

Supporting Information

Mesoporous Silica Thin Films for Improved Electrochemical Detection of Paraquat

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Electrochemical methods for paraquat detection reported in the literature

Table S1: Comparison of different electrochemical methods for detection of paraquat

Electrode material	Modification	Method	Linear range (mol L ⁻¹)	LOD (mol L ⁻¹)	Preconcentration	Reference
Mercury	--	SWV	5×10^{-8} - 1×10^{-5}	1.5×10^{-8}	No	¹
Boron doped diamond	--	SWV	8×10^{-7} - 1.6×10^{-4}	7×10^{-8}	No	²
Boron doped diamond	--	SWV	5×10^{-9} - 1×10^{-6}	1.5×10^{-9}	No	³
Au-μE	--	SWV	No data	1.76×10^{-8}	No	⁴
Au-μE	--	SWV	5×10^{-7} - 1×10^{-5}	8.2×10^{-8}	No	⁵
Cu	Bismuth film	DPV	0.1×10^{-6} - 4.2×10^{-6}	1.2×10^{-8}	No	⁶
Cu	Bismuth Film	DPV	6.6×10^{-7} - 4.8×10^{-5}	9.3×10^{-8}	No data	⁷
Au	Au NPs-DNA	SWV	5×10^{-6} - 1×10^{-3}	1.9×10^{-6}	No data	⁸
CPE	Ag NP	SWV	1×10^{-7} - 1×10^{-3}	2.01×10^{-8}	10 minutes	⁹
CPE	Sb NPs-Biochar	SWV	0.2 - 2.9×10^{-6}	3.4×10^{-8}	2 minutes	¹⁰
Au	Au NPs-DNA	DPV	7×10^{-9} - 1.5×10^{-6}	2×10^{-10}	No data	¹¹
Graphene	(Ppy-g-NGE)	DPV	5×10^{-8} - 2×10^{-6}	4.1×10^{-8}	5 minutes	¹²
GCE	Carbon nanotubes - DHP	SWV	5×10^{-8} - 1.5×10^{-6}	1×10^{-8}	No data	¹³
GCE	PVP-GNs/Cu ₂ O	DPV	1×10^{-6} - 2×10^{-4}	2.7×10^{-7}	No data	¹⁴
Pyrolytic graphite	Metallo-phthalocyanine	SWV	5×10^{-7} - 2.9×10^{-5}	1.03×10^{-7}	10 minutes	¹⁵
CPE	Biochar	DPAAdSV	3×10^{-8} - 1×10^{-6}	7.5×10^{-9}	5 minutes	¹⁶
GCE	Nafion	CDPV	3.9×10^{-9} - 3.9×10^{-7}	1.9×10^{-9}	3 minutes	¹⁷
GCE	Nafion - Clay	SWV	3.9×10^{-8} - 2.7×10^{-7}	1.9×10^{-9}	4 minutes	¹⁸

CPE	Amberlite XAD-2 resin	CSV	4.2×10^{-6}	3.9×10^{-7}	3 minutes	¹⁹
CPE	Natural phosphate	SWV	2.3×10^{-8} - 3×10^{-6}	7.8×10^{-10}	10 minutes	²⁰
CPE	Hydroxyapatite	SWV	8×10^{-7} - 2×10^{-5}	1.5×10^{-8}	30 minutes	²¹
GCE	Mesoporous silica	SWV	1×10^{-8} - 5×10^{-8}	4×10^{-9}	3 minutes	This study

GCE: Glassy carbon electrode; CPE: Carbon paste electrode; AuNPs: Gold nanoparticles; PVP-GNs/Cu₂O-Polyvinyl pyrrolidone-graphene nanosheets/Cuprous oxide; Au-μE-Gold microelectrode; DHP: dihexadecylhydrogenophosphate; DPAdSV: Differential pulse adsorptive stripping voltammetry; DPV: Differential pulse voltammetry; SWV: Square wave voltammetry; CDPV: Cathodic differential pulse voltammetry; CSV: Cathodic stripping voltammetry.

