## Supporting information

# Impact of solid state form on the disproportionation of miconazole mesylate salt 

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## Methods

## Single crystal X-ray diffraction

Single crystal X-ray measurements were conducted on a Rigaku Rapid II curved image plate diffractometer with a $\mathrm{Cu}-\mathrm{K} \alpha$ X-ray microsource $(\lambda=1.54178 \AA)$ with a laterally graded multilayer (Goebel) mirror for monochromatization. Single crystals were mounted on Mitegen microloop mounts using a trace of mineral oil and cooled in-situ to $100(2) \mathrm{K}$ for data collection. Data were collected using the dtrek option of CrystalClear-SM Expert 2.1 b32 [1]. Data were processed using HKL3000 [2] and data were corrected for absorption and scaled using Scalepack [2]. The space group was assigned and the structures were solved by direct methods using XPREP within the SHELXTL suite of programs [3] and refined by full matrix least squares against $F^{2}$ with all reflections using Shelx12014 [4] with the graphical interface Shelxle [5]. The
crystal under investigation was found to be non-merohedrically twinned. HKL3000 lacks the ability to simultaneously integrate more than one twin domain. Rigaku programs compatible with the diffractometer (twinsolve) gave unsatisfactory results. The data were thus instead handled as if not twinned, with only the major domain integrated, and converted into an hklf 5 type format hkl file after integration using the "Make HKLF5 File" routine as implemented in the program WinGX [6]. The twin law and matrix were obtained using the program ROTAX as implemented in WinGX. The twin operation was identified as a $180^{\circ}$ rotation around the reciprocal a-axis, the twin matrix as $101.313,0-10,00-1$. The Overlap R1 and R2 values in the "Make HKLF5 File" routine used were 0.23 , i.e. reflections with a discriminator function less or equal to overlap radius of 0.23 were counted overlapped, all others as single. The discriminator function used was the "delta function on index non-integrality". No reflections were omitted.

The structure was solved using direct methods with all reflections of component 1 . The structure was refined using the hklf 5 routine with all reflections of component 1 (including the overlapping ones) as obtained from WinGX, resulting in a BASF value of 0.297(3). No $\mathrm{R}_{\text {int }}$ value is obtainable for the hklf 5 type file using the WinGX routine.

H atoms attached to carbon and nitrogen atoms were positioned geometrically and constrained to ride on their parent atoms, with carbon hydrogen bond distances of $0.95 \AA$ for aromatic and alkene H atoms, $0.99 \AA$ for $\mathrm{CH}_{2}, 0.88 \AA$ for NH , and $0.98 \AA$ for $\mathrm{CH}_{3}$ moieties, respectively. Methyl H atoms were allowed to rotate but not to tip to best fit the experimental electron density. Water H atoms were freely refined with O-H bond distances restrained to $0.84(2) \AA . \mathrm{U}_{\text {iso }}(\mathrm{H})$ values were set to a multiple of $\mathrm{U}_{\mathrm{eq}}(\mathrm{C} / \mathrm{N} / \mathrm{O})$ with 1.5 for OH and $\mathrm{CH}_{3}$, and 1.2 for $\mathrm{N}-\mathrm{H}, \mathrm{C}-\mathrm{H}$ and $\mathrm{CH}_{2}$ units, respectively.

Table 1. Experimental details

|  | MICONZ04 (MM dihydrate) |
| :---: | :---: |
| Crystal data |  |
| Chemical formula | $\mathrm{C}_{18} \mathrm{H}_{15} \mathrm{Cl}_{4} \mathrm{~N}_{2} \mathrm{O} \cdot \mathrm{CH}_{3} \mathrm{O}_{3} \mathrm{~S} \cdot 2\left(\mathrm{H}_{2} \mathrm{O}\right)$ |
| Moiety formula | $\mathrm{C}_{18} \mathrm{H}_{15} \mathrm{Cl}_{4} \mathrm{~N}_{2} \mathrm{O}, \mathrm{CH}_{3} \mathrm{O}_{3} \mathrm{~S}, 2 \mathrm{H}_{2} \mathrm{O}$ |
| $M_{\text {r }}$ | 548.24 |
| Crystal system, space group | Monoclinic, C2/c |
| Temperature (K) | 100 |
| $a, b, c(\AA)$ | 38.449 (3), 9.0878 (5), 13.6903 (9) |
| $\beta$ ( ${ }^{\circ}$ ) | 103.528 (5) |
| $V\left(\AA^{3}\right)$ | 4650.9 (5) |
| Z | 8 |
| $F(000)$ | 2256 |
| $D_{x}\left(\mathrm{Mg} \mathrm{m}^{-3}\right)$ | 1.566 |
| Radiation type | Cu K $\alpha$ |
| No. of reflections for cell measurement | 39176 |
| $\theta$ range ( ${ }^{\circ}$ ) for cell measurement | 4.7-73.5 |
| $\mu\left(\mathrm{mm}^{-1}\right)$ | 5.82 |
| Crystal shape | Plate |
| Colour | Colourless |
| Crystal size (mm) | $0.12 \times 0.06 \times 0.02$ |
|  |  |
| Data collection |  |
| Diffractometer | Rigaku Rapid II curved image plate diffractometer |
| Radiation source | microfocus X-ray tube |
| Monochromator | Laterally graded multilayer (Goebel) mirror |
| Scan method | $\omega$ scans |
| Absorption correction | Multi-scan, SCALEPACK (Otwinowski \& Minor, 1997) |
| $T_{\text {min }}, T_{\text {max }}$ | 0.465, 0.893 |
| No. of measured, independent and observed $[I>2 \sigma(I)]$ reflections | 39176, 39176, 33902 |
| $R_{\text {int }}$ | n/a |
| $\theta$ values ( ${ }^{\circ}$ ) | $\theta_{\text {max }}=72.4, \theta_{\text {min }}=4.7$ |
| $(\sin \theta / \lambda)_{\text {max }}\left(\AA^{-1}\right)$ | 0.618 |


