
The toxic effects of bisphenol S showing immunomodulation in fish macrophages

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SI Materials and Methods

Phagocytic activities of primary macrophages. The phagocytic activities of primary macrophages against the pathogenic gram-positive *Staphylococcus aureus* (CGMCC 1.363), gram-negative *Vibrio parahaemolyticus* (CGMCC 1.1615), and green fluorescent protein (GFP)-expressing *Escherichia coli* (CGMCC 1.2389) were measured following modified protocols described in previous studies^{1, 2}. The GFP-expressing *E. coli* were generated prior to the experiment. A GFP-expressing plasmid was constructed using a pET-28a expression vector (Novagen). After confirming by DNA sequencing, the pET-28a/GFP plasmid was used to transform *E. coli* BL21(DE3)pLysS cells. A single colony of *E. coli* BL21(DE3)pLysS harboring the pET28/GFP plasmid was induced by adding 0.5 mM isopropyl- β -D-thiogalactopyranoside (IPTG) to LB medium at 37°C. The culture was incubated with shaking at 180 rpm and 28°C for 3 h until the OD₆₀₀ reached 0.2–0.3. *S. aureus* and *V. parahaemolyticus* were cultured to the logarithmic phase and counted using the OD value.

Detached primary macrophages were separated on a 34%/51% (v/v) Percoll density gradient (GE, USA) by centrifugation. After exposure to bisphenol S (BPS)-supplemented culture medium for 6 h, the macrophages were washed three times and resuspended in 0.85 mL of antibiotic-free culture medium. Fifty milliliters of freshly thawed ice-cold normal serum and 0.1 mL of bacteria were added to each well ensuring that the proportion of macrophages to bacteria was 1:10. After cultivation on a Labquake shaker at 8 rpm for 90 min at 26°C, the cells were washed four times to remove the bacteria that had not been engulfed. Final suspensions of macrophages were resuspended in 200 μ L of PBS and applied to slides using a cytocentrifuge. The phagocytic activity of macrophages against GFP-expressing *E. coli* was evaluated under a fluorescence microscope (LSM 780 NLO, ZEISS) or directly in a fluorescence microplate (Infinite® M1000 Pro, Tecan). The phagocytic activity of macrophages against *S. aureus* and *V.*

parahaemolyticus was analyzed at $\times 100$ under an oil-immersion microscope (Axio Observer A1, ZEISS) after staining using a Diff-Quik Kit (Nanjing Jian Cheng Bioengineering Institute, China). The phagocytosis of macrophages was evaluated using a phagocytic index, which as determined by using the following formula: phagocytic index = (percentage of macrophages containing at least one bacterium) \times (mean number of bacteria per positive cell).

REFERENCES

1. Qiu, W. H.; Liu, S.; Chen, J. S.; Hu, L.; Wu, M. H.; Yang, M. The primary culture of carp (*Cyprinus carpio*) macrophages and the verification of its phagocytosis activity. *In Vitro Cell Dev-An.* **2016**, *52* (1), 10-19.
2. Campbell, P. A.; Canono, B. P.; Drevets, D. A. Measurement of bacterial ingestion and killing by macrophages. *Curr. Protoc. Immunol.* **2001**, *Chapter 14*, Unit 14 6.