Electrochemical impedance spectroscopy

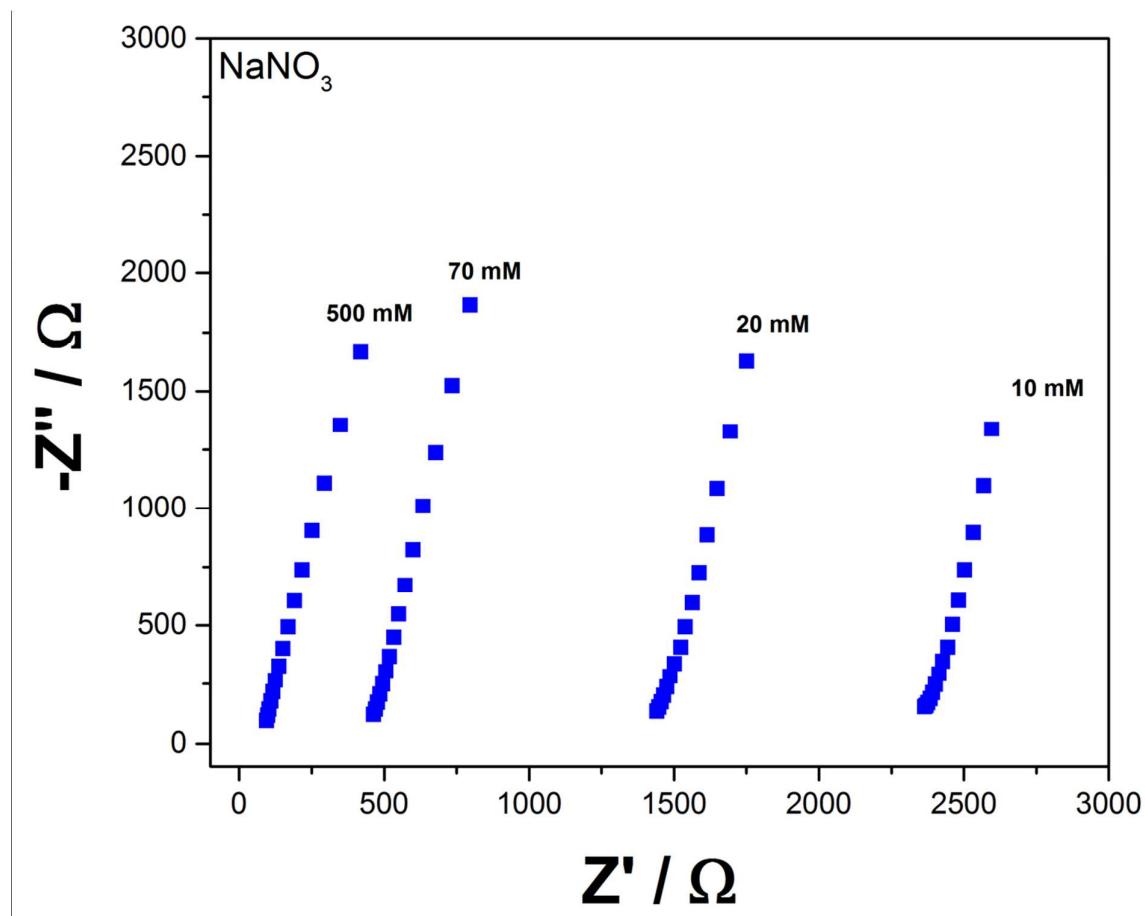


Figure SI1: Nyquist plot for the impedance measured at bare electrodes in solutions of different NaNO_3 concentrations ($\text{pH} = 6$). Frequencies were between 60 and 2000 Hz.

