SUPPORTING INFORMATION

Decoration of Polyfluorene-Wrapped Carbon Nanotubes via Strain-Promoted Azide-Alkyne

Cycloaddition

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Experimental

General

Semi-purified plasma torch SWNTs (RN120, batch #RNB735-120-X464) were purchased from Raymor Industries Inc. and used without further purification. Raw HiPCO SWNTs were purchased from NanoIntegris (batch #HR27-104, 10 wt % in anhydrous EtOH) and also used without further purification. Reagents were purchased from commercial suppliers and used as received. Monomethyl ether mPEG₅₀₀₀ was purchased from Fluka. Flash chromatography was performed using an IntelliFlash 280 system from Analogix. Unless otherwise noted, compounds were monitored using a variable wavelength detector at 254 nm. Solvent amounts used for gradient or isocratic elution were reported in column volumes (CV). Columns were prepared in Biotage[®] SNAP KP-Sil cartridges using $40 - 63 \mu m$ silica or 25 – 40 µm silica purchased from Silicycle. NMR was performed on a Bruker Avance 600 MHz or 700 MHz instrument and shift-referenced to the residual solvent resonance. Polymer molecular weights and dispersities were analyzed (relative to polystyrene standards) via GPC using a Waters 2695 Separations Module equipped with a Waters 2414 refractive index detector and a Jordi Fluorinated DVB mixed bed column in series with a Jordi Fluorinated DVB 10⁵ Å pore size column. THF with 2% acetonitrile was used as the eluent at a flow rate of 2.0 mL/min. Sonication was performed in a Branson Ultrasonic B2800 bath sonicator. Centrifugation of the polymer-SWNT samples was performed using a Sorvall Legend X1R centrifuge (semi-purified plasma torch SWNT dispersions) or a Beckman Coulter Allegra X-22 centrifuge (raw HiPCO SWNT dispersions). Infrared spectra were recorded using a Thermo Scientific Nicolet 6700 FT-IR spectrometer equipped with a Smart iTX attenuated total reflectance (ATR) sample analyzer. UV-Vis-NIR spectra were recorded on a Cary 5000 spectrometer in dual beam mode, using matching 10 mm quartz cuvettes. Fluorescence spectra were measured on a Jobin-Yvon SPEX Fluorolog 3.22 equipped with a 450 W Xe arc lamp, digital photon counting photomultiplier, and an InGaAs detector, also using a 10 mm quartz cuvette. Slit widths for both excitation and emission were set to 10 nm band-pass, and correction factor files were applied to account for instrument variations. Photoluminescence maps were obtained at 25 °C, with 5 nm intervals for both the excitation and emission. Raman spectra were collected using a Renishaw InVia Laser Raman spectrometer, with three different lasers: a 25 mW argon ion laser (514 nm, 1800 L/mm grating); a 500 mW HeNe Renishaw laser (633 nm, 1800 L/mm grating); and a 300 mW Renishaw laser (785 nm, 1200 L/mm grating). Laser intensity was set to 0.5% for the 785 nm excitation wavelength and 5% for the 514 nm and 633 nm excitations for the polymer-SWNT samples. For the SWNT sample dispersed in CHCl₃, laser intensity was set to 1% for 514 nm and 633 nm, and 10% for 785 nm.



Scheme S1. Synthesis of monomers 3 and 4.

Synthetic Procedures

2,7-dibromofluorene (1) (adapted from reference 1)

A round bottom flask equipped with a stir bar was charged with fluorene (33.2 g, 200 mmol), NBS (89.0 g, 500 mmol) and acetic acid (400 mL). While the mixture was stirring, conc. HBr (10 mL) was slowly added and then the reaction mixture was stirred at RT for 1.5 h. Water (200 mL) was added and the resulting suspension was filtered and washed with water to obtain an orange-white solid. The solid was recrystallized from a 1.5:1 v/v mixture of EtOH:acetone (~1.8 L total volume), and the mother

liquor was recrystallized again from the same solvent mixture (~1.5 L total volume). The crops were combined to afford **1** (41.2 g, 64%) as a white solid. ¹H-NMR (600 MHz; CDCl₃): δ 7.67 (d, *J* = 1.1 Hz, 2H), 7.61 (d, *J* = 8.1 Hz, 2H), 7.51 (dd, *J* = 8.1, 1.8 Hz, 2H), 3.88 (s, 2H).