SI Figures

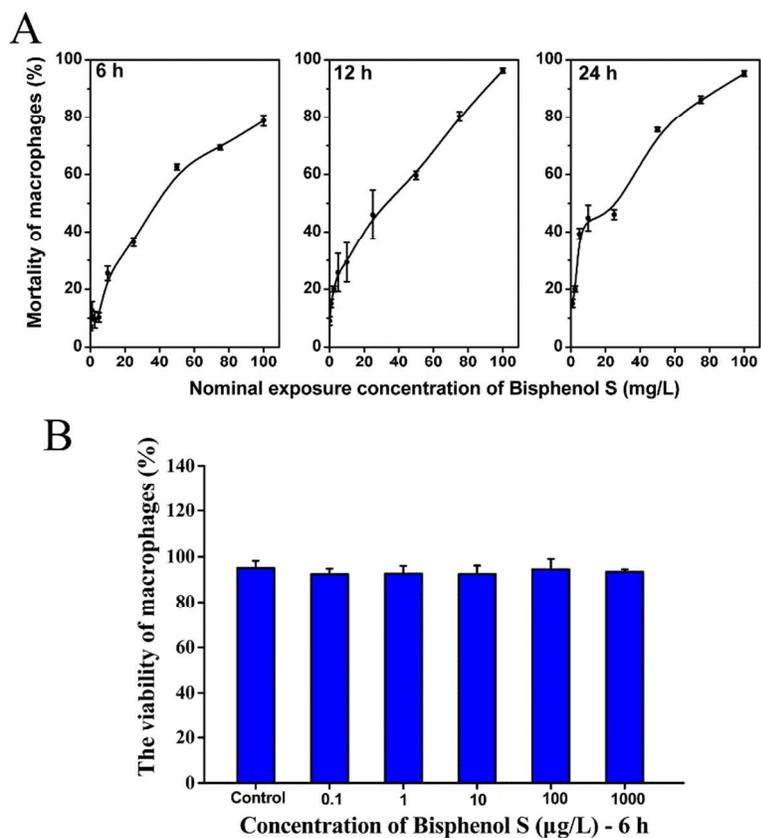


Figure. S1. Acute toxicity (A) and sublethal toxicity (B) of bisphenol S to the primary macrophages of red carp. Mortality was observed at 6, 12, and 24 h after exposure. Values are the means and standard error of the means ($n = 6$) relative to the control.

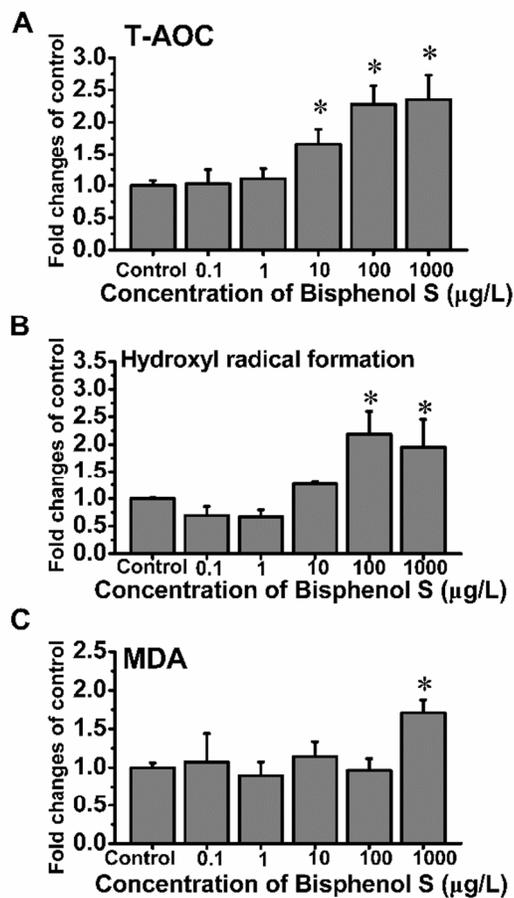


Figure. S2. Induction of total antioxidant capacity (T-AOC, A), hydroxyl radical formation level (B) and malonaldehyde (MDA) content (C) in primary macrophages following a 6-h exposure to bisphenol S (BPS). The level of MDA represents the lipid peroxidation level. Values are the means \pm standard deviation ($n = 3$). An asterisk (*) indicates a significant difference versus the control at $p < 0.05$ (ANOVA, Tukey's test).

