Supporting Information for:

HU-331 and Oxidized Cannabidiol Act as Inhibitors of Human Topoisomerase II α and β

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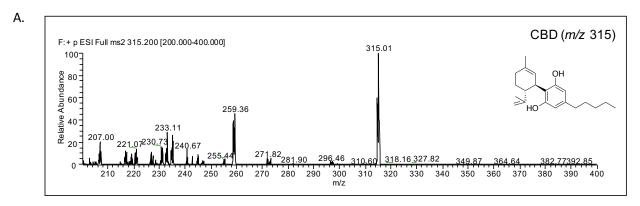
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LC-MS/MS Analysis of Cannabidiol, Oxidized Cannabidiol, and HU-331. Cannabidiol and HU-331 were subjected to LC-MS/MS analysis to develop a method for detection and relative quantitation. LC-MS/MS analysis was performed using a Thermo TSQ Quantum Triple Quadrupole mass spectrometer coupled to a Thermo Accela HPLC/UHPLC system. An aliquot (20 μ l) was injected into the equilibrated HPLC system using a Thermo PAL autoinjector. The following solvents were used – Mobile Phase A: Optima LC-MS-grade water (Fisher Scientific) with 0.1% (v/v) formic acid, and Mobile Phase B: Optima LC-MS-grade acetonitrile (Fisher Scientific) with 0.1% (v/v) formic acid. Compound separation was achieved using a Kinetex EVO C18 column (2.6 μ m, 50 mm x 2.1 mm, 100 Å) (Phenomenex Inc., Torrance, CA) with a flow rate of 0.3 ml/min and column oven temperature 30°C. The LC gradient system was as follows: 30 to 95% B (0 – 2 minutes), 95% B (2 – 4 minutes), 95 to 30% B (4 – 4.5 minutes), and 30% B (4.5 – 5 minutes). The mass spectrometer was operated in positive ion mode using electrospray ionization (ESI+). MS spectral data were analyzed by Thermo Xcalibur software 2.0.

Standard solutions of CBD and HU-331 (20 mM in DMSO) were diluted to 20 μ M in acetonitrile for LC-MS/MS analysis. Collision induced dissociation of the precursor ion m/z 315 for CBD and m/z 329 for HU-331 was performed in product ion mode at collision energy 21 V to identify the characteristic fragmentation pattern for each compound (**Figure S1**). Next, a selected reaction monitoring (SRM) method was developed for compound detection using the precursor-to-product ion transitions m/z 315 \rightarrow 259 for CBD and m/z 329 \rightarrow 287 for HU-331. Relative levels of oxidized CBD were measured based on the SRM peak area (m/z 329 \rightarrow 287), similar to HU-331 (**Figures S2 and S4**). Peak areas were determined using the auto-integration function in Thermo Xcalibur software.



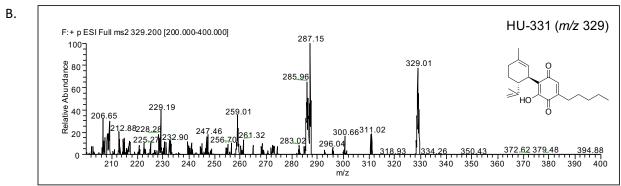


Figure S1: LC-MS/MS Analysis of Cannabidiol (CBD) and HU-331. Standard solutions of 20 μ M CBD (A) and HU-331 (B) were subjected to liquid chromatography tandem mass spectrometry (LC-MS/MS) analysis via electrospray ionization in positive ion mode. CBD has a precursor ion of m/z 315, and HU-331 has a precursor ion of m/z 329. Shown are representative product ion spectra generated from collision induced dissociation of CBD (m/z 315) (A) and HU-331 (m/z 329) (B) at collision energy 21 V.

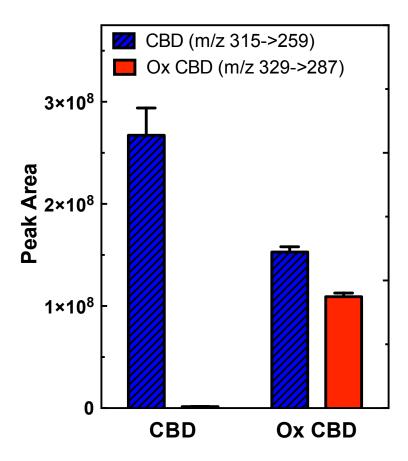


Figure S2: Analysis of Oxidation of Cannabidiol in Solution. Pure cannabidiol has a molecular ion at m/z 315 when analyzed by LC-MS/MS in positive ion mode, while HU-331 is observed at m/z 329. Samples of cannabidiol (CBD) and oxidized cannabidiol (Ox CBD) were subjected to LC-MS/MS analysis at matching concentrations. The precursor-to-product ion transitions for CBD (m/z 315>259) and Ox CBD (m/z 329>287) were used to measure relative levels of each compound by selected reaction monitoring. Results above display the peak areas for CBD (m/z 315>259) and Ox CBD (m/z 329>287) in two samples from an initial concentration of CBD at 200 μ M. Cannabidiol appears to oxidize in solution over time. Results represent the mean of two samples and error bars represent the standard error of the mean.