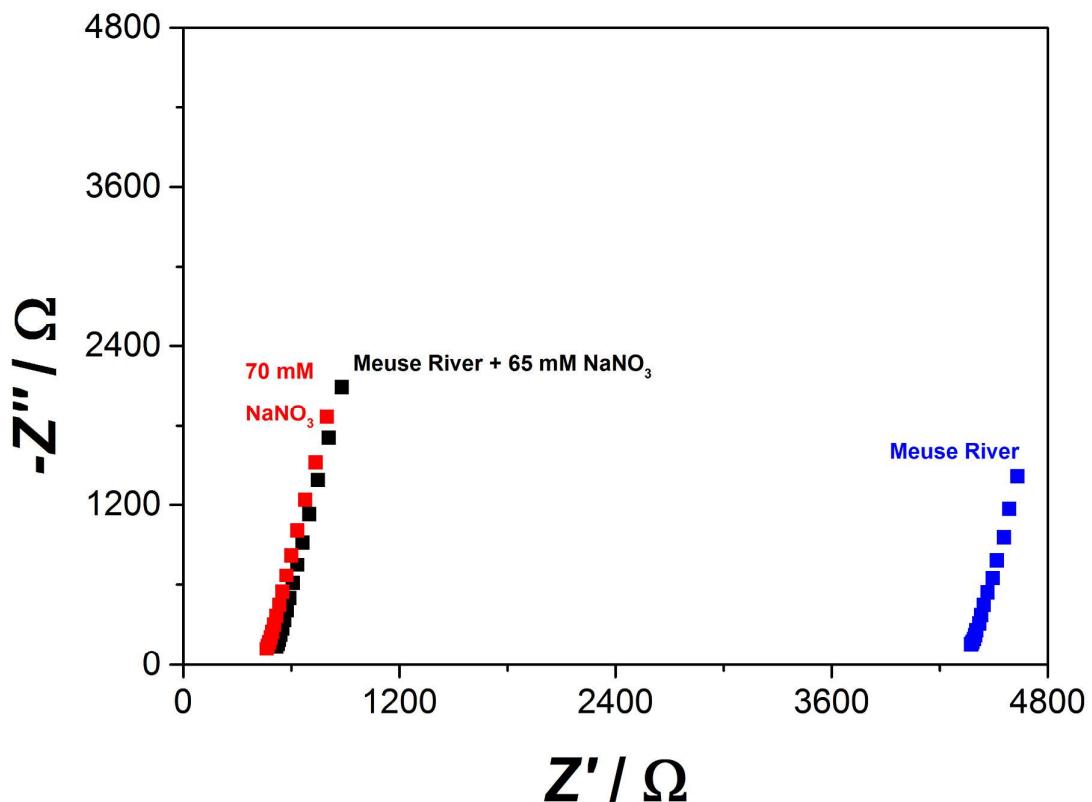


Figure SI2: Nyquist plot for the impedance measured in a laboratory solution of 70 mM NaNO₃ (red squares), in Meuse River sample before (blue squares) and after the additions of 65 mM NaNO₃ at a bare electrode.

Table SI2: Solution resistances obtained by electrochemical impedance spectroscopy

<i>I</i> / mM	<i>R_s</i> / Ω ^a
10	2344
20	1392
50	573
70	422
100	295
500	64
Real sample	4335
Real sample + 65mM NaNO ₃	460

R_s : Solution resistance extracted from Nyquist plot obtained at the bare electrode. Same conditions as Figure SI1.