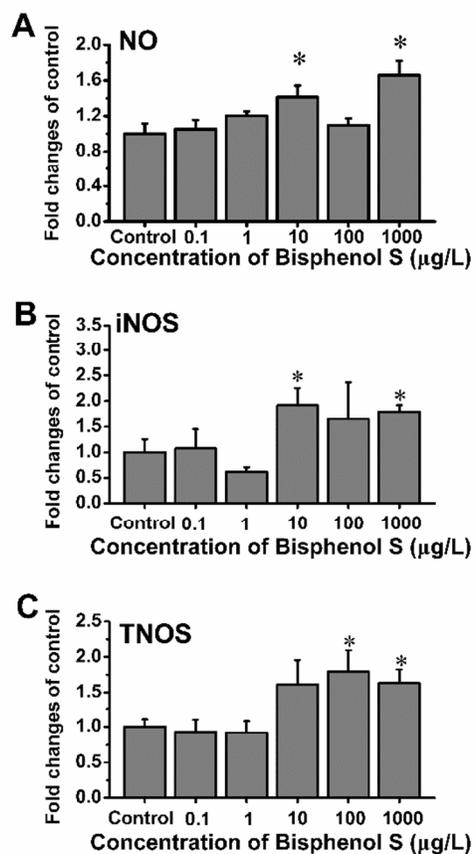


Figure S3. Induction of nitric oxide production (NO, A), and induced nitric oxide synthase (iNOS, B) and total nitric oxide synthase (TNOS, C) activities in primary macrophages following a 6-h exposure to bisphenol S (BPS). Values are the means \pm standard deviation ($n = 3$). An asterisk (*) indicates a significant difference versus the control at $p < 0.05$ (ANOVA, Tukey's test).

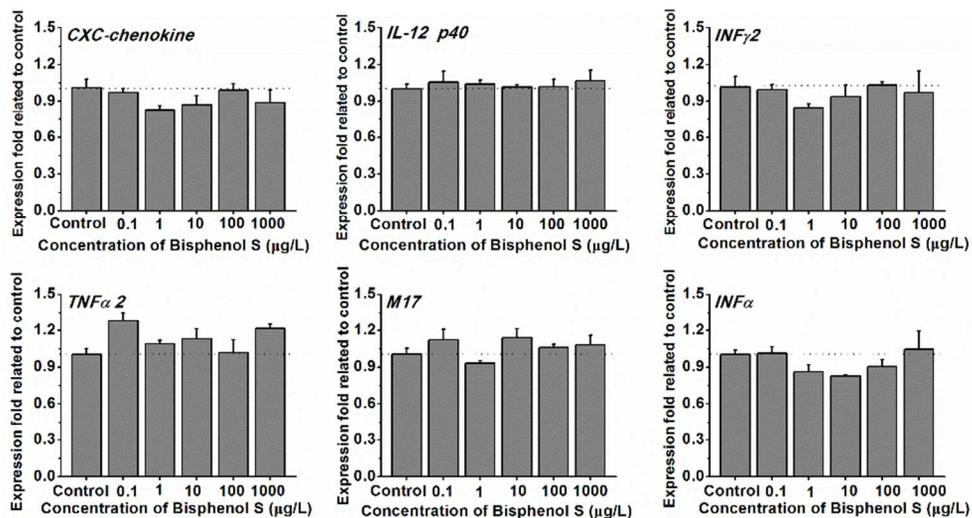


Figure S4. The expression of immune-related genes in primary macrophages following a 6-h exposure to bisphenol S (BPS). Values are the means \pm standard deviation ($n = 3$). An asterisk (*) indicates a significant difference versus the control at $p < 0.05$ (ANOVA, Tukey's test).