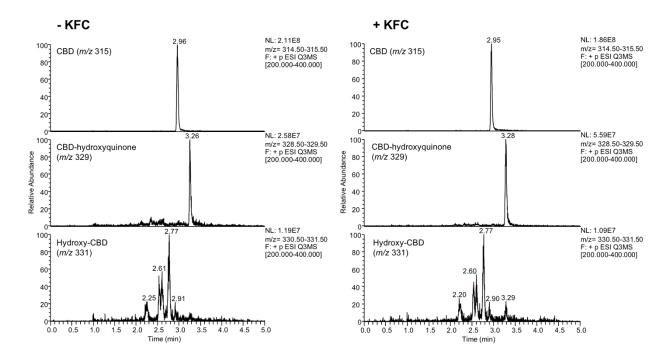


Figure S3: KFC-oxidized CBD. LC-MS/MS analysis of oxidized CBD products with and without KFC treatment. Shown are LC-MS extracted ion chromatograms for CBD (*m/z* 315), CBD-hydroxyquinone (*m/z* 329), and hydroxy-CBD products (*m/z* 331) +/- KFC. It should be noted that the hydroxy-CBD products in the –KFC reaction (left lower chromatogram) are the result autooxidation by air since we have also observed no hydroxy-CBD in spectra of pure CBD (see figure 6).

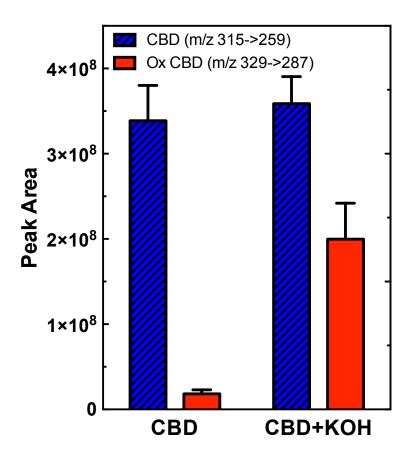


Figure S4: Analysis of KOH-Oxidized Cannabidiol. Samples of cannabidiol (CBD) and KOH-oxidized cannabidiol (Ox CBD) were subjected to LC-MS/MS analysis at matching concentrations. The precursor-to-product ion transitions for CBD (m/z 315>259) and Ox CBD (m/z 329>287) were used to measure relative levels of each compound by selected reaction monitoring. Results above display the peak areas for CBD (m/z 315>259) and Ox CBD (m/z 329>287) in two samples from an initial concentration of CBD at 200 μ M. Results are plotted as the mean of four replicates and error bars represent the standard deviation of those samples.

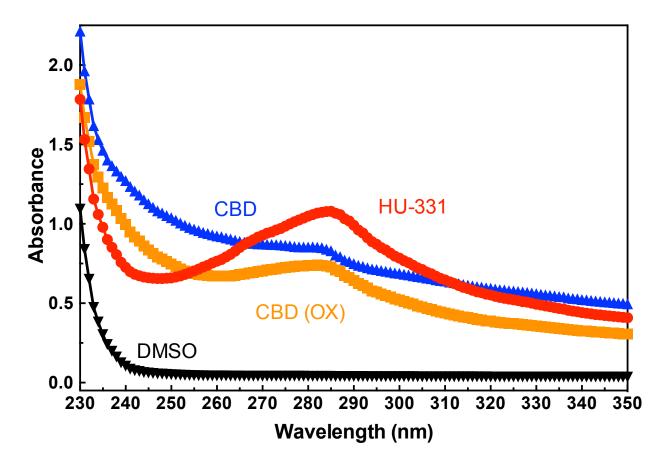


Figure S5: UV Absorbance Spectra for CBD, Oxidized CBD, and HU-331. The absorbance spectra at 1 nm resolution for 20 mM CBD, oxidized CBD (ox CBD), and HU-331 are shown.

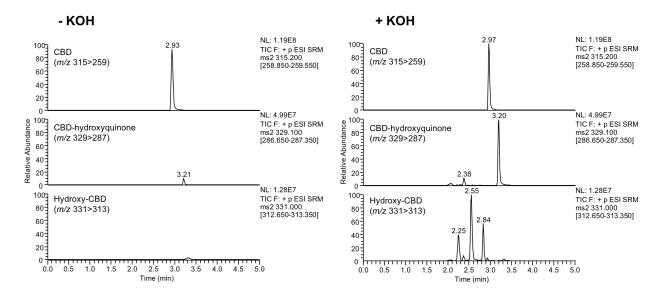


Figure S6: KOH-oxidized CBD. LC-MS/MS analysis of CBD oxidized products with and without KOH treatment. Shown are LC-MS/MS chromatograms from selected reaction monitoring (SRM) of CBD (m/z 315>259), CBD-hydroxyquinone (m/z 329>287), and hydroxy-CBD products (m/z 331>313) +/- KOH. The graphs have been scaled equally for comparison of treatment conditions.