2,7-dibromo-9,9-didodecylfluorene (2) (adapted from reference 1)

A round bottom flask equipped with a stir bar was charged with 1 (5 g, 15.4 mmol), 1-bromododecane (7.88 g, 31.6 mmol), toluene (31 mL), and sat. KOH (31 mL). ⁿBu₄NBr (1.0 g, 3.1 mmol) was then added and the reaction mixture was heated to 60 °C and stirred vigorously for 1 h under a nitrogen atmosphere. The biphasic mixture was allowed to separate, and the organic layer was isolated. The aqueous phase was extracted twice with diethyl ether (2 x 120 mL) and the organic extracts were combined and concentrated *in vacuo* to obtain a viscous green oil. The crude mixture was purified by flash chromatography (100 g column, 100% hexanes) to afford **2** as a white solid (9.34 g, 92%). ¹H-NMR (600 MHz; CDCl₃): δ 7.51 (d, *J* = 8.0 Hz, 1H), 7.45-7.43 (m, 2H), 1.92-1.89 (m, 2H), 1.29-1.04 (m, 18H), 0.87 (t, 3H), 0.59-0.56 (m, 2H).

2,7-dibromo-9,9-bis(6-bromohexyl)fluorene (3) (adapted from reference 1)

A round bottom flask equipped with a stir bar was charged with **1** (5 g, 15.4 mmol), 1,6dibromohexane (37.7 g, 154 mmol), toluene (31 mL), and sat. KOH (31 mL). ⁿBu₄NBr (1.0 g, 3.1 mmol) was then added and the reaction mixture was heated to 60 °C and stirred vigorously for 1 h under a nitrogen atmosphere. The biphasic mixture was allowed to separate, and the organic layer was isolated. The aqueous phase was extracted twice with diethyl ether (2 x 120 mL) and the organic extracts were combined and concentrated *in vacuo* to obtain a viscous green oil. Excess 1,6-dibromohexane was removed using vacuum distillation (1 mbar, 115 °C) to obtain a viscous yellow oil. The crude mixture was purified by flash chromatography (100 g column, 0 to 20% CH₂Cl₂ in hexanes over 10 CV) to obtain a white solid containing two spots by TLC. The crude product was recrystallized from MeOH (~250 mL) to afford **3** as a white solid (4.4 g, 44%). ¹H-NMR (600 MHz; CDCl₃): δ 7.537.52 (m, 1H), 7.47-7.43 (m, 2H), 3.31-3.28 (t, 2H), 1.94-1.91 (m, 2H), 1.68-1.66 (m, 2H), 1.22-1.19 (m, 2H), 1.10-1.07 (m, 2H), 0.60-0.57 (m, 2H).

2,2'-(9,9-didodecylfluorene-2,7-diyl)bis(4,4,5,5-tetramethyl-1,3,2-dioxaborolane) (4) (adapted from reference 1)

A round bottom flask equipped with a stir bar was charged with **2** (7.0 g, 10.6 mmol), B₂Pin₂ (5.92 g, 23.3 mmol), KOAc (3.12 g, 31.8 mmol), and dioxane (100 mL). Pd(dppf)₂Cl₂ (260 mg, 318 µmol) was added and the reaction mixture was stirred at 80 °C for 12 h. The reaction mixture was partitioned with water and extracted thrice with Et₂O. The organic extracts were combined and dry loaded onto silica (10 g). The crude mixture was purified by flash chromatography (100 g column, 0 to 80% CH₂Cl₂ in hexanes over 10 CV) to obtain a yellow solid. The solid was recrystallized from a 1.25:1 v/v mixture of MeOH:acetone (~450 mL total volume) to afford **4** as a white solid (4.88 g, 63%). ¹H-NMR (600 MHz; CDCl₃): δ 7.80 (d, *J* = 7.6 Hz, 1H), 7.74-7.71 (m, 2H), 2.00-1.97 (m, 2H), 1.39 (s, 12H), 1.26-0.99 (m, 18H), 0.86 (t, *J* = 7.1 Hz, 3H), 0.54 (m, 2H).