Debye Length as a function of ionic strength

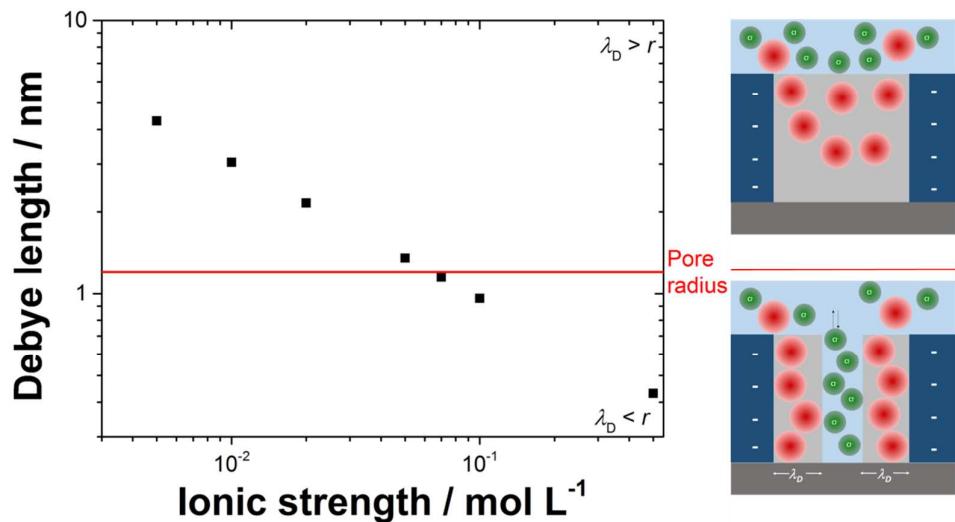


Figure SI3: Debye length as a function of the ionic strengths used in the SWV experiments. The red line symbolizes the value of the mesochannel radius. On the right, two schemes represent anion exclusion of the mesochannels (top) and the ‘free’ anion zone.

Electrochemical paraquat detection at three different kind of electrodes

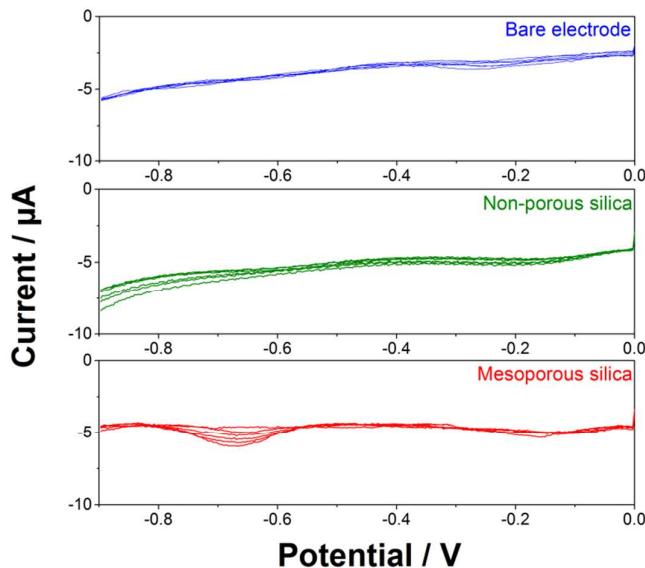


Figure SI4: Square wave voltammograms of 10 – 50 nM paraquat at a bare electrode (top graph, blue curves); a non-porous silica modified electrode (middle graph, green curves) and a mesoporous silica modified electrode (bottom graph, red curves) obtained in the optimal experimental conditions (i.e. 70 mM NaNO₃ and pH 6).

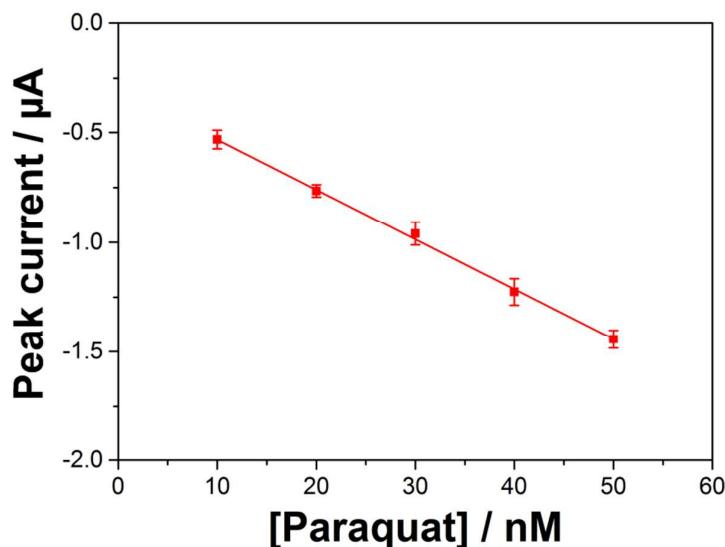


Figure S15: Calibration curve for 10 – 50 nM paraquat at a mesoporous silica modified electrode obtained in the optimal experimental conditions (i.e. 70 mM NaNO₃ and pH 6).

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