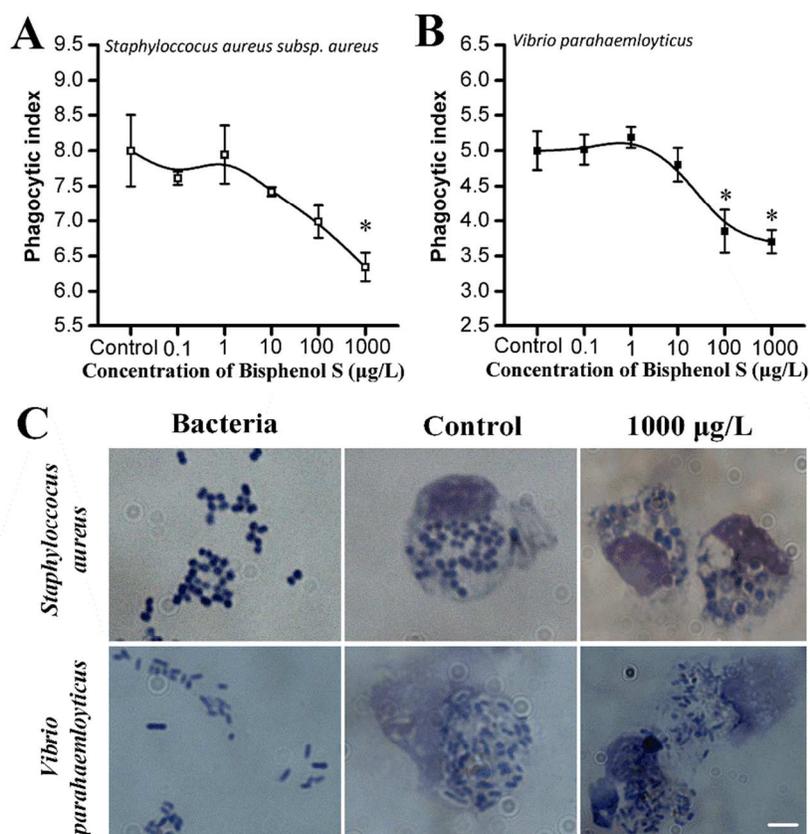


Figure S5. Changes in the phagocytic index of primary macrophages after a 6-h exposure to bisphenol S (BPS). The phagocytic index was measured by counting the intracellular *Staphylococcus aureus* (n = 6, A) and the intracellular *Vibrio parahaemolyticus* (n = 6, B) (C) Representative images comparing the phagocytic activities of the control group and the 1000 µg/L BPS group (scale bar = 5 µm). Values are the means ± standard deviation. An asterisk (*) indicates a significant difference versus the control at $p < 0.05$ (ANOVA, Tukey's test).

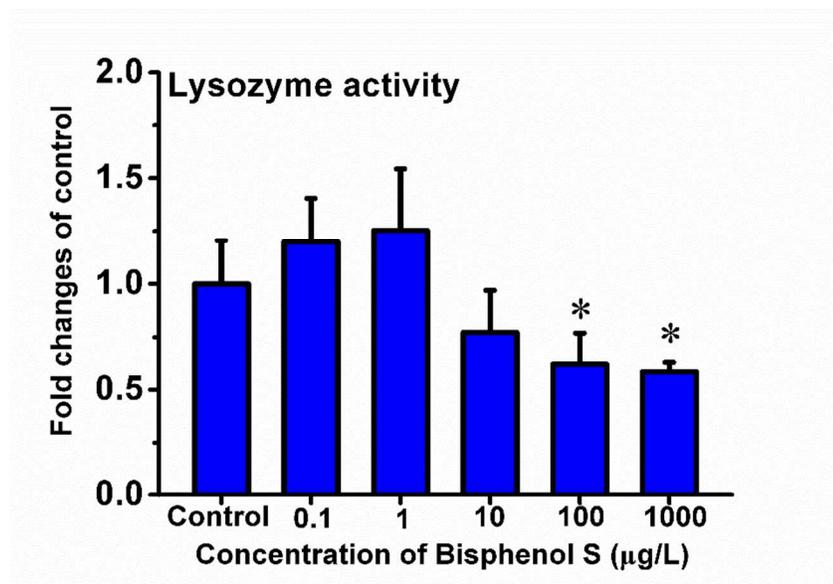


Figure S6. Changes in lysozyme activity of primary macrophages after a 6-h exposure to bisphenol S (BPS). Values are the means \pm standard deviation ($n = 3$). An asterisk (*) indicates a significant difference versus the control at $*p < 0.05$ (ANOVA, Tukey's test).