Poly(didodecylfluorene-alt-bis(bromohexyl)fluorene) (PF-Br)

A Schlenk tube equipped with a stir bar was charged with **3** (646 mg, 0.99 mmol), **4** (750 mg, 0.99 mmol), toluene (7.1 mL), and 3M K₃PO_{4(aq)} (7.1 mL). The biphasic mixture was degassed by three freeze-pump-thaw cycles, then, while frozen under liquid nitrogen, $[(o-tol)_3P]_2Pd$ (36 mg, 50 µmol) was added under a positive pressure of nitrogen. The Schlenk tube was evacuated and backfilled with nitrogen four times, and the reaction mixture was vigorously stirred at 80 °C for 12 h. The phases were allowed to separate, and the organic layer was isolated and filtered through a single plug of celite and neutral alumina. The plug was thoroughly washed with THF and the flow-through was concentrated *in vacuo*. The crude polymer was precipitated into MeOH (~400 mL) and then filtered to afford **P1** as a

yellow solid (760 mg, 77%). ¹H-NMR (600 MHz; CDCl₃): δ 7.86-7.83 (m, 4H), 7.73-7.66 (m, 8H), 3.31-3.28 (m, 4H), 2.20-2.10 (m, 4H), 1.72-1.69 (m, 4H), 1.29-1.11 (m, 50H), 0.86 (t, *J* = 7.1 Hz, 12H).



Poly(didodecylfluorene-alt-bis(azidohexyl)fluorene) (PF-N₃)

A round bottom flask equipped with a stir bar and reflux condenser was charged with **PF-Br** (600 mg, 0.61 mmol), NaN₃ (394 mg, 6.1 mmol), ⁿBu₄NBr (390 mg, 1.2 mmol), and THF (90 mL) and the reaction mixture was heated to reflux for 12 h. The reaction mixture was filtered through an alumina plug and washed thoroughly with THF, then the solution was concentrated *in vacuo* and precipitated into MeOH (~400 mL) to afford **PF-N₃** as a yellow solid (407 mg, 73%). ¹H-NMR (600 MHz; CDCl₃): δ 7.86-7.83 (m, 4H), 7.73-7.64 (m, 8H), 3.15-3.12 (m, 4H), 2.16-2.11 (m, 4H), 1.45-1.42 (m, 4H), 1.27-1.10 (m, 50H), 0.86 (t, *J* = 7.0 Hz, 12H).



mPEG₅₀₀₀-**DIBAC.** A round bottom flask equipped with a stir bar was charged with mPEG₅₀₀₀-OH (250 mg, 50 μ mol), DIBAC-COOH (46 mg, 150 μ mol) and 4-dimethylaminopyridine (3 mg, 25 μ mol) and CH₂Cl₂ (1 mL). To the solution, *N*-(3-dimethylaminopropyl)-*N*'-ethylcarbodiimide hydrochloride (EDC·HCl) (33 mg, 175 μ mol) was added, and the reaction mixture was stirred overnight at RT. The reaction mixture diluted with 5 mL of CH₂Cl₂ and then precipitated into 1:1 Et₂O:hexanes (~100 mL).

The precipitate was collected on a Hirsch funnel, then washed with ice cold ethanol (3 x 20 mL), Et₂O (3 x 20 mL), and dried *in vacuo* to afford **mPEG**₅₀₀₀-**DIBAC** as a fine white powder (246 mg, 93%). ¹H NMR (600 MHz; DMSO-d6): δ 7.66 (dd, J = 7.6, 1.2, 1H), 7.62 (d, J = 7.3, 1H), 7.52-7.45 (m, 3H), 7.38 (td, J = 7.4, 1.5, 1H), 7.34 (td, J = 7.4, 1.3, 1H), 7.30 (dd, J = 7.5, 1.2, 1H), 5.03 (d, J = 14.2, 1H), 4.03-3.99 (m, 1H), 3.95-3.91 (m, 1H), 3.51 (s, 1H), 3.24 (s, 3H), 2.67-2.62 (m, 2H), 2.37-2.35 (m, 1H), 2.29 (dt, J = 17.1, 6.4, 1H), 1.81 (dt, J = 16.7, 6.3, 1H).

General SPAAC procedure. A glass vial was charged with 4 mL of polymer-SWNT dispersion and 2.05 equivalents of DIBAC derivative was added directly. Equivalents were calculated by determining the polymer concentration in solution (based on the SWNT dispersion protocol), calculating the mass of polymer present, and then using the molecular weight of the repeat unit to determine the number of moles of azide moiety (two per repeat unit). The glass vial was agitated until the DIBAC derivative was dissolved, and then the reaction progress was monitored by IR spectroscopy.

SPAAC with DIBAC-COOH. 4 mL of PF-N₃-SWNT dispersion, 1.4 mg of DIBAC-COOH.

