>wsu@hawaii.edu</u> Tel: (+1) 808-956-3531, Fax: (+1) 808-956-3542

TABLE OF CONTENTS

SUPPLEMENTARY FIGURES AND TABLES	Page
Figure S1: Characterization of lipophilic magnetic nanoparticle	S-3
Figure S2: Energy dispersive X-ray spectroscopy (EDS) data of ZnFe ₂ O ₄ and Fe ₃ O ₄	S-4
Figure S3: Confirmation of antibody binding on functional oleosome	S-5
Figure S4: Confocal images of oleosome to normal breast cell and neuroblastoma	S-6
Figure S5: Cytotoxic effects of carmustine on different types of cells	S-7

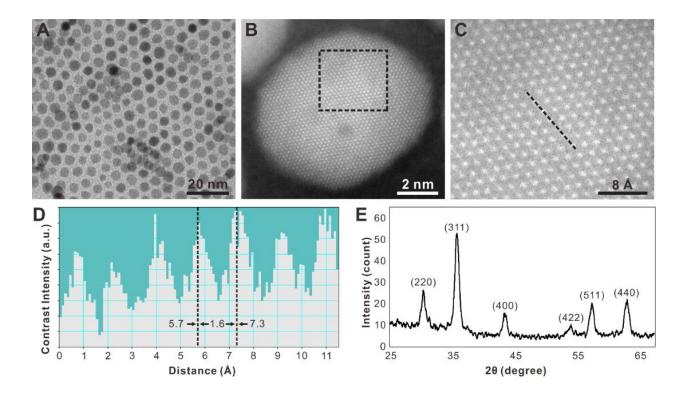


FIGURE S1: Characterization of the lipophilic magnetic nanoparticle. (A-C) STEM images of the magnetic $ZnFe_2O_4$ nanoparticles. (B) The lattice structure of single nanoparticle. (C) a zoomed-in image of the dash-lined box in (B). (D) The distance between lattice projections showing a d-spacing of ~0.16 nm (1.6 Å), corresponding to the (511) lattice spacing of the face-centered cubic spinel. (E) Magnetic nanoparticle x-ray diffraction patterns.

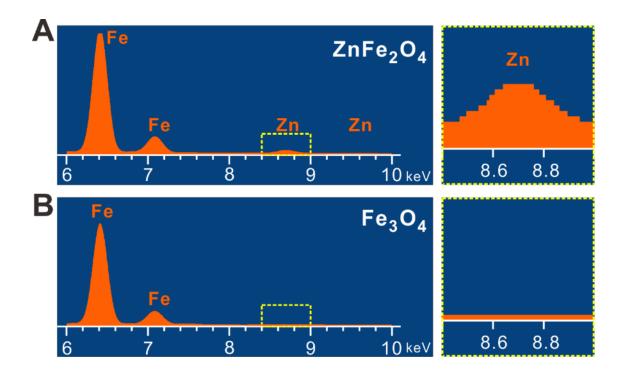


FIGURE S2: Energy dispersive X-ray spectroscopy (EDS) data of $ZnFe_2O_4$ (A) and Fe_3O_4 (B). The right panel is a zoomed-in image of the dash-lined box in (A) and (B).

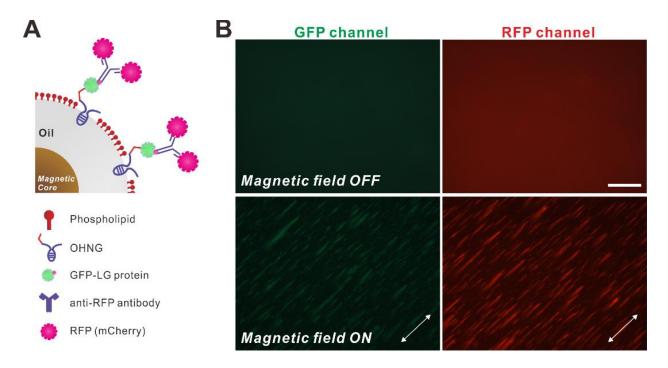


FIGURE S3: Confirmation of antibody binding on functional oleosome. (A) Schematic illustration of red fluorescence protein (RFP, mCherry) decorated oleosome with the anti-RFP antibody. (B) After the sequential treatment of anti-RFP antibody and RFP on the oleosome, the fluorescence signal of RFP was showed the magnetic response and it was well-aligned with GFP signal. The double-headed arrow indicates the direction of the applied magnetic field. Scale bar: 100 μm.

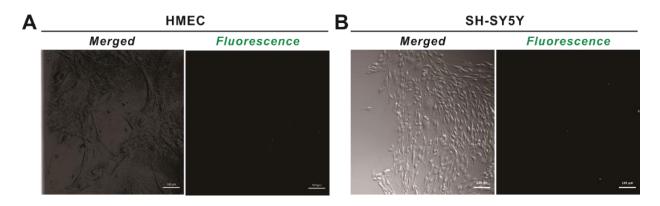


FIGURE S4: Confocal images of oleosome to normal breast cell and neuroblastoma. (A) oleosome-EGFR treated normal cells (HMECs), (B) oleosome-EGFR treated neuroblastoma (SH-SY5Y). Fluorescence images were obtained with FITC filter (495 nm excitation, 520 nm emission). Scale bar is 100 μm.

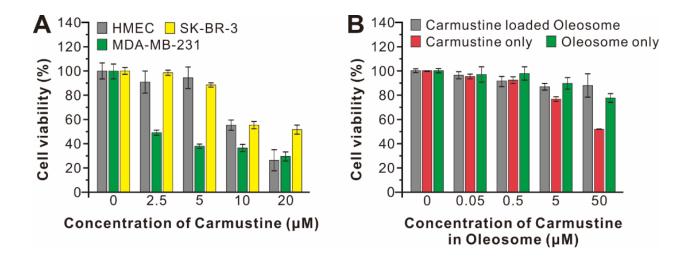


FIGURE S5: Cytotoxic effects of carmustine on different types of cells. (A) The result of cytotoxic effects of carmustine on breast cancer cell lines (MDA-MB-231 and SK-BR-3) and normal cell line (HMEC). 2 mM Carmustine in DMSO was treated with the growth media and the final ratio between DMSO and growth media was matched for every condition by adding an extra volume of DMSO. (B) HUVEC was not showed a cytotoxic response to carmustine loaded oleosome-EGFR treatment while 50 % of cell death with direct treatment of 50 µM carmustine.