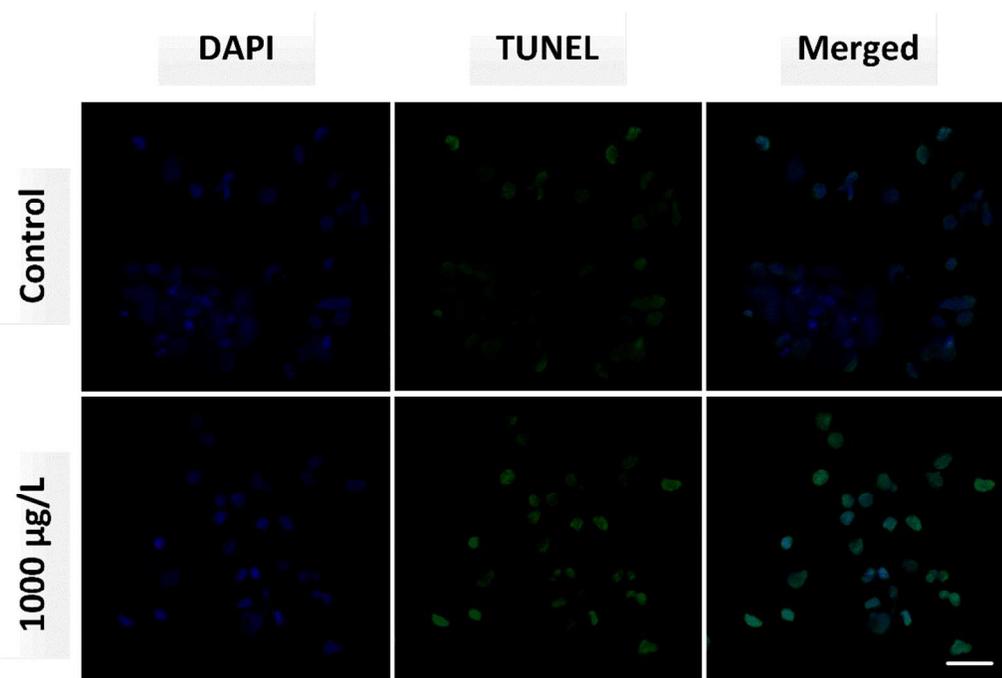


Figure S7. Representative fluorescent images showing a significant increase in TUNEL-positive nuclei (green) in 1,000 µg/L bisphenol S (BPS)-treated samples are compared with the control (Scale bar = 20 µm).

SI Tables

Table S1. Primers used for real-time PCR

Gene name	GenBank accession number	Sequence of the primer (5'-3')
<i>40S ribosomal protein S11</i>	AB012087.1	F: GTTCTCGCTGTTGAAGGAAGTGG R: TTGCGGATGTAATGCAAGTAGTC
<i>Interleukin-1β</i>	AJ245635.1	F: AAGGAGGCCAGTGGCTCTGT R: CCTGAAGAAGAGGAGGCTGTCA
<i>Tumor necrosis factor-α1</i>	AJ311800.2	F: CACGCTCAACAAGTCTCAGAACAA R: CGAAGTAAATGCCGTACAGTAGGA
<i>Interleukin-10</i>	AB110780.1	F: AGCGGGATATGCGGAAATGTAGG R: TGCCAAATACTGCTCGATGTACTTAA
<i>CXCL8</i>	AB470924.1	F: TCACTTCACTGGTGTGCTC R: GGAATTGCTGGCTCTGAATG
<i>γ-Interferon-1</i>	AM261214.1	F: TGCACTTGTCTAGTCTCTGCT R: TGTACTTGTCCCTCAGTATTT
<i>Interleukin-12 p35</i>	AJ580354.1	F: TGCTTCTCTGTCTCTGTGATGGA R: CACAGCTGCAGTCGTTCTTGA
<i>Interleukin-6</i>	KC858889.1	F: GCGTATGAAGGAGCGAAGA R: ATCTGACCGATAGAGGAGCG
<i>Interleukin-11</i>	AJ632159.1	F: GCTGTCACGTCATGAACGAGAT R: CCCGCTTGAGATCCTGAAATAT
<i>CXC chemokine</i>	AB082985.1	F: GTGTGAACATGGTTCCTCCA R: GGATTGAAGCATTCTGCTCT
<i>Interleukin-12 p40</i>	AJ621425.1	F: GAGCGCATCAACCTGACCAT R: AGGATCGTGGATAGTGACCTCTAC
<i>γ-Interferon-2</i>	AM168523.1	F: GAGGAACCTGAGCAGAATCT R: CCTTGATCGCCCATAGTGTT
<i>Tumor necrosis factor-α2</i>	AJ311801.2	F: AGAACAATCAGGAAGGTGGAAATG R: CTGCCGTAGGACTCAGAGTAGCG
<i>Interleukin-6 subfamily-like cytokine M17</i>	AY102632.1	F: AGCTAAATTCAGAATGATCCTCGCTAT R: GCAGAAACTCCTCAGGTGGGTG
<i>α-Interferon</i>	AB376667.1	F: TGCATATGGCTCGGCCAATA R: GTCAAGACAAGAAACCTCACC
<i>Estrogen receptor α</i>	AB334722.1	F: CACAGCCGCCATACACCGAGAT R: GGAAGCCTGGTACTTTCTTAGCC
<i>Estrogen receptor β1</i>	AB334723.1	F: GCCGTGCTCCTCTTTGTTGGTAGC R: ACCTCGGCCATGACTTCACCACTC
<i>Estrogen receptor β2</i>	AB334724.1	F: TTCCCTTCAGGGGACAGAGCTGAG R: AGTCCAGCAGCCTCAGAACCTTCC