SPAAC with mPEG₅₀₀₀**-DIBAC (plasma torch SWNTs).** 4 mL of **PF-N**₃-SWNT plasma torch dispersion, 23.7 mg of mPEG₅₀₀₀-DIBAC.

SPAAC with mPEG₅₀₀₀-DIBAC (HiPCO SWNTs). 4 mL of **PF-N₃-SWNT** HiPCO dispersion, 35.5 mg of mPEG₅₀₀₀-DIBAC.

HiPCO Dispersion with PF-N₃-mPEG₅₀₀₀

PF-N₃ (3 mg) and mPEG₅₀₀₀-DIBAC (35.5 mg) were dissolved in toluene (4 mL) and allowed to stand at RT overnight. HiPCO SWNTs (3 mg dry weight) were then added and the mixture was sonicated for 2 h in a bath sonicator chilled with ice. The suspension was centrifuged at 8,346 g for 30 min, and the supernatant was carefully removed and filtered through a cotton plug. The as-produced sample was analyzed using UV-Vis-NIR spectroscopy, and was diluted 16-fold (200 μ L of the as-produced dispersion was added to 3.0 mL of toluene) to obtain the spectrum shown in Figure S10.



Figure S1. ¹H NMR overlay of (a) PF-Br (blue) and (b) PF-N₃ (red).



Figure S2. FT-IR overlay of (a) PF-Br (orange) and (b) PF-N₃ (blue).





Figure S4. Raman spectra collected at 785 nm of PF-N₃-SWNT dispersions (a) pre- (green) and (b) post-SPAAC (purple) with DIBAC-COOH. Spectra were normalized to the G-band at ~1590 cm⁻¹ and offset for clarity.



Figure S5. FT-IR overlay of the SPAAC reaction between **PF-N₃-SWNT** and mPEG₅₀₀₀-DIBAC (a) pre- (dark red) and (b) post-SPAAC (light blue). The post-SPAAC spectrum was obtained after 30 min.



Figure S6. UV-Vis-NIR spectra of **PF-N₃-SWNT** dispersions in 50/50 THF:D₂O with (i) nothing, (ii) two equivalents of mPEG₅₀₀₀-OH (containing no DIBAC), or (iii) two equivalents of mPEG₅₀₀₀-DIBAC. Photographs of the filtered dispersions are colour-coded accordingly with coloured dots above.



Figure S7. UV-Vis-NIR spectra of **PF-N₃-HiPCO** dispersions pre-SPAAC (toluene, dark yellow trace) and post-SPAAC with mPEG₅₀₀₀-DIBAC (D₂O, blue trace). Spectra are offset for clarity. The photograph shows the pre-SPAAC polymer-SWNT dispersion in toluene.



Figure S8. UV-Vis-NIR spectra of PF-N₃-HiPCO dispersions diluted for PL mapping.



Figure S9. PL maps of **PF-N₃-**HiPCO in (a) toluene and (b) toluene mixed with 2.05 eq of mPEG₅₀₀₀-OH.

Table S1. Emission Wavelengths of sc-SWNT Chiralities Dispersed by $PF-N_3$ in Toluene and Toluene with mPEG₅₀₀₀-OH

| Chirality | λ_{em} (nm) (-PEG _{5k}) | λ_{em} (nm) (+PEG _{5k}) | $\Delta[\lambda_{em}]$ (nm) |
|----------------|---|---|-----------------------------|
| (6,5) | 990 | 990 | 0 |
| (7,5) | 1047 | 1046 | -1 |
| (7,6) | 1133 | 1133 | 0 |
| (8,4) | 1131 | 1130 | -1 |
| (8,6) | 1196 | 1196 | 0 |
| (8,7) | 1280 | 1279 | -1 |
| (9,4) | 11245 | 1126 | +1 |
| (9 <i>,</i> 5) | 1265 | 1264 | -1 |



Figure S10. UV-Vis-NIR spectrum of **PF-N₃-mPEG₅₀₀₀** used to disperse HiPCO SWNTs in toluene. Note that the spectrum was obtained by diluting the as-produced dispersion by a factor of 16 in toluene. The photograph shows the as-produced polymer-SWNT dispersion (undiluted).

References

(1) Xia, C.; Advincula, R. C. Decreased Aggregation Phenomena in Polyfluorenes by Introducing

Carbazole Copolymer Units. *Macromolecules* **2001**, *34*, 5854–5859.