| Range of $h, k, l$ | $h=-47 \rightarrow 47, k=-11 \rightarrow 11, l=-16 \rightarrow 16$ |  |
| :--- | :--- | :---: |
|  |  |  |
| Refinement | $F^{2}$ |  |
| Refinement on | $0.070,0.205,1.05$ |  |
| $R\left[F^{2}>2 \sigma\left(F^{2}\right)\right], w R\left(F^{2}\right), S$ | 39176 |  |
| No. of reflections | 303 |  |
| No. of parameters | 4 |  |
| No. of restraints | H atoms treated by a mixture of independent and <br> constrained refinement |  |
| H-atom treatment | $w=1 /\left[\sigma^{2}\left(F_{\mathrm{o}}^{2}\right)+(0.0633 P)^{2}+49.6689 P\right]$ <br> where $P=\left(F_{\mathrm{o}}^{2}+2 F_{\mathrm{c}}^{2}\right) / 3$ |  |
| Weighting scheme | 0.001 |  |
| $(\Delta / \sigma)_{\max }$ | $0.85,-0.82$ |  |
| $\Delta \rho_{\max }, \Delta \rho_{\min }\left(\mathrm{e} \AA^{-3}\right)$ |  |  |

## Figures



Figure S1. XRPD pattern of (A) MCZ and (B) AH.


Figure S2. FTIR spectrum of (a) MCZ and (b) AH. The dotted vertical lines at $1509 \mathbf{c m}^{-1}$ and 1036 $\mathbf{c m}^{-1}$ represents unique peaks of MCZ and AH , respectively.


Figure S3. DSC traces of (A) AH and (B) AMO.


Figure S4. XRPD pattern of (a) AMO stored in a desiccated container for 4 weeks at $25^{\circ} \mathrm{C}$, (b) AMO stored in $57 \%$ RH for 2 days at $25^{\circ} \mathrm{C}$ (c) AH stored in $75 \%$ RH for 2 days at $25{ }^{\circ} \mathrm{C}$, and (d) calculated XRPD pattern from DH single crystal structure.


Figure S5. Polarized light microscopic images of (A) AMO, (B) AH, and (C) DH.


Figure S6. Particle size distribution for (A) AMO, (B) AH, and (C) DH.


Figure S7. XRPD patterns for various sample after storage for 2 days at (A) at $57 \% \mathbf{R H}$, (B) at 43 \% RH, and (C) at 33 \% RH for (a) AMO + TSPd, (b) AMO + CCS, (c) AH + TSPd, (d) AH + CCS, (e) DH + TSPd, and (f) DH + CCS; (D) control samples of (a) neat AMO at $57 \%$ RH, (b) neat AH at $57 \%$ RH, (c) neat AMO at $43 \% \mathrm{RH}$, (d) neat AH at $43 \% \mathrm{RH}$, (e), neat AMO at $33 \% \mathrm{RH}$, and (f) neat AH at 33 \% RH. (E) Samples for TSPd stability analysis (a) neat TSPd at $33 \% \mathbf{R H}$, (b) neat TSPd at $43 \%$ RH, (c) neat TSPd at $57 \%$ RH, and (d) neat TSPd before storage. The dotted and dashed lines indicate the position of unique peak for MCZ and DH, respectively.


Figure S8. XRPD pattern of (a) neat DH, (b) neat DH after exposure to $57 \%$ RH for 7 days, (C) neat MCZ, and (d) neat MCZ after exposure to $57 \%$ RH for 7 days.


Figure S9. Raman spectra of (a) neat DH, (b) neat DH after exposure to 57 \% RH for 7 days, (C) neat MCZ, (d) neat MCZ after exposure to $57 \%$ RH for 7 days, (e) $\mathbf{D H}+\mathrm{MCZ}$ (1:1) mixture, and (f) DH + MCZ (1:1) mixture after exposure to $57 \%$ RH for 7 days. The composition of the mixture of the DH ( 50 mole \%) and MCZ ( 50 mole \%) did not change upon storage at 57 \% RH for 7 days.


Figure S10. (A) Raman and (B) XRPD spectra of (a) MM solid form used in this study and (b) MM solid formed obtained by following method described in Guerrieri et. al (2009).


Figure S11. DSC traces of AMO stored at $33 \% \mathrm{RH}, 43 \% \mathrm{RH}$, and $57 \% \mathrm{RH}$.


Figure S12. XRPD pattern of solid residues obtained after slurrying DH in water for $\mathbf{2}$ days.

## References

[1] Rigaku Corp., The Woodlands, Texas, USA.
[2] Otwinowski Z, Minor W., Processing of X-ray diffraction data collected in oscillation mode. Methods Enzymol. 1997, 276, 307-326.
[3] a) SHELXTL suite of programs, Version 6.14, 2000-2003, Bruker Advanced X-ray Solutions, Bruker AXS Inc., Madison, Wisconsin: USA) b) Sheldrick GM. A short history of SHELX. Acta Crystallogr A. 2008, 64(1), 112-122.
[4] a) Sheldrick GM. University of Göttingen, Germany, 2013. b) Sheldrick GM. Crystal structure refinement with SHELXL. Acta Crystallogr Sect C Struct Chem. 2015, 71(1), 3-8.
[5] Hübschle CB, Sheldrick GM, Dittrich B. ShelXle: a Qt graphical user interface for SHELXL. J. Appl. Crystallogr. 2011, 44(6), 1281-1284.
[6] Farrugia, LJ, J. Appl. Crystallogr. 2012, 45, 849-854.