Table S2. The lethal concentrations of bisphenol S against fish primary macrophages

Exposure time (h)	LC ₅₀ ^a (mg/L)	LC ₅ ^b (mg/L)
6	39.1 (34.2–44.9)	1.52 (1.21–1.99)
12	29.7 (25.4–35.7)	0.25 (0.07–0.57)
24	16.8 (13.3–21.5)	0.12 (0.04–0.26)

a. Lethal concentration for 50% of the tested populations calculated using the trimmed Spearman–Kärber (TS–K) method. The values in parentheses represent 95% confidence limits.

b. Lethal concentration for 5% of the tested populations calculated using the Probit analysis method. The values in parentheses represent 95% confidence limits.

Table S3. Correlation coefficients between all evaluated parameters in primary macrophages after a 6-h exposure to bisphenol S

	<i>ROS</i>	<i>TAOC</i>	<i>OH ·</i>	<i>MDA</i>	<i>NO</i>	<i>INOS</i>	<i>TNOS</i>	<i>IL1β</i>	<i>TNFA1</i>	<i>IL10</i>	<i>CXC18</i>	<i>INFγ1</i>	<i>IL-12 p35</i>	<i>IL-6</i>	<i>IL11</i>	<i>BA (S)</i>	<i>BA (E)</i>	<i>BA (V)</i>	<i>LYSO</i>	<i>Caspase 3</i>	<i>Tunel</i>	<i>ERα</i>	<i>ERβ1</i>	
<i>TAOC</i>	0.564**																							
<i>OH ·</i>	0.511**	0.794**																						
<i>MDA</i>	-0.206	-0.0353	0.150																					
<i>NO</i>	0.092	0.132	0.111	0.122																				
<i>INOS</i>	0.240	0.281	0.554**	0.364*	0.473**																			
<i>TNOS</i>	0.072	0.212	0.436**	0.130	0.299	0.804**																		
<i>IL1β</i>	0.662**	0.762**	0.749**	-0.23	0.318	0.450**	0.265																	
<i>TNFA1</i>	0.530**	0.623**	0.558**	-0.341*	0.108	0.234	0.157	0.746**																
<i>IL10</i>	0.263	0.250	0.273	-0.020	-0.059	0.164	0.074	0.373*	0.484**															
<i>CXC18</i>	0.620**	0.460**	0.352*	-0.512**	0.118	0.123	0.063	0.637**	0.575**	0.362*														
<i>INFγ1</i>	0.519**	0.608**	0.603**	-0.069	0.346*	0.306	0.088	0.699**	0.542**	0.136	0.452**													
<i>IL-12 p35</i>	0.459**	0.512**	0.607**	-0.272	0.102	0.311	0.268	0.652**	0.496**	0.186	0.508**	0.539**												
<i>IL-6</i>	0.511**	0.686**	0.768**	-0.134	0.105	0.390*	0.263	0.823**	0.700**	0.394*	0.452**	0.590**	0.825**											
<i>IL11</i>	0.391*	0.503**	0.553**	-0.248	-0.105	0.202	0.274	0.543**	0.716**	0.485**	0.489**	0.355*	0.344*	0.518**										
