Synthesis of Hollow Biomineralized CaCO₃-Polydopamine Nanoparticles for Multimodal Imaging-Guided Cancer Photodynamic Therapy with Reduced Skin Photosensitivity

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Figure S1. Nitrogen adsorption/desorption isotherms and the corresponding pore-size distribution curve (inset) of the CaCO₃-PDA sample.



Figure S2. Digital photographs of $CaCO_3$ -PDA nanoparticles grown in the presence or absence of oxygen, as well as $CaCO_3$ and PDA nanoparticles grown in the presence of oxygen.



Figure S3. The time dependent particle size increases based on the TEM imaging of samples collected at different reaction time.



Figure S4. Thermogravimetric analysis curves of CaCO₃-PDA and CaCO₃-PDA-PEG hollow nanoparticles.



Figure S5. Digital photographs of CaCO₃-PDA and CaCO₃-PDA-PEG hollow nanoparticles in various solutions. Those nanoparticles showed excellent colloidal stability after PEGylation.



Figure S6. Drug loading capacities of CaCO₃-PDA. (a1, a2, a3) molecular structures of DOX (a1), SN38 (a2) and Mitoxantrone (a3). (b1, b2, b3)UV-Vis-NIR spectra of DOX (b1), SN38 (b2) and Mitoxantrone (b3) loaded Ca-CO₃-PDA at different feeding ratios of drug to CaCO₃-PDA. Inserted: photos of drug loaded CaCO₃-PDA. (c1, c2, c3) Quantification of DOX (c1), SN38 (c2) and Mitoxantrone (c3) loading capacities of CaCO₃-PDA at different drug: CaCO₃-PDA-PEG feeding ratios. In our experiments, drugs ((DOX, SN38 or mitoxantrone) at different weight ratios with CaCO₃-PDA was added to the CaCO₃-PDA (2 mg/mL) ethanol solutions under stirring overnight. Later, the free drugs were purified by repeatedly centrifuged with fresh ethanol solutions. The UV-Vis-NIR spectra was then used to quantitatively evaluate the drug loading capacity.



Figure S7. Dynamic light scattering (DLS) data of Ce6@CaCO₃-PDA-PEG (a) and Ce6@liposome (b) and their corresponding particle dispersion index (PDI) in the aqueous solution.



Figure S8. Confocal images of calcein AM (green: live cells) and propidium iodide (red: dead cells) co-stained 4T1 cells with different treatment.



Figure S9. EDX spectra of Fe^{3+} , Zn^{2+} , Mn^{2+} and Co^{2+} doped CaCO₃-PDA hollow nanoparticles.

lons Agents	CaCO ₃ -PDA	CaCO ₃
Zn	0.34	0.3
Fe	4.08	0.82
Со	2.93	1.42
Mn	1.37	0.43

Table S1. Metal ions chelation capacities (Zn, Fe, Co or Mn) of $CaCO_3$ -PDA hollow nanoparticles and bare Ca-CO₃ nanoparticles based on the quantitative energy dispersive spectrometer (EDS) analysis.



Figure S10. (a) T1 weighted MR imaging of CaCO₃-PDA(Mn)-PEG at various Mn^{2+} concentrations. (b) The T1 relaxation rates of CaCO₃-PDA(Mn)-PEG. The longitudinal relaxivity (r1) of CaCO₃-PDA(Mn)-PEG was determined to be 9.5 mM⁻¹ s⁻¹.



Figure S11. UV-Vis-NIR spectra of CaCO₃-PDA-PEG nanoparticles.



Figure S12. (a) PA imaging of CaCO₃-PDA-PEG at different concentrations under the Visualsonic Vevo® 2100 LAZER system (700 nm, 21 MHz). (b) Linear correlation of PA signal intensities of CaCO₃-PDA-PEG at 700 nm against its corresponding PDA concentrations.



Figure S13. H&E staining of major organs collected from mice of different groups post treatments. No noticeable signs of organ damage appeared in all major organs of mice. Group (i) & (ii) were used to represent the group of mice treated with PBS injection only and Ce6@CaCO₃-PDA-PEG + Light exposure, respectively.



Figure S14. (a) Tumor growth curves of mice with i.v. injection of Ce6@CaCO₃-PEG or Ce6@CaCO₃-PDA-PEG and irradiated with 660 nm LED light at a power density of 5 mW cm⁻² for 1 h, at 24 h post injection (5 mice for each group). (b) The body weight variation curves of the two groups of mice post different treatments.