<i>BA (S)</i>	-0.218	-0.054	-0.222	0.168	-0.044	-0.152	-0.091	-0.309	-0.333*	-0.235	-0.255	-0.186	-0.292	-0.230	-0.236									
<i>BA (E)</i>	-0.362*	-0.318	-0.332*	0.118	0.104	-0.137	-0.164	-0.321	-0.402*	-0.046	-0.163	-0.260	0.041	-0.074	-0.475**	0.545**								
<i>BA (V)</i>	-0.191	-0.090	-0.319	0.116	-0.050	-0.193	-0.135	-0.320	-0.308	-0.204	-0.194	-0.195	-0.334*	-0.283	-0.245	0.978**	0.534**							
<i>LYSO</i>	-0.409*	-0.599**	-0.639**	0.168	-0.029	-0.207	-0.185	-0.587**	-0.487**	-0.179	-0.363*	-0.582**	-0.506**	-0.595**	-0.353*	0.109	0.180	0.190						
<i>Caspase 3</i>	0.830**	0.522**	0.479**	-0.433**	0.013	0.138	0.059	0.681**	0.502**	0.287	0.682**	0.400*	0.540**	0.555**	0.463**	-0.114	-0.229	-0.118	-0.444**					
<i>Tunel</i>	0.388*	0.625**	0.559**	-0.280	0.101	0.261	0.227	0.700**	0.720**	0.547**	0.630**	0.338*	0.668**	0.772**	0.563**	-0.086	0.112	-0.080	-0.426**	0.508**				
<i>ERα</i>	0.718**	0.563**	0.559**	-0.297	0.038	0.204	0.091	0.687**	0.568**	0.384*	0.634**	0.563**	0.531**	0.652**	0.513**	-0.221	-0.215	-0.214	-0.449**	0.710**	0.552**			
<i>ERβ1</i>	0.068	0.210	0.245	0.433**	0.267	0.364*	0.160	0.177	-0.089	0.134	-0.072	0.206	0.108	0.147	-0.152	0.009	0.233	0.009	-0.144	-0.011	0.074	0.016		
<i>ERβ2</i>	0.599**	0.470**	0.454**	-0.305	0.264	0.317	0.082	0.662**	0.552**	0.404*	0.606**	0.578**	0.445**	0.495**	0.414*	-0.327	-0.174	-0.284	-0.314	0.565**	0.527**	0.727**	0.246	

* $p < 0.05$ and ** $p < 0.01$ based on Spearman's test.

ROS = reactive oxygen species; TAOC = total antioxidant capacity; OH = hydroxyl radical formation; MDA = lipid peroxidation level; NO = nitric oxide; iNOS = induced nitric oxide synthase; TNOS = total nitric oxide synthase; *IL-1 β* = interleukin-1 β ; *TNF α 1* = tumor necrosis factor α 1; *IL-10* = interleukin-10; *CXCL8* = Cxc chemokine-8; *INF γ 1* = γ -interferon-1; *IL-12 p35* = interleukin-12 p35; *IL-6* = interleukin-6; *IL-10* = interleukin-11; BA (S.) = bactericidal activity against *Staphylococcus aureus* subsp. *aureus*; BA (E.) = bactericidal activity against *Escherichia coli*; BA (V.) = bactericidal activity against *Vibrio parahaemolyticus*; LYSO = lysozyme; Caspase 3 = caspase 3 activity; *ER α* = estrogen receptor α ; *ER β* = estrogen